RECENT ADVANCEMENTS IN THETOXICOLOGICAL ASSESSMENT **ANDBACTERIAL - MEDIATED BIODEGRADATION** PATHWAYS OF PARACETAMOL

Abstract

Paracetamol or acetaminophen (N- Asmita Gupta acetyl-para-aminophenol) $(C_8H_9NO_2)$ is the most widely used over the Daulat Ram College counter (OTC) non-opioid analgesic and University of Delhi antipyretic agent used to treat mild to New Delhi, India. moderate pain and fever. Being an OTC asmita@dr.du.ac.in drug, it is freely available, resulting in its rampant overuse, especially during Covid Swati and in the post Covid period. Ingestion of Department of Biosciences Paracetamol at doses above recommended levels has been found to New Delhi, India. causehistopathological and neurotoxic effects. Thus, this global issue of improper Madan Kumar environmental and use paracetamol and the associated risks raise Technology (CRDT) concerns and demand for a systematic Indian Institute of Technology (IIT) approach towards its monitoring and New Delhi, India. remediation. Bioremediation of paracetamol from environmental samples using bacteria Rama Pasricha such as *Pseudomonas* sp., *Bacillus* sp., Department of Botany Enterococcus sp. has been shown to be Daulat Ram College aneffective, environment-friendly and low- University of Delhi cost method. The present chapter highlights New Delhi, India. the recent advancements in the toxicity studies on unnatural exposure to paracetamol and its bacterial biodegradation pathways.

Keywords: Acetaminophen, Wastewater, Biodegradation, Bacteria, Toxicity.

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I. INTRODUCTION

Paracetamol or Acetaminophen is an antipyretic, non-opioid analgesic drug that is used to treat fever and moderate pain. It is the most commonly used OTC analgesic drug worldwide and is also recommended by the World Health Organization (WHO) as a first-line treatment drug for pain conditions. Initially approved by the U.S. FDA, 1951 (Swann 1994), paracetamol is currently available in various forms such as regular and effervescent tablets, syrup, and injection, and is also used in combination with other drugs in many other medications for allergy, cold, pain, sedative, among others (Jóźwiak-Bebenista and Nowak 2014).

Among all the pharmaceutical compounds contaminating the surface and wastewaters worldwide, acetaminophen prevalence and concentrations have been found to be among the highest (more than 100 ng/L in rivers and 100 μ g/L in sewage treatment plant influents) Rios-Miguel et al. (2022). The exposure with paracetamol at undesirable concentration shows various pathological conditions. Mossa et al. (2012) showed that liver of rats has various biochemical and histopathological changes when exposed to the paracetamol at concentration 66mg/kg. Blecharz Klin et al. (2015a) has studied the neurotoxicity associated with the exposure of acetaminophen at 5 and 15mg/kg concentration and found that it causes defect in the neurotransmission process associated with medulla oblongata in early developmental stages.

Direct effects on spinal cord were also observed by Blecharz Klin et al. (2015b). Hepatotoxicity has also been observed in alcoholic people and it can be fatal at over dosage (Artnak and Wilkinson 1998; Bridger et al. 1998; Denison et al. 1987; Himmelstein et al. 1984; Johnson et al. 1981; Yasunaga et al. 1985). Although the effect of paracetamol on pregnant ladies and there unborns is still unclear but few studies interlinked the paracetamol exposure in offsprings with ADHD (attention deficit hyperactivity disorder) and ASD (autism spectrum disorder) later in life (Endesfelder et al. 2021). The exposure with paracetamol whether therapeutic or environmental, causes ill effects in the human beings and other organisms.

Recent knowledge about the accumulation of medicinal products in the environment matrices and its possible effect on aquatic speciesshows the necessity of conducting environmental risk assessment (ERA)studyof medicinal product before marketing it for human use as per the European Council Directive 2001/83/EC, 2001 (Valverde 2006).The environmental exposure with any pharmaceutical compound can be calculated as the ratio of the predicted environmental concentration (PEC) and measured environmental concentration (MEC) to the predicted no-effect concentrations (PNEC) of the pharmaceuticals (Ashfaq et al.2017a).

Thus, the easy access and widespread use of paracetamol and the increased use of its combination medications have become one of the main causes of the intentional or non-intentional overdose, resulting in the commonly observed drug induced liver injury (DILI) (Rotundo and Pyrsopoulos 2020). Also, the discharge of excess drug into the environment through patients' urine and hospital wastewater has caused environmental pollution and toxicity. Several physical and chemical processes have been used for removal of Paracetamol from environmental samples. Nonetheless, the degradation of paracetamol by microbes is

considered as an environmentally friendly and cost-effective process, and results in the complete mineralization of the contaminant. The biodegradation of paracetamol in the environment is reported for bacteria, fungi, algae. Bacterial degradation of paracetamol has been shown to give promising results by several researchers. In the light of the above, the present chapter focuses on the recent advances in the toxicology of paracetamol and its bacterial mediated degradation pathway analysis.

II. CHARACTERISTICS OF PARACETAMOL

Paracetamol or para-hydroxyacetanilide is a derivative of phenol that is chemically named N-acetyl-para-aminophenol, in which one of the hydrogen atoms of amino group is replaced by an acetyl group. Known as Acetaminophen in the Americas and Paracetamol in the rest of the world, it is sold under different brands like Crocin, Tylenol, Excedrin, Calpol, Panadol, etc). Chemically, paracetamol or 4-hydroxyacetanilide is a white, odorless crystalline solid (Di Martino et al. (1997). Some of the important physical properties of paracetamol are described in Table 1. The 2D and 3D structures of paracetamol has been depicted in Figure 1.

Parameter	Physical Property	Reference	
Appearance	white crystalline powder	Di Martino et al., (1997)	
Colour/Form	White colourless crystalline powder	IARC (1990)	
Molecular	151.16 g/mol	IARC (1990)	
Weight			
Molecular	C ₈ H ₉ NO ₂ or HOC ₆ H ₄ NHCOCH ₃	IARC (1990)	
formula			
Taste	Slightly Bitter	Lewis (2007)	
Odour	Odorless	Lewis (2007)	
pН	about 6 (saturated aqueous solution)	NTP (1992)	
Melting point	169-170.5 °C	Di Martino et al., (1997)	
Boiling point	>500 °C	ILO (2008)	
Solubility	Soluble in water (1:70, 1:20 at 100°C)	NTP (1992)	
Density	1.293 g/cu cm at 21 °C	Haynes, (2014)	
Environmental	Toxic to aquatic life.	ILO (2008)	
risk			

 Table 1: Physical Properties of Paracetamol

In the 1980s, acetanilide was found to be effective in treating fever, but due to its high toxic effects, it was later replaced by one of its safer chemical derivatives, that is, *N*-acetyl*para*-aminophenol, or paracetamol. The mechanism of action of paracetamol is complex and it has an effect on the central and peripheral nervous system. It is a selective cyclooxygenase (COX-1 and COX-2) inhibitor, suppresses prostaglandin production and is involved in the endocannabinoid system and serotonergic pathways, thus resulting in analgesic and antipyretic effects (Ayoub 2021; Przybyła et al. 2021). However, paracetamol is known for its hepatotoxic effects at doses above the recommended levels. A dose of 4000 mg within 24 h has been recommended by the US FDA, and for the European countries, a maximum dose of a 3000 mg of paracetamol in older adults has been recommended (Rotundo and Pyrsopoulos 2020).



Figure 1: Structure of Paracetamol (a) 2D structure, (b) 3D structure

III. TOXICITY ASSESSMENT OF PARACETAMOL

During COVID -19 prevalence as a pandemic the usage of paracetamol for the treatment of fever and pain increased a lot. Due to its excessive usage, the degradation products of paracetamol alongwith the unused parent compound has accumulated in environmental matrices specially the water bodies. In the last decade, the increased concentration of pharmaceuticals in wastewater has been observed and paracetamol is one of them.Paracetamol has multidirectional mechanism of action (Figure 2) which is different from other drugs belonging to this group of non-steroidal anti-inflammatory drug (NSAID). It causes inhibition of cyclooxygenase (COX), a constitutive enzyme that is involved in the synthesis of prostaglandin-an inflammatory pain mediator. Prostaglandin is further transformed into other active compounds having different effects on human body(Ghanem et al. 2016).

The exposure risk associated with the paracetamol in the environmental matrices has been studied by several researchers. They have done toxicity testing both *in vitro* and *in vivo*. It has been observed that toxicity mechanism of paracetamol is highly conserved and it is same in both vertebrates and invertebrates. The neurotoxicity which is an important parameter in ecotoxicity testing has been widely studied by Oliveira et al. (2015). It has been observed that acetylcholinesterase (AChE) and Se-GPx activity is inhibited in *Daphnia magna* when exposed to 10ug/L of APAP (Oliveira et al.2015). Seawater mussel *Mytilus galloprovincialis* also showed similar effect when exposed to paracetamol at concentration ranging from 0.023 to 0.403 mg/L (Solé et al. 2010). Nunes et al. (2015) and Ramos et al. (2014) postulated that the inactivation of enzymes in the presence of paracetamol is due to the oxidative stress that is the most common effect of paracetamol exposure.*Torpedo californica* (fish) (Weiner et al. 1994), Rodents (Delwing-de Lima et al.2010), *Brachionus koreanus* (rotifer) (Rhee et al.2013), and *Anguilla anguilla* (European eel) (Nunes et al.2015) have shown similar effect because of the oxidative stress from paracetamol exposure.





The oxidative changes in the sample can be assessed by measuring changes in the activity of glutathione peroxidase and catalase which are the most common biomarkers of oxidative stress.Different authors have reported contradictory results on the activity of oxidative stress biomarkers. Studies done by Oliveira et al. (2015) and Monteiro et al. (2006) have reported the decrease in activity of biomarkers related to oxidative stress whereas studies on *Orcheochromis mossambicus* (Kavitha et al. 2011), *Cyprinus carpio* (Nava-Álvarez et al. 2014), or *Dreissena polymorpha* (Parolini and Binelli 2012) showed opposite results i.e. increase in oxidative stress biomarkers were observed when exposed to paracetamol. The various studies on paracetamol toxicity have been summarized in Table 2.

Table 2: Toxicity assessment of paracetamol on different orga

Organism	Assays	Exposure	Tested	Toxicity	Reference
		time	conc.		
Cyprinus	Biomarkers tests	96 h	100 µg/L	 Oxidative 	Nava-
carpio	were done: lipid			stress	Álvarez
	peroxidation,			 Increase in 	et al.
	superoxide dismutase			GPx activity	(2014)
	(SOD), catalase				
	(CAT), and total and				
	selenium-dependent				
	glutathione-				
	peroxidase (GPx),				

Rhamdia	Piscine micronucleus	21 days	0.25, 2.5	• Oxidative	Guiloski
quelen	test, of steroid		µg/L	stress	et al.
	hormone dosage and			 Neuronal 	(2017)
	comet assay assessed			disruption	
				 Reduction 	
				inhemoglobin	
				• Decrease in	
				leukocytes and	
				thrombocytes	
				• Increase in	
				estradiol	
				levels	
				hepatotoxicit	
				У	
Diopatra	Tissue regenerative	30 days	5 25	• Diminished	Fraitas at
neapolitana	capacity as a	39 uays	125		(2015)
neupoinana	biomarker		625.	capacity	ul. (2015)
			3125	cupucity	
			µg/L		
Daphnia	Total cholinesterases,	48 h	0.00001,	• Inhibition of	Oliveira
magna	CAT, glutathione-S-		0.001,	enzymes like	et al.
	transferases (GSTs),		0.25	AChE and	(2015)
	GPx		mg/L	GPx	
Matilus	Different	10 dave	20, 200	• Hanatia linid	Soló at al
alloprovinc	biochemical assays	10 uays	20, 200 ug/I	• Hepatic lipid	(2010)
ialis	were done like		μg/L	levels	(2010)
ians	acetylcholinesterase			increased	
	(AChE), GSTs, CAT.			• inhibition of	
	Lipid peroxidation			AChE	
	levels (LPL)			activity	
				5	
Anguilla	Battery of	96h	5, 25,	• Inhibition of	Nunes et
anguilla	antioxidant		125,	AChE	al. (2015)
	biomarkers: CAT,		625,	activity	
	AChE and GSTs,		3125		
	thiobarbituric acid		µg/L		
	(TDADS) constances				
	(IBAKS) assay,				
	dehydrogenase				
	(LDH)				
Brachionus	AChE activity	24h	0.1.1.	• Inhibition of	Rhee et
koreanus	assessed		10, 100.	AChE	al. (2013)
-			1000,	activity	· - /
			10,000,	5	

			100,000 μg/L		
Oncorhynch us mykiss	Biochemical biomarkers: CAT, GPx, glutathione reductase (GRed) and GSTs activity and also LPL, TBARS assessed	96 h (acute exposure), 28 days (chronic exposure)	0.05, 0.5, 5 mg/L (acute exposur e), 12.5, 25, 50 µg/L (chronic exposur e)	 Increase in catalase activity Increase in GPx activity 	Ramos et al. (2014)
Tisbebatta gliai.	Evaluation was done by calculating risk quotients (RQs) based on MEC (the highest exposure concentration of the compound in the medium)/PNEC (predicted no effect concentration) ratios	48 h	1.7, 2.2 ,3.4, 4.5 ,6.7, 8.4, 11.2, 16.8, 17.0, 22.6, 25.5, 34.0, 33.9, 51.0 mg/L (in mixture with other pharmac euticals)	• At environmenta l conc., low or no risk.	Trombini et al. (2016)
<i>Hediste</i> <i>diversicolor</i>	Biomarkers such as CAT, GSTs, AChE, and cyclooxygenase (COX) activities, LPL, TBARS	28 days	0, 30, 60, and 120 µg/L	 No changes are observed in COX activity alteration are observed in CAT activity decrease in GSTs Activity 	Daniel et al. 2022

The other effects related to paracetamol exposure as observed in male fish *Rhamdia quelen* are severe changes of hematological parameters, i.e., reduction of hemoglobin and hematocrit, mild blood congestion and leukocyte infiltration, and disruption of hypothalamic-pituitary-gonadal axis (Guiloski et al. 2017). The regenerative tissue capacity of a polychaete

Diopatraneapolitana has found to be decreased when exposed to environmentally relevant concentration of paracetamol ($25 \mu g/L$) (Freitas et al. 2015). Different plant species such as *L. culinaris* Med. and *P. sativum* L have shown cytotoxic effect in the roots when exposed to paracetamol. The paracetamol exerted effect on these plants species at each concentration tested. It hampered the mitotic processes, inhibits root growth and showed the presence of micronucleus (Mercado and Galvis 2023).

Other than *in vivo* and *in vitro* toxicological studies, the ratio between predicted or measured environmental concentration to the predicted or measured no effect concentration of the compound is also commonly used in ecological risk assessment of pharmaceutical compounds (Ashfaq et al. 2017a, b) which is generally termed as risk quotient (RQ). It gives a direct insight on the harmful dosages of various pollutants. The value of RQhigher than 1 indicates high risk to the Aquatic community as shown by Bouissou-Schurtz et al. (2014). It has been observed that the values of RQ for paracetamol as calculated in *Streptocephalusproboscideus*, *Daphnia*, green algae and *D. magna* were, 9.2, 64, 0.11 and 5.0 respectively. The values clearly indicate that except green algae all the other three under study are in high risk to paracetamol exposure.

IV. MECHANISM OF PARACETAMOL DEGRADATION BY BACTERIA

The microorganisms such as Bacteria, Algae and Fungi utilize paracetamol as energy and carbon source. The degradation depends on environmental factors such as pH, temperature, and cellular structure such as cell structure and physiology of cell membrane. At high and low temperature, the porosity and rigidity of cell membrane hinders the efficient uptake of molecules or enzyme activity thereby the reaction becomes slow. pH regulated the toxicity and uptake of molecules through the cell membrane. At acidic pH, the paracetamol is present in protonated form whereas at alkaline pH the phenolate form occurs.

The degradation of paracetamol occurs at optimum temperature and pH (Żur et al. 2018). The degradation and mineralization of paracetamol is done with the help of a diverse array of enzymes and pathways. The degradation of paracetamol is reported under aerobic as well as anaerobic environments (Figure 3 and 4).

There are several pathways reported for paracetamol degradation under aerobic conditions, and it occurs mainly through hydroquinone pathway and pyrocatechol pathway (Figure 3). In hydroquinone pathway, the paracetamol is converted to hydroquinone through amidohydroxylation in which the paracetamol is first converted to 4-aminophenol by releasing acetate. Thereafter, amino groups are replaced by hydroxyl group to form hydroquinone. In the case of hydroxylation, the paracetamol is converted to hydroquinone releasing acetamide. The hydroquinone is converted to 4-hydroxymuconic semialdehyde by hydroquinone 1,2-dioxygenase for further metabolism through tricarboxylic cycle. The enzymes involved in amidohydroxylation are amidohydroxylase and amino acid oxidase. The enzymes involved in initial hydroxylation are hydrolytic enzymes.

In another pathway i.e., pyrocatechol pathway, the paracetamol is converted to N-(4-hydroxyphenyl)-acetamide by hydroxylase. The N-(4-hydroxyphenyl)-acetamide can be degraded through the following routes (a) N-(4-hydroxyphenyl)-acetamide oxidation to benzoquinone and hydroquinone, (b) N-(4-hydroxyphenyl)-acetamide conversion to

pyrocatechol by oxygenase and formamide CGTase. These two routes finally generate intermediates (p-phenol) which can undergo ring fission for further degradation through TCA cycle (Wu et al. 2012; Żur et al. 2018).



Figure 3: Pathways of aerobic degradation of Paracetamol degradation by bacteria (Li et al. 2014; Takenaka et al. 2003; Wu et al. 2012; Żur et al. 2018),

A different pathway of paracetamol degradation was observed in *Burkholderia* sp. AK-5, in which the p-aminophenol is transformed to 1,2,4-trihydroxybenzene by two successive hydroxylation generation 1,4-benzenediol as intermediate. Thereafter, 1,2,4-trihydroxybenzene undergo conversion to maleylacetic acid by 1,2,4-trihydroxybenzene 1,2-dioxygenase (Takenaka et al. 2003). Another pathway for paracetamol degradation was proposed while studying its microbial degradation in soil, where the paracetamol is converted to 3-hydroxyacetaminophen. The enzymes involved in the process are cytochrome P-450 hydroxylase. This 3-hydroxyacetaminophen can be further degraded via two routes (a) methylation to p-acetaniside which further gets degraded via intermediates such as 4-methoxy phenol, 1,4, dimethoxybenzene and 2-hexenoic acid, (b) oxygenation to N-acetylp-benzoquinone imine which further degraded via., para-benzoquinone (Li et al. 2014).

When the genetic studies were performed to identify the genes involved in paracetamol degradation, it was observed that plasmid plays an important role in paracetamol degradation. The *Pseudomonas* sp. ST-4 has been shown to degrade 4-aminophenol through plasmid (Khan et al. 2006). The genes responsible for hydroquinone degradation and their regulation have been elucidated in *Pseudomonas sp. tida* DLL-E4 (Hu et al. 2014). There are several reports available where the *Pseudomonas* sp. has been studied for paracetamol degradation in the concentration range 50-4000 mg/L and the pathway have been suggested based on the detection of intermediated or enzymes (Poddar et al. 2022; Zur et al. 2018;

Zhang et al. 2013; Hu et al. 2013). In a similar study, microbes such as *Brevibacterium, Corynebacterium, Enterococcus* and *Bacillus* sp., isolated from activated sludge sample were studied for paracetamol degradation. Cultures of *Brevibacterium, Corynebacterium* and *Enterococcus* resulted in 86-97% degradation of paracetamol at 200mg/L concentration but in case *Bacillus* complete mineralization of paracetamol was observed along with detection of degradation intermediates viz., 4-aminophenol, hydroquinone and 2-hexenoic acid (Palma et al. 2021).

The degradation of paracetamol under anaerobic condition was also attempted by the researchers. Recently a study on paracetamol degradation by *Bacillus drentensis* strain S1 was performed under anaerobic conditions and the pathway was also deciphered based on the detection of degradation intermediates viz., phenothiazine and 2-isopropyl-5-methylcyclohexanone (Figure 4). This strain was reported to degrade 94.5% (initial concentration 300mg/ L) at pilot scale under anaerobic conditions (Chopra and Kumar 2020).



Figure 4: Pathways of anaerobic degradation of Paracetamol degradation by bacteria (Chacón et al. 2022; Chopra and Kumar 2020).

In another study, the degradation of paracetamol was reported in mangrove sediments. A total of 16 microbial genera (*Bacillus, Enterobacter and Arthrobacter* could be identified) were found to be associated with paracetamol degradation. Furthermore, they have studied the role of electron acceptor viz., NaNO₃, Na₂SO₄ and NaHCO₃ addition for enhanced paracetamol degradation under anaerobic conditions and NaNO₃ showed the best degradation in case of mangrove sediments (Yang et al., 2020). Chacón et al (2022) studied the role of raw biochar and designer redox-active biochars in paracetamol degradation. They observed that under anaerobic conditions, the degradation of paracetamol occurs via methylation route as 4-methoxyphenol was predominantly present during the degradation.

V. CONCLUSION

Paracetamol or acetaminophen is one of the most commonly used over-the-counter pharmaceutical drug worldwide. It is the standard first-line treatment for fever and pain. Its use has further increased post-Covid. Paracetamol's easy availability has resulted in excessive use leading to an increase in its concentration in the environment. Acetaminophen levels have been found to be among the highest of all the pharmaceuticals in surface and river waters. Several researchers have tested the risks associated with exposure to paracetamol in the environment in several organisms and have reported hepatotoxic and neurotoxic effects. It has been observed that the mechanism of toxicity of paracetamol is highly conserved and same in both vertebrates and invertebrates. For removal of paracetamol from the environment, biological route through bacterial biodegradation has been shown to be quiet effective and eco-friendly. Various pathways of paracetamol degradation and mineralization like hydroquinone and pyrocatechol pathways among others have been reported in different bacterial strains. Bacterial biodegradation of paracetamol under anaerobic conditions have also been studied. Further research into more effective and widely applicable routes of removal of paracetamol from the environment along with its rational and efficient use is much needed for better management of this most commonly used medicine that is here to stay for many more years to come.

REFERENCES

- [1] Artnak KE, Wilkinson SS (1998) Fulminant hepatic failure in acute acetaminophen overdose. Dimensions Crit Care Nursing 17(3): 135-144. 51.
- [2] Ashfaq M, Khan KN, Saif-Ur-Rehman M, Mustafa G, Nazar MF, Sun Q, Iqbal J, Mulla SJ, Yu C-P (2017a) Ecological risk assessment of pharmaceuticals in the receiving environment of pharmaceutical wastewater in Pakistan. Ecotoxicol Environ Saf 136:31–39. https://doi.org/10.1016/j.ecoenv.2016.10.029
- [3] Ashfaq M, Noor N, Saif-Ur-Rehman M, Sun Q, Mustafa G, Nazar MF, Yu C-P (2017b) Determination of commonly used Pharmaceuticals in hospital waste of Pakistan and evaluation of their ecological risk assessment. Clean Water Air Soil 45. https://doi.org/10.1002/clen. 201500392
- [4] Ayoub SS (2021) Paracetamol (acetaminophen): A familiar drug with an unexplained mechanism of action. Temperature, 8(4), pp.351-371.
- [5] Blecharz Klin K, Joniec Maciejak I, Jawna K, Pyrzanowska J, Piechal A, et al. (2015a) Developmental exposure to paracetamol causes biochemical alterations in medulla oblongata. Environ Toxicol Pharmacol 40(2): 369-374.
- [6] Blecharz Klin K, Joniec Maciejak I, Jawna K, Pyrzanowska J, Piechal A, et al. (2015b) Effect of prenatal and early life paracetamol exposure on the level of neurotransmitters in rats-focus on the spinal cord. Int J Dev Neurosci 47(pt B): 133-139.
- [7] Bouissou-Schurtz C, Houeto P, Guerbet M, Bachelot M, Casellas C, Mauclarie AC, Panetier P, Delval C, Masset D (2014) Ecological risk assessment of the presence of pharmaceutical residues in a French national water survey. Regul Toxicol Pharmacol 69:296–303. https://doi.org/10.1016/j.yrtph.2014.04.006
- [8] Bridger S, Henderson K, Glucksman E, Ellis AJ, Henry JA, et al. (1998) Deaths from low dose paracetamol poisoning. Br Med J 316(7146): 1724-1725.
- [9] Chacón FJ, Cayuela ML, & Sánchez-Monedero MA (2022) Paracetamol degradation pathways in soil after biochar addition. Environmental Pollution, 307, 119546.
- [10] Chopra S, & Kumar D (2020) Characterization, optimization and kinetics study of acetaminophen degradation by Bacillus drentensis strain S1 and waste water degradation analysis. Bioresources and Bioprocessing 7, 1-18.
- [11] Daniel D, Nunes B, Pinto E, Ferreira IM, & Correia AT (2022) Assessment of paracetamol toxic effects under varying seawater pH conditions on the marine polychaeteHedistediversicolor using biochemical endpoints. Biology 11(4), 581.
- [12] Delwing-de Lima D, Wollinger LF, Casagrande ACM, Delwing F, da Cruz JGP, Wyse ATS, Magro DD-D (2010) Guanidino compounds inhibit acetylcholinesterase and butyrylcholinesterase activities: effect

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neuroprotector of vitamins E plus C. Int J Dev Neurosci 28:465–473. https://doi.org/10.1016/j.ijdevneu.2010.06.008

- [13] Denison H, Kaczynski J, Wallerstedt S (1987) Paracetamol medication and alcohol abuse: a dangerous combination for the liver and kidney. Scand J Gastroenterol 22(6): 701-704
- [14] Di Martino P, Conflant P, Drache M, Huvenne J-P, & Guyot-Hermann A-M (1997) Preparation and physical characterization of forms II and III of paracetamol. Journal of Thermal Analysis 48(3), 447– 458. doi:10.1007/bf01979491
- [15] Endesfelder CB, Scheuer T, Schmitz T (2021) Paracetamol (Acetaminophen) and the Developing Brain. International Journal of Molecular Sciences 22(20): 11156.
- [16] Freitas R, Coelho D, Pires A, Soares AMVM, Figueira E, Nunes B (2015) Preliminary evaluation of Diopatraneapolitana regenerative capacity as a biomarker for paracetamol exposure. Environ Sci Pollut Res 22:13382–13392. https://doi.org/10.1007/s11356-015-4589-1
- [17] Ghanem CI, Perez MJ, Manautou JE, Mottino AD (2016) Acetaminophen from liver to brain: New insights into drug pharmacological action and toxicity. Pharmacol Res 109: 119-131.
- [18] Guiloski IC, Ribas JLC, Piancini LDS, Dagostim AC, Cirio SM, Favaro LF, Boschen SL, Cestari MM, da Cunha C, de Assis HC (2017) Paracetamol causes endocrine disruption and hepatotoxicity in male fish Rhamdia quelen after subchronic exposure. Environ Toxicol Pharmacol 57:111–120. https://doi.org/10.1016/j.etap.2017.05.005
- [19] Haynes WM (ed.) (2014) CRC Handbook of Chemistry and Physics. 94th Edition. CRC Press LLC, Boca Raton: FL 2013-2014, p. 3-314
- [20] Himmelstein DU, Woolhandler SJ, Adler RD (1984) Elevated SGOT/SGPT ratio in alcoholic patients with acetaminophen hepatotoxicity. Am J Gastroenterol 79(9): 718-720.
- [21] Hu J, Zhang L L, Chen JM, & Liu Y (2013) Degradation of paracetamol by Pseudomonas aeruginosa strain HJ1012. Journal of Environmental Science and Health Part A, 48(7), 791-799.
- [22] Hu X, Wang J, Wang F, Chen Q, Huang Y, & Cui Z (2014) Complete genome sequence of the pnitrophenol-degrading bacterium Pseudomonas putida DLL-E4. Genome announcements 2(3), 10-1128.
- [23] IARC (1990) IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Pharmaceutical Drugs. Lyon (FR): International Agency for Research on Cancer; 1990. (IARC Monographs on the Evaluation of the Carcinogenic Risks to Humans, No. 50.) Paracetamol (Acetaminophen) Available from: https://www.ncbi.nlm.nih.gov/books/NBK526213/
- [24] ILO (2008) ILO-WHO International Chemical Safety Cards (ICSCs) ICSC: 1330 (November 2008) https://www.inchem.org/documents/icsc/icsc/eics1330.htm
- [25] Irwin RD, Boorman GA, Cunningham ML, Heinloth AN, Malarkey DE and Paules RS (2004) Application of toxicogenomics to toxicology: basic concepts in the analysis of microarray data. Toxicologic pathology 32(1_suppl), pp.72-83.
- [26] Johnson MW, Friedman PA, Mitch WE (1981) Alcoholism, nonprescription drugs and hepatotoxicity: the risk from unknown acetaminophen ingestion. Am J Gastroenterol 76: 530-533.
- [27] Jóźwiak-Bebenista M and Nowak JZ (2014) Paracetamol: mechanism of action, applications and safety concern. Acta poloniaepharmaceutica 71(1), pp.11-23.
- [28] Kavitha P, Ramesh R, Bupesh G, Stalin A, Subramanian P (2011) Hepatoprotective activity of Tribulus terrestris extract against acetaminophen-induced toxicity in a freshwater fish (Oreochromis mossambicus). In Vitro Cell Dev Biol Anim 47:698–706. https:// doi.org/10.1007/s11626-011-9457-9
- [29] Khan SA, Hamayun M, & Ahmed S (2006) Degradation of 4-aminophenol by newly isolated Pseudomonas sp. strain ST-4. Enzyme and Microbial Technology 38(1-2), 10-13.
- [30] Lewis RJ Sr (2007) Hawley's Condensed Chemical Dictionary 15th Edition. John Wiley & Sons, Inc. New York, NY., p. 11
- [31] Li J, Ye Q, & Gan J (2014) Degradation and transformation products of acetaminophen in soil. Water Research 49, 44-52.
- [32] Mercado SAS, Galvis DGV (2023) Paracetamol ecotoxicological bioassay using the bioindicators Lens culinaris Med. and Pisum sativum L. Environ Sci Pollut Res 30, 61965–61976. https://doi.org/10.1007/s11356-023-26475-7
- [33] Monteiro DE, de Almeida JA, Rantin FT, Kalinin AL (2006) Oxidative stress biomarkers in the freshwater characid fish, Brycon cephalus, exposed to organophosphorus insecticide Folisuper 600(methyl parathion). Comp BiochemPhysiol C 143:141–149. https://doi.org/10.1016/j.cbpc.2006.01.004
- [34] Mossa ATH, Heikal TM, Omara EAA (2012) Physiological and histopathological changes in the liver of male rats exposed to paracetamol and diazinon. Asian Pac J Trop Biomed 2(3): S1683-S1690.

- [35] Mullins ME, Yeager LH and Freeman WE (2020) Metabolic and mitochondrial treatments for severe paracetamol poisoning: a systematic review. Clinical Toxicology 58(12), pp.1284-1296.
- [36] Nava-Álvarez R, Raz-Estrada AC, Garcia-Medina S, Gómez-Oliván LM, Galar-Martínez M (2014) Oxidative stress induced by mixture of diclofenac and acetaminophen on common carp (Cyprinus carpio). Water Air Soil Pollut 225:1873. https://doi.org/10.1007/s11270-014-1873-5
- [37] NTP (1992) National Toxicology Program, Institute of Environmental Health Sciences, National Institutes of Health (NTP). 1992. National Toxicology Program Chemical Repository Database. Research Triangle Park, North Carolina.
- [38] Nunes B, Verde MF, Soares AMVM (2015) Biochemical effects of the pharmaceutical drug paracetamol on *Anguilla anguilla*. Environ Sci Pollut Res 22:11574–11584. https://doi.org/10.1007/s11356-015- 4329-
- [39] Oliveira LLD, Antunes SC, Gonçalves F, Rocha O, Nunes B (2015) Evaluation of ecotoxicological effects of drugs on Daphnia magna using different enzymatic biomarkers. Ecotoxicol Environ Saf 119: 123–131. https://doi.org/10.1016/j.ecoenv.2015.04.028
- [40] Palma TL, Magno G, & Costa MC (2021) Biodegradation of paracetamol by some gram-positive bacterial isolates. Current microbiology, 78, 2774-2786.
- [41] Parolini M, Binelli A (2012) Sub-lethal effects induced by a mixture of three non-steroidal antiinflammatory drugs (NSAIDs) on the freshwater bivalve *Dreissena polymorpha*. Ecotoxicology 21:379– 392. https://doi.org/10.1007/s10646-011-0799-6
- [42] Poddar K, Sarkar D, Chakraborty D, Patil PB, Maity S, & Sarkar A (2022) Paracetamol biodegradation by Pseudomonas strain PrS10 isolated from pharmaceutical effluents. International Biodeterioration & Biodegradation 175, 105490.
- [43] Przybyła GW, Szychowski KA and Gmiński J (2021) Paracetamol–An old drug with new mechanisms of action. Clinical and Experimental Pharmacology and Physiology 48(1), pp.3-19.
- [44] Ramos AS, Correia AT, Antunes SC, Gonçalves F, Nunes B (2014) Effects of acetaminophen exposure in Oncorhynchus mykiss gills and liver: detoxification mechanisms, oxidative defense system and peroxidative damage. Environ Toxicol Pharmacol 37:1221–1228. https://doi.org/10.1016/j.etap.2014.04.005
- [45] Rhee J-S, Kim BM, Jeong CB, Park HG, Leung KMY, Lee YM, Lee JS (2013) Effect of pharmaceuticals exposure on acetylcholinesterase (AchE) activity and on the expression of AchE gene in the monogonont rotifer, Brachionuskoreanus. Comp BiochemPhysiol C 158:216–224. https://doi.org/10.1016/j.cbpc.2013.08.005
- [46] Rios-Miguel AB, Smith GJ, Cremers G, van Alen, T, Jetten MS, den Camp HJO and Welte CU (2022) Microbial paracetamol degradation involves a high diversity of novel amidase enzyme candidates. Water Research X, 16, p.100152.
- [47] Rotundo L and Pyrsopoulos N (2020) Liver injury induced by paracetamol and challenges associated with intentional and unintentional use. World Journal of Hepatology 12(4), p.125.
- [48] Solé M, Shaw JP, Frickers PE, Readman JW, Hutchinson TH (2010) Effects on feeding rate and biomarkers responses of marine mussel experimentally exposed to propranolol and acetaminophen. Anal Bioanal Chem 396:649–656. https://doi.org/10.1007/s00216-009-3182-1
- [49] Swann JP (1994) FDA and the practice of pharmacy: prescription drug regulation before the Durham-Humphrey Amendment of 1951. Pharmacy in History 36(2), pp.55-70.
- [50] Takenaka S, Okugawa S, Kadowaki M, Murakami S, & Aoki K (2003) The metabolic pathway of 4aminophenol in Burkholderia sp. strain AK-5 differs from that of aniline and aniline with C-4 substituents. Applied and Environmental Microbiology 69(9), 5410-5413.
- [51] Trombini C, Hampel M, & Blasco J (2016) Evaluation of acute effects of four pharmaceuticals and their mixtures on the copepod Tisbebattagliai. Chemosphere 155, 319-328.
- [52] Valverde JL (2006) Proposal for a Regulation of the European Parliament and of the Council on Advanced Therapy Medicinal Products and amending Directive 2001/83/EC & Regulation (EC) No. 726/2004 (COM (2005) 567).
- [53] Weiner L, Kreimer D, Roth E, Silman I (1994) Oxidative stress transforms acetylcholinesterase to a molten globule-like state. BiochemBiophys Res Commun 198:915–922. https://doi.org/10.1006/bbrc. 1994.1130
- [54] Wu S, Zhang L, & Chen J (2012) Paracetamol in the environment and its degradation by microorganisms. Applied microbiology and biotechnology 96, 875-884.
- [55] Yang CW, Chen YE, & Chang BV (2020) Microbial communities associated with acetaminophen biodegradation from mangrove sediment. Sustainability, 12(13), 5410.

- [56] Yasunaga M, Matsuda S, Murata M (1985) Two cases of acute liver injury caused by ingesting small dose of acetoaminophen. Acta Hepatologica Japonica 26: 493- 499
- [57] Zhang L, Hu J, Zhu R, Zhou Q, & Chen J (2013) Degradation of paracetamol by pure bacterial cultures and their microbial consortium. Applied microbiology and biotechnology 97, 3687-3698.
- [58] Żur J, Piński A, Marchlewicz A, Hupert-Kocurek K, Wojcieszyńska D, & Guzik U (2018) Organic micropollutants paracetamol and ibuprofen—toxicity, biodegradation, and genetic background of their utilization by bacteria. Environmental Science and Pollution Research 25, 21498-21524.