

IN VITRO ANTI-INFLAMMATORY AND ANTIDIABETIC ACTIVITY OF *HIBISCUS ROSA SINENSIS*

Abstract

Natural plant products are growing in popularity nowadays as a result of increase in the number of illnesses. The Malvaceae family includes the plant *Hibiscus rosa sinensis* Linn, which may exist everywhere in the globe. Its leaves, bark, roots, and flowers have been employed to treat a number of ailments in traditional Indian medicine. *Hibiscus rosa sinensis*, commonly known as the Chinese hibiscus, a plant whose traditional medical uses date back a long time. This research sought to determine if an extract of *Hibiscus rosa sinensis* (Red tea) would have any *in vitro* anti-inflammatory and anti-diabetic properties. Through an *in vitro* experiment that measured the suppression of protein denaturation and anti-proteinase activity, the anti-inflammatory efficacy was assessed. The anti-diabetic activity was assessed through *in vitro* suppression of the enzymes α -glucosidase and α -amylase. *Hibiscus rosa sinensis* red tea was used for the activity and compared to acarbose in doses of 10, 20, 30, 40, and 50 g/ml. Anti-proteinase and Protein denaturation model's activity was measured and compared to diclofenac sodium 10–50 g/ml. The findings showed that extracts of *Hibiscus rosa sinensis* had strong anti-inflammatory and anti-diabetic effects that varied with dosage. Both the anti-inflammatory and the antidiabetic effects of red tea were notable. However, to effectively use these effects as medicinal treatments, these effects must first be verified using *in vivo* models.

Keywords: *Hibiscus rosa-sinensis*, Antidiabetic activity, Anti-inflammatory activity, Red Tea.

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I. INTRODUCTION

The prevalence of diabetes, a fatal and incurable condition, is rising globally. Diabetes incidence is steady [1]. The critical factor controlling blood sugar is insulin. In the case of diabetes mellitus, the body cannot utilize or create enough insulin on its own. Blood sugar is a result of this. Diabetes can cause major health issues like amputations, heart disease, blindness, and kidney failure. It is the seventh most frequent cause of death in the United States. In adults over the age of 18, diabetes was expected to affect 9% of people worldwide in 2014. Over 80% of fatalities from diabetes take place in low- and middle-income nations [2]. In 2030 diabetes will overtake smoking as the seventh biggest cause of death, according to the WHO [3]. The prevalence of diabetes is expected to more than triple from 171 million cases in 2000 to 366 million cases in 2030, with India experiencing the biggest rise [4]. Up to 79.4 million Indians are expected to have diabetes by 2030. Due to the negative side effects of using insulin and oral hypoglycaemic medications, there is an increasing demand from the market for alternative diabetes treatment options such as herbal remedies [5]. More than 200 plants are thought to possess anti-diabetic qualities. The 21,000 plant species used as medicines worldwide are recorded by the World Health Organization (WHO) [7]. The capacity to reduce glucose levels has been found in more than 400 medicinal plants [8]. Different civilizations throughout the world have employed herbal plants for treating diabetes for a number of years [9].

Heat, redness, discomfort, swelling, and altered physiological processes are all signs of inflammation, which is a complicated biological reaction of human tissues to adverse stimuli such as pathogens, damaged cells, irritants, injury, infection, or destruction [10]. Chemical mediators released from the damaged tissue and migratory cells serve as its trigger. Non-steroidal anti-inflammatory medicines (NSAIDs) are often prescribed medications for the treatment of inflammatory disorders. These medications inhibit COX-1 and COX-2 enzymes from producing prostaglandins. NSAIDs have a number of side effects, but stomach irritation and subsequent gastric ulcer development are among the most serious. The liver, gastrointestinal tract and other human biological systems are damaged when these medications are used over an extended period of time. Therefore, a novel anti-inflammatory medicine that is safe, effective, nontoxic or less toxic is needed. In many impoverished nations, plant medicines are quite significant in primary healthcare. Anti-inflammatory medications function as stabilizers of the lysosomal membrane. The *in vitro* anti-diabetic and anti-inflammatory activities of *Hibiscus rosa-sinensis* flowers were assessed in the current work. The red tea obtained from the flowers was tested for α -glucosidase inhibition activity, α -amylase inhibition activity, anti-proteinase, and protein denaturation ability.

II. MATERIALS AND METHODS

- 1. Collection of Hibiscus Flowers:** Flowers from *Hibiscus rosa-sinensis* were acquired in February in Hyderabad, Telangana, and were verified by botanist Harikrishna from Osmania University.
- 2. Preparation of Red Tea:** Pick fresh hibiscus flowers separate all the petals from the flowers and add the petals into a sufficient quantity of water to allow it to boil for some time (purplish pink color). Allow it to cool; filter it to get red tea extract [11] represented in Figure 1



Figure 1: Red Tea Extract Obtained from *Hibiscus Rosa-Sinensis* Flowers

- 3. Calculation of the Extraction Yield:** The extraction yield was calculated using the following equation (%)

$$\text{Extraction yield} = \frac{\text{weight of the extract following solvent evaporation and freeze drying}}{\text{dry mass of the sample}} \times 100$$

- 4. Phytochemical Screening:** Screening for phytochemicals was done for the red tea [12].
- 5. Proteinase Inhibitory Assay:** The Oyedepo and Femurewa-modified technique for the proteinase inhibitory test was used. The reaction mixture (2 ml) contained varied concentrations of the test plant extract sample, 0.06 mg of trypsin, and 1 ml of Tris-HCl buffer (20 mM, pH 7.4). The reaction mixture received 1 ml of 0.8% (w/v) casein after 5 minutes at 37 °C of incubation. For an additional 20 minutes, the mixture was incubated. To halt the process, 2 ml of 70% perchloric acid was added. The supernatant's absorbance at 210 nm was measured using a Tris-HCl buffer as a reference after centrifuging the cloudy solution. Three times the experiment was run. [13].
- 6. Inhibition of Protein Denaturation Method:** With minor adjustments, the approach was used to determine the inhibition of protein denaturation. The reaction mixture included 1% BSA (aqueous solution) and various amounts of the test extract. 1 N HCl was used to change the pH of the reaction mixture. The samples were heated for 20 minutes at 37 °C, followed by 20 minutes at 57 °C, and then allowed to cool [14]. At 660 nm, the samples' turbidity was determined. Three duplicates of the experiment were run.

The following formula was used to compute the percent inhibition of protein denaturation:

$$\text{Percentage inhibition} = (A_C \text{ of control} - A_C \text{ of test sample}) \times 100 / A_C, \text{ where } A_C \text{ and } A_S \text{ represent the absorbances of the control and sample, respectively, at 600 nm.}$$

- **α -amylase Inhibitory Activity:** Red tea's ability to inhibit α -amylase was tested using the conventional procedure with a few minor modifications. The reaction mixture was preincubated at 37°C for 20 min using the following ingredients: 50 μ l phosphate buffer (100 mM, pH = 6.8), 10 μ l α -amylase (2 U/ml), and 20 μ l of various

concentrations of red tea (10, 20, 30, 40, and 50 µg/ml). The substrate was then added, 20 µl of 1% soluble starch (100 mM phosphate buffer pH 6.8), and incubated for a further 30 minutes at 37°C. Next, 100 µl of the DNS color reagent was added and heated for 10 minutes. Using a UV Spectrophotometer, the mixture's absorbance at 540 nm was determined. As a standard, acarbose was utilized in a range of concentrations (10, 20, 30, 40, and 50 µg/ml). Each experiment run in triplicates and uses a control drug (red tea) in parallel to the test substance.

Percentage inhibition was used to express the results, and it was computed using the method [15].

Inhibitory activity (%) = $(1 - A_s/A_c) \times 100$, Both A_s and A_c represent the absorbances in the presence of the test material and control, respectively.

- α-glucosidase Inhibitory Activity:** Red tea's α-glucosidase inhibitory activity was tested using the conventional procedure with a few minor modifications. The reaction mixture was pre-incubated at 37°C for 15 minutes and contained 50 µl of phosphate buffer (100 mM, pH = 6.8), 10 µl of α-glucosidase (1 U/ml), and 20 µl of various quantities of extract and fractions (0.1, 0.2, 0.3, 0.4, and 0.5 mg/ml). The reaction was then preceded for a further 20 minutes at 37°C with the addition of the substrate to 20 µl of P-NPG (5 mM). With the addition of 50 µl of 0.1 M Na₂ CO₃, the reaction was stopped. Using a UV Spectrophotometer, the emitted p-nitrophenol's absorbance was determined at 405 nm. As a standard, acarbose was added at various quantities (10, 20, 30, 40, and 50 µg/ml). A control configuration that was parallel to the test setup was used to conduct each experiment three times. The findings were obtained using the formula [16]

Inhibitory activity (%) = $(1 - A_s/A_c) \times 100$, Where, A_s represents the absorbance while the test drug is present and A_c represents the absorbance of the control.

III. RESULTS

According to phytochemical study, the flavonoids, tannins, terpenoids, saponins, and alkaloids listed in Table 1 are the principal bioactive chemicals responsible for the chemical composition.

Table 1: Preliminary Phytochemical Screening

Phytochemicals constituents	Results
Flavonoids	++
Tannins	+
Terpenoids	+++
Saponins	++
Alkaloids	+

Note: + indicates presence

1. **Assessment of *in-vitro* Anti-Inflammatory Activity:** Different *Hibiscus rosa sinensis* extracts were tested for their ability to reduce protein denaturation, proteinase activity, and *in vitro* anti-inflammatory potential was estimated.
2. **Antiproteinase Action:** In this method the red tea extract from *Hibiscus rosa sinensis* showed graded inhibition which was found to be increasing with increasing concentrations i.e., at 10 µg/ml it showed 15% inhibition and at 50 µg/ml it exhibited 71% when compared to the standard Diclofenac Sodium at 10 µg/ml it showed 10% inhibition and at 50 µg/ml it exhibited 57%. Comparing the IC₅₀ value of red tea to that of diclofenac sodium, which was determined to be 34.09 µg/ml in Table 2, the red tea value was found to be 38.46 µg/ml.

Table 2: Antiproteinase Activity of *Hibiscus rosa sinensis* red Tea and Standard Drug

Name of the compound	Concentration (µg/ml)	% inhibition	IC ₅₀
<i>Hibiscus rosa sinensis</i> red tea	10	15	38.46
	20	23	
	30	39	
	40	57	
	50	71	
Diclofenac Sodium	10	10	34.09
	20	15	
	30	44	
	40	52	
	50	57	

3. Inhibition of Protein Denaturation Method

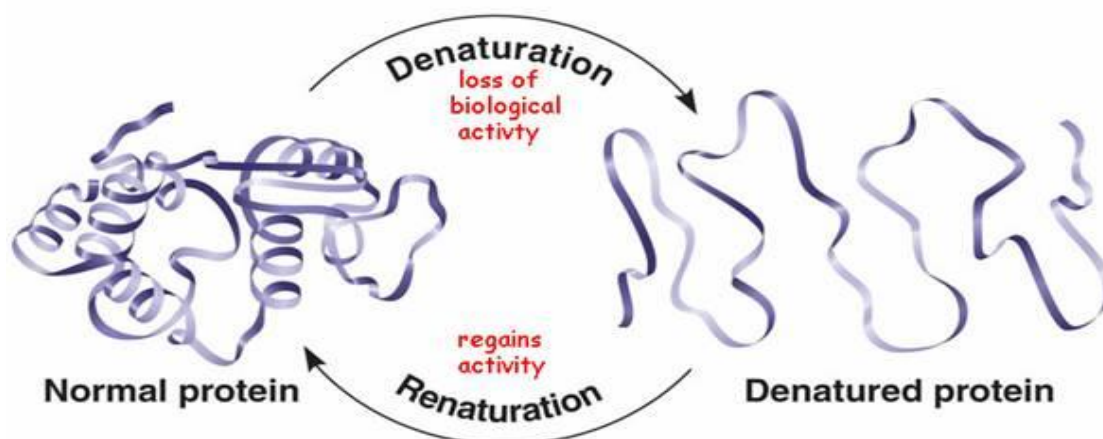


Figure 2: The Way that the Proteindenaturates



Figure 3: Extraction of Egg Albumin

4. **Protein Denaturation Inhibition:** In this method, the red tea extract from *Hibiscus rosa sinensis* showed graded inhibition, which was found to be increasing with increasing concentrations, i.e., at 10 $\mu\text{g/ml}$ it showed 29% inhibition, and at 50 $\mu\text{g/ml}$ it exhibited 69% when compared to the standard Diclofenac Sodium at 10 $\mu\text{g/ml}$ it showed 24% inhibition and at 50 $\mu\text{g/ml}$ it exhibited 90%. According to Table 3, red tea's IC₅₀ value was determined to be 31.323 $\mu\text{g/ml}$, which was higher than diclofenac sodium's IC₅₀ value of 3.479 $\mu\text{g/ml}$ Figure 2 depicts the denaturation process, while Figure 3 shows how to remove albumin from eggs.

Table 3: Protein Denaturation of *Hibiscus rosa sinensis* Red Tea and Standard Drug

Name of the compound	Concentration ($\mu\text{g/ml}$)	% inhibition	IC ₅₀
Diclofenac sodium	10	29	31.323
	20	38	
	30	47	
	40	53	
	50	69	
<i>Hibiscus rosa sinensis</i> red tea	10	24	39.47
	20	25	
	30	38	
	40	68	
	50	90	

5. Assessment of *in vitro* Anti Diabetic Activity

- **α -Amylase Inhibition Assay:** In this method, the red tea extract from *Hibiscus rosa sinensis* showed graded inhibition, which was found to be increasing with increasing concentrations, i.e., at 10 $\mu\text{g/ml}$ it showed 31% inhibition, and at 50 $\mu\text{g/ml}$ it exhibited 85% of the standard acarbose, and at 50 $\mu\text{g/ml}$ it exhibited 80% inhibition. In comparison to acarbose, which exhibited an IC₅₀ value of 21.6 $\mu\text{g/ml}$, red tea was

discovered to have an IC₅₀ value of 38.46 µg/ml, as shown in Table 4. Figure 4 shows the method by which it functions.

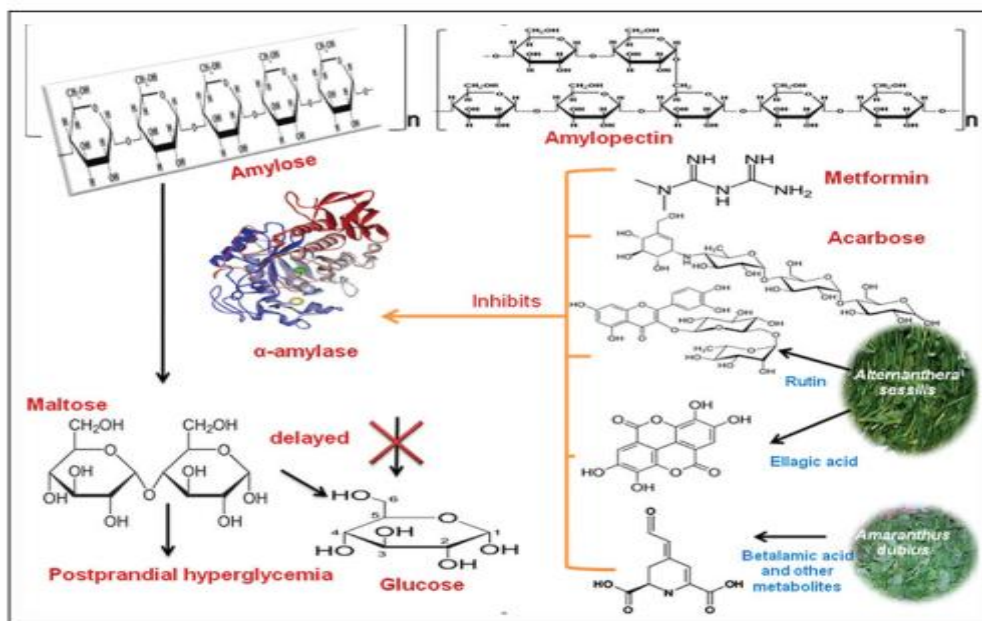


Figure 4: Mechanism of α-amylase Enzyme in Inhibition Assay

Table 4: α-amylase Activity of *Hibiscus rosa sinensis* Red Tea and Standard Drug

Name of the compound	Concentration (µg/ml)	% inhibition	IC ₅₀
Acarbose	10	31	21.6
	20	46	
	30	55	
	40	83	
	50	85	
<i>Hibiscus rosa sinensis</i> (red tea)	10	14	38.6
	20	26	
	30	39	
	40	53	
	50	80	

$$\text{Percentage inhibition} = (\text{Abs control} - \text{Abs sample}) \times 100 / \text{Abs control}$$

- **α-Glucosidase Inhibition Assay:** In this method, the red tea extract from *Hibiscus rosa sinensis* showed graded inhibition, which was found to be increasing with increasing concentrations, i.e., at 10 µg/ml it showed 31% inhibition, and at 50 µg/ml it exhibited 89% of the standard acarbose, and at 50 µg/ml it exhibited 78%. According to Table 5, red tea's IC₅₀ value was discovered to be 44.1 µg/ml, which was higher than acarbose's IC₅₀ value of 27.7 µg/ml.

Table 5: α -Glucosidase Inhibition Assay of *Hibiscus rosa sinensis* Red Tea and Standard Drug

Name of the compound	Concentration ($\mu\text{g/ml}$)	% inhibition	IC ₅₀
Acarbose	10	31	27.7
	20	36	
	30	57	
	40	78	
	50	89	
<i>Hibiscus rosa sinensis</i> (red tea)	10	10	44.11
	20	21	
	30	34	
	40	63	
	50	78	

$$\text{Percentage inhibition} = (\text{Abs control} - \text{Abs sample}) \times 100 / \text{Abs control}$$

IV. DISCUSSION

Hibiscus rosa sinensis red tea extract was tested for its anti-inflammatory properties using the anti-proteinase and protein denaturation method. Alkaloids, tannins, flavonoids, steroids, and phenols, which are polyphenolic chemicals found in the plant extract, imply that the extract has potent anti-inflammatory properties. The well-understood cause of inflammation is the denaturation of proteins. Plant extracts' ability to stop the denaturation of protein is investigated. When compared to the standard medication diclofenac sodium, the extract was efficient at preventing protein denaturation [17]. Proteinase inhibitors have been demonstrated to provide a high level of protection against tissue damage, which has been linked to leukocyte proteinase's role in the occurrence of inflammatory reactions. Contrasting the red tea extract with the generic medication [18].

The Pancreatic α -amylase and intestinal α -glucosidase enzyme are responsible for the carbohydrate metabolism, which raises blood sugar levels. When these enzymes are inhibited, the metabolism of carbohydrates slows down, which reduces the amount of glucose absorbed from the gut. The main phytoconstituents such as phenols, flavonoids, tannins and other bioactive substances possess α -amylase and α -glucosidase inhibitory activity resulting in regulation of postprandial hyperglycaemia. The efficacy of the red tea extract of *Hibiscus rosa sinensis* to inhibit alpha-amylase and alpha-glucosidase was investigated in this work. As a result, the current findings show that *Hibiscus rosa sinensis* may be useful in the treatment of postprandial hyperglycemia. [19].

V. CONCLUSION

Flowers contain a lot of flavonoids and other bioactive substances, which may explain why they have a hypoglycaemic impact. Additional research is required to clarify the mechanism of action and establish its anti-diabetic potential in people. The red tea extract

stopped heat-induced denaturation of proteins and proteinase activity from occurring due to the existence of polyphenolic compounds such as alkaloids, flavonoids, tannins, steroids, and phenols that have significant anti-inflammatory characteristics. This study suggests that a constituent of the *Hibiscus rosa sinensis* plant may provide the basis for the creation of potent anti-inflammatory and anti-diabetic drugs that may potentially be utilized to treat other ailments.

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