BIOTECHNOLOGICAL INTERVENTIONS FOR MINERAL ACCUMULATION IN RICE

Abstract

Fighting against hunger is still a challenge for humanity because of the continuously growing global population and it cause food shortages. Millions of individuals have "hidden hunger" or a lack of micronutrients (Zn and Fe), especially in developing nations. Mineral accumulation in food grains is an effective strategy to tackle this issue and provide an affordable and feasible method for distributing micronutrients to a population through a diverse diet. The popularity and consumption of white/polished rice have increased and its resulting in a loss of nutritional value. As a result, to promote beneficial health effects to the population, enhanced nutrigenomic value of white rice by incorporating the gene of rice bran layer of brown rice through biofortification processes like agronomic practices, traditional breeding, gene overexpression, CRISPR technology, and RNAi techniques. Therefore, the focus of this chapter is on improving the nutritional properties of white or polished rice by accumulation mineral using biotechnological and molecular approaches.

Keywords: Biofortification, Rice, Antinutrient, Malnutrition, CRISPR technology, RNAi

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I. INTRODUCTION

Due to "hidden hunger," or a diet lacking in iron and zinc, about 14.5 million males, 28.2 million pregnant women, and 85.7 million children in India experience anemia each year [1]. Increasing the amount of micronutrients in food crops is known as biofortification. Rice is the most essential food source for humans and will feed more people for longer than any other crop [2]. Rice is the primary source of calories for more than 3,500 million people worldwide. In the future, rice will remain an essential staple food for billions of people. It will become one of the world's most important agricultural products, close to the country's food security, employment, and economic development. The rice crop is estimated to cover 164.7 million acres [3]. In countries where human Fe deficiency is frequent, rice is a widely consumed staple food. The amount of Fe and Zn in polished/white rice is insufficient to meet human nutritional needs. The average grain Fe content in regularly grown rice cultivars must be increased by 7.5 times to reach the goal Fe concentration of 15 mg kg-1 for biofortification in polished rice. In contrast, there is only a small variance in the polished grain Fe contents in rice germplasm, which lie broadly between 1 and 11 mg kg-1 [5]. We need to produce more staple grains that should be of the highest nutritional value in sustainable ways.

There are three primary methods for incorporating vital micronutrients into agricultural plants through biofortification (Fig 1): Agronomical practices, Conventional breeding, and new breeding technologies (Molecular and biotechnological approach)

- **1. Agronomic techniques:** In this method we are applying fertilizer, foliar spray, nutrient primers and microbes' base culture on plants for increasing their nutrients value. Also known as ferti-fortification.
- 2. Conventional breeding: cross between two parents in which one is high yielding variety and one is mineral rich variety
- **3.** New breeding technologies: It includes Molecular Breeding & Transgenic Approach in which transfer genetic materials in plants.

There are many limitations of agronomic and breeding approaches. When we are applying fertilizer, small fraction of the applied fertilizer is absorbed by plants, and a large portion is lost, causing serious environmental pollution. Other disadvantages like, breeding approaches is depending on existing gene pool, takes a long time, in crosses no guarantee of particular gene combination, undesirable genes can be transferred (Linkage drag), effects of environment and genotype interaction, relies only on phenotypic selection and improvement of many traits is not possible [4]. Through these new molecular techniques, we are able to directly improve crops by insertion or deletion of a certain segment of a gene and in the end we get the desired plant without interference for other good characters, to be more productive and save time. These NBTs can be distinguished from other GMO plants due to their stable and definite mutation [2].



Image Source: [15]

II. WHY RICE?

- Rice is a staple food For more than half of the world's population
- India is the world's 2nd largest producer of rice and the 1st largest exporter of rice in the world
- Rice is the maximum cultivated crop in India. So, rice is major source of employment for our Indian farmers
- Rice is an excellent option to overcome malnutrition
- Rice plays an essential role in food security
- Time has come to play a role in nutritional security

III. LIMITATION FOR MINERAL ACCUMULATION IN RICE

Brown rice contains, on average 90% portion of endosperm, 6-7% rice bran and 2-3% embryo by weight [7]. Bran, unlike endosperm, is a large storehouse of lipids, vitamins, proteins, minerals, and dietary fiber [8], [9]. Recent X-ray micro fluorescence studies have shown that Zn, Fe, and potassium (K) concentrations reduced in the following order: bran > hull > whole grain > brown and polished rice [10], [11]. Zn and Fe are distributed throughout the endosperm, but the concentration in the bran is approximately three times higher than that in the rice husk and endosperm [12] [10], the husking and polishing of the rice removes the bran from the rice, thereby enabling the rice to be polished. It consumes elements that are not found in the food of many consumers. Therefore, it is vital to increase the Zn and Fe concentration in the rice endosperm [6]. The phytic acid in grains is another limitation due to their chelator nature. About 80% of the total phytic acid in most cereals builds up in the aleurone layer. Phytic acid stored as mixed salts and this complex called phytate. Due to its six negatively charged ions, phytate is a powerful chelator of divalent cations such Fe2+, Zn2+, Ca2+, Mg2+ and lowers their bioavailability [13]. Monogastric animals cannot secrete phytase enzyme so that they cannot digest or breakdown phytate minerals complex. It is necessary to reduce phytic acid for achieve enough availability of minerals from rice. NBTs improve the mineral content in grains by up regulating metal-storage proteins such lactoferrin, ferritin and down regulating phytic acid pathways [14].

IV. MOLECULAR AND BIOTECHNOLOGICAL APPROACHES

Along with fertile fortification, biotechnological approaches can be employed to incorporate genes that increase the content of micronutrients, their bioavailability and decrease of the concentration of antinutrients. This chapter focuses on three techniques: Overexpression of genes, RNA interference, and CRISPR-Cas9.

1. Over Expression of Gene: Gene over-expression is the process that leads to abundant target protein expression by adding constitutive/vital promoter regulatory elements before the target gene so that genes can be transcribed and translated efficiently. There are many ways of obtaining the target gene like getting from the gene libraries, PCR-amplification of the target gene, designing and synthesizing the target gene construct.

Promoters are added to the upstream region of the gene and this transgene is inserted into the plasmid by restriction enzyme and ligase enzyme. This recombinant artificial Vector is transferred into a plant by many gene transformation techniques like Agrobacterium-mediated transfer, electroporation, particle bombardment method, microinjection, etc. It makes a transgenic plant with desired traits.



Figure 1: Schematic Representation of Transgenic Plant Development

2. RNA Interference: Tissue-specific RNAi-mediated silencing of the phytic acid gene significantly reduced the phytate levels in seeds without interfering germination or plant growth. The short RNA molecule bind with the target mRNA and make it functionally inactivates target mRNA and sometimes its leads to the degradation for prevent translation. The small interfering RNA molecules (siRNAs) affiliated with the three catalytic core elements Dicer, Argonaute (AGO), and RNA-dependent RNA polymerase (RdRP). The ribonuclease enzyme, one of the several proteins in RISC which are responsible for enzymatic cleavage of double stranded RNA [16].



Figure 2: Mechanism of RNA interference through SiRNA biosynthesis

Double strand RNA with sense and anti-sense strands is inserted into a vector and makes an artificially constructed Vector. This Vector transforms into a plant cell and once this construct is rich in the plant cell cytoplasm, the dicer enzyme cuts this ds RNA strand (hairpin structure) and makes the duplex structure. Next, the RISC complex binds and this whole complex structure binds with targeted m-RNA and silences their function.

3. CRISPR-Cas9 Technique: This technique is one type of natural immunity system of bacteria for defense against viruses. When viruses attack on bacteria, a small DNA template inserted into the bacteria. Bacteria memorized it and that's called RNA arrays. Now again this viruses attack, this RNA arrays guide and detected infectious DNA template. The cas9 enzyme of bacteria cut this infectious DNA template and disabled viruses



Figure 3: CRISPR-Cas9 Gene Editing

Nowadays in biotech research, this cas9 enzyme used in many experiments. Cas9 isolated from bacteria and guide RNA (gRNA) construct for target gene. This cluster called CRISPR and insert into target host for disable the target protein.

V. CASE STUDIES OF MINERAL ACCUMULATION BY BIOTECHNOLOGY

1. Increasing Iron Mineral in rice by Molecular Breeding Method

• **Objective:** Overexpression of *Osfer2* gene (endogenous Ferritin gene) for the development of cisgenic rice plants

• Methodology:

- The gene Osfer2 from the Swarna rice variety was overexpressed in Pusa Sugandhi II.
- > Total RNA was isolated from Swarna seeds and cloned the Osfer2 gene
- Genetic transformation via the biolistic method in Pusa Sugandhi II
- Total RNA was extracted from the transgenic plants (mature dehusked seeds) for semi-qRT-PCR
- Analysis of Fe & Zn concentration in grains histochemical localization of iron in grain tissues

Results



Figure 4: PCR and qRT-PCR Analysis

The result shown that PCR analysis of different 7 transgenic plant seeds and roots sample compared with non-transgenic (NT) plants. As per the PCR analysis, NT plants showing lower ferritin genes with tubulin reference gene. Transgenic seeds showed greater ferritin transcript expression compared to other plant components (shoots). The qRT-PCR of 7 transgenic plants showed 7.8-fold overexpression of ferritin gene in transgenic rice seeds over NT rice seeds.



Figure 5: Morphology of T2 Pusa Sugandhi II and Prussian blue Staining

Morphology of transgenic and non-transgenic pusa Gandhi II showed that no any physical difference after gene transformation. The distribution of ferric ions in the endosperm of transgenic seed was validated by Pearl's Prussian blue staining of transgenic seeds. This distribution was revealed by the insoluble blue coloring of potassium ferric ferrocyanide complex due to ferric ion (Fe3+) interacting with potassium ferrocyanide. Due to the lack of iron in the endosperm, non-transgenic seeds were found to be lighter in color [17].

2. Decrease Anti-Nutrient Cadmium in rice by CRISPRCas9 Genome Editing Technology

- **Objective:** Knockout mutants of *OsCCX2* genes (Cadmium) are generated by the CRISPR/cas9 editing method
- Methodology
 - Genome editing technology (CRISPRCas9) to target the cadmium transporter gene (OsCCX2) in Nipponbare rice
 - Genetic transformation of Agrobacterium cells EHA105 with CRISPR-Cas9CCX2 gene
 - Rice seeds grow in a hydroponic solution, and the plants were harvested for Cd determination in WT, ccx2-1 and ccx2-2
 - > Total RNA sample isolated for semi-qRT-PCR
 - > Analysis of Cd Concentration in grains was quantified by Spectroscopy

35 level(/OsActin2) storn node nowe 23 21 sCCX2 expression ctin 7 Relative root shoot leaf stem node flow b d h a e C g (a) The leaf sheath. (b) Node I. (c) Node II. (d) Node III. (e) Node IV. (f) Unelongated basal stem. (g) Four-day-old seedling grown on MS agar plates. (h) Two-week-old seedling grown in

Results

Figure 6: The semi-quantitative-RT-PCR analysis of *OsCCX2* transcript levels & Histochemical GUS staining of *OsCCX2*

They initially examined the expression pattern of OsCCX2 using a semiquantitative RT-PCR technique and found a clear DNA band in the node tissue and weak bands in other tissues. Strong GUS signal was exclusively found in node tissues while weak GUS signal was seen in internode tissues. The OsCCX2 gene tends to be expressed near the nodes, which suggests a connection to cation buildup in grains.



Figure 7: Translocation ratio of Cd

The calculated root-to-shoot and shoot-to-root translocation ratio of transgenic ccx2 mutants (average, 5.0%) much lower than those of the wild type plants [18].

3. Reduction of phytic acid for enhancing phosphorus concentration in rice by molecular breeding

- Objective: Generate low phytic acid rice by over-expressing *appA* gene
- Methodology:
 - ▶ Isolation and cloning of *appA* gene from E. coli
 - Vector construction with *pUC- zein-appA-nos* gene
 - Rice tissue culture and Agrobacterium-mediated genetic transformation
 - > DNA isolated from leaves of transgenic plants for PCR
 - Total RNA isolated from transgenic seeds for cDNA synthesis and gene expression
 - Determination of total phosphorus, inorganic phosphorus, and phytic acid by Spectroscopy



Figure 8: Semi qPCR Analysis of Transgenic Seeds

The semi qPCR analysis results shown that the non-transgenic seeds (L2) did not exhibit any transgene amplification while L3 to L9 transgenic seeds shown positive results. The housekeeping gene β -tubulin showed amplification in both transgenic and nontransgenic plants.



Figure 9: Histochemical *gus* analysis of transgenic rice lines

The phosphorus quantity increased four times whereas the phytate content of the transgenic seeds lowered by about 45%. Polished seeds from transgenic plants contained two and three times as much increased iron and zinc, respectively. Transgenic seeds' maturation behavior and other physical features remained unchanged. This finding demonstrates that *appA* gene reduce phytic acid in rice seeds without impairing the plant's other physiological functions or morphological cost [19].

4. RNAi technology use for reduces phytic acid content and enhance mineral distribution in rice seeds

- **Objective:** RNAi-mediated silencing of *OsITP5/6K-1* gene
- Methodology
- > Cloning of rice *ITP5/6K-1* gene and RNAi vector construction
- > Tissue culture of rice plants and *Agrobacterium* mediated genetic transformation
- DNA was extracted from transgenic leaves for PCR-based screening of transgenic plants
- > Total RNA isolated from transgenic seeds for qRT-PCR expression analysis
- Determination of total phosphorus
- > Analysis of (Fe^{2+}) , (Mg^{2+}) , (Mn^{2+}) and (Zn^{2+}) content in milled seeds of nontransgenic (NT) and transgenic seeds by Atomic Absorption Spectroscopy

	A 510 um	Non-transgenic	Zn 510 um	Transgenic	B503 µr	Non-transgenic	Fe 503 (Transgenic
Unstained	Em.		En		End Em_	-		End
	AL5 <u>06.5</u> μm		AL- 501.3 µm		AL	1	AL	m
Stained with dithizone	Em	10	ind AL		End En-		End Em-	End

• Results

Figure 10: Histochemical localization of Zn and Fe

Dithizone and Perl's Prussian blue were used to stain NT and transgenic rice grains in order to evaluate the histochemical localization of Zn and Fe. μ -XRF imaging analysis, in which transgenic rice grains displayed higher color intensity indicating a greater accumulation of Zn and Fe in the endosperm region compared to NT rice grains.



Figure 18: Analysis of Iron, Magnesium, Manganese and Zinc Content in Milled Seeds by Atomic Absorption Spectroscopy

Fe content increased by 1.3 times when compared to NT, whereas Mg, Mn, and Zn also increased by 1.4, 1.7, and 1.6 times, respectively. Iron and zinc concentration in the grain endosperm were both increased by 1.3 and 1.6 times in low phytate polished seeds, respectively. their findings demonstrated that RNAi technology of the *OsITP5/6K-1* gene considerably decreased the amounts of phytate in seeds without impairing seed germination or growth of the plant [20].

VI. CONCLUSION

Globally, more than a billion people experience Fe and Zn deficiency. In areas where human Fe insufficiency is frequent, rice (Oryza sativa) is a common staple [5]. But polished/ white rice does not provide enough Fe and Zn to match human nutritional requirements. In rice germplasm, there is only a limited amount of minerals due to polishing. The development of Fe and Zn in rice is considered the best way to solve these problems. However, biological enrichment of Fe and Zn by Fertilizer and breeding methods in rice is very difficult due to insufficient genetic modification. At the same time, biotechnological intervention has led to an increase in the amount of Fe and Zn in rice. The development of effective genetic biological amplification techniques relies on knowledge of the functions of different genes involved in the uptake, translocation and storage of Fe and Zn. Overexpression of Osfer2 (increase 3.4-fold higher Fe) and *appA* (increase Zn) genes by molecular breeding method, Knockout OsCCX2 anti-nutrient gene (decrease 32.41% Cd) by CRISPR/Cas9 technology and silencing OsITP5/6K-1 gene (decrease 46.2% phytic acid and 3, 1.3, 1.6-fold enhancement in Pi, Fe, Zn respectively) by RNAi technology. It has become evident with different case studies that molecular and biotechnological approaches do increase micronutrients and decrease antinutrients.

Varieties name	Minerals increased	Year of Release	Released by		
CR Dhan 315	Zn 24.9 ppm	2020	ICAR-National Rice Research Institute, Cuttack		
DRR Dhan 45	rich Zn 22.6 ppm	2016	ICAR-Indian Institute of Rice Research,		
DRR Dhan 48	rich Zn 24 ppm	2018	Hyderabad		
DRR Dhan 49	rich Zn 25.2 ppm	2018			
Zinco Rice MS	rich Zn 27.4 ppm	2018	Indira Gandhi Krishi Vishwavidyalaya, Raipur (under ICAR-All India Coordinated Research Project on Rice)		
Mukul	rich Zn 20.1 ppm	2018	ICAR-National Rice Research Institute, Cuttack		
Swarna	2.54 fold Fe & 1.54 fold Zn		IRRI		
Surabhi	Rich Zn 22.84 ppm	2017	Nuziveedu Seeds Limited		
Chhattisgarh Zinc Rice-1	Rich Zn 22-24 ppm	2016	Indira Gandhi Krishi Vishwavidyalaya, Raipur		

Table 1: Biofortified Varieties of Rice in India

(Normal rice contain 12 ppm Zn)

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