MICROFLUIDICS: AN EMERGING TECHNIQUE

Abstract

Microfluidics, an innovative field that blends fluid dynamics, engineering, and biology, has revolutionized the manipulation and control of tiny liquid volumes within microscale channels. This abstract investigates recent progress in microfluidics and its profound impact on analytical and biomedical applications. Compared to traditional large-scale systems, microfluidic devices offer numerous benefits, such as reduced sample and reagent usage, heightened sensitivity, and improved control over fluid behavior. These devices consist of microchannels, chambers, valves, and pumps created using microfabrication techniques, allowing precise manipulation of fluids at the microliter or even nanoliter scale. In the realms of analytical and biomedical fields, microfluidics has proven to be a versatile and potent tool, enabling precise manipulation and analysis of small-scale fluids. The integration of microfluidics with other disciplines and technologies holds great potential for advancing diagnostics, drug development, personalized medicine, and more. Continuous research and collaboration in this field are poised to drive transformative breakthroughs in the near future.

Keywords: Microfluidics, DNA analysis and sequencing, Drug discovery, Tissue engineering.

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I. INTRODUCTION

The fields of microfluidics and microminiaturized device manufacture are related to the study of fluid behaviour in micro-channels and the confinement or movement of fluids via chambers and tunnels, respectively. In microfluidics, fluid quantities as tiny as femtoliters or one quadrillionth of a litre. On a micrometric scale, fluids behave quite differently from how they do in daily life; these distinctive characteristics are crucial for new scientific research and inventions. The main idea behind microfluidics is to include functions that often require an entire laboratory into a small, straightforward device.

Microfluidics is a multidisciplinary field that combines physics, engineering, chemistry, and biology to manipulate and control small volumes of fluids at the microscale level. In the context of biotechnology, microfluidics offers numerous applications and advantages due to its precise control over fluid flow, small sample requirements, high throughput, and integration of various functionalities on a single chip.This technology enables researchers to perform a wide range of biological and chemical experiments with high precision, speed, and efficiency [1].

In biotechnology, microfluidics offers a multitude of advantages over conventional techniques. The miniaturization of experimental systems brings several benefits, such as reduced sample and reagent volumes, shorter reaction times, and enhanced sensitivity. Additionally, microfluidic devices can integrate multiple functions onto a single chip, allowing for the automation and parallelization of complex experiments, leading to highthroughput analysis [2]. One of the primary applications of microfluidics in biotechnology is the development of lab-on-a-chip devices. These miniaturized platforms bring together various laboratory processes, such as sample preparation, mixing, separation, and detection, onto a single chip [3]. Furthermore, microfluidic systems have revolutionized DNA analysis and sequencing. By utilizing miniaturized reaction chambers, microfluidics enables highthroughput DNA amplification, sequencing, and genotyping with reduced costs and time. In the realm of drug discovery and development, microfluidics offers powerful tools for highthroughput screening of compounds, accelerating the identification of potential drug candidates [4].

Here are some prominent fields where microfluidics has contributed:

- **1. Diagnostic testing using a "lab-on-a-chip":** By combining several analytical procedures onto a single chip, microfluidic technologies have revolutionised diagnostic testing. With applications in point-of-care diagnostics, pathogen detection, and disease monitoring, these lab-on-a-chip devices make it possible to analyse biological samples quickly, sensitively, and affordably [5].
- **2. DNA Analysis and Genomics:** Microfluidics has played a crucial role in advancing DNA analysis techniques. It enables efficient DNA amplification methods such as polymerase chain reaction (PCR) by reducing reaction volumes and improving reaction kinetics. Microfluidic platforms also enable high-throughput DNA sequencing, single-cell genomics, and the analysis of genetic variations [6].
- **3. Cell Analysis and Manipulation:** Microfluidic systems provide precise control over cell manipulation and analysis. They allow for the sorting, separation, and isolation of cells based on various parameters such as size, morphology, and biomarker expression. Microfluidics also enables single-cell analysis, cell culture, and the study of cellular behaviour and interactions [7].
- **4. Drug Discovery and Screening:** Microfluidics has transformed the field of drug discovery by enabling high-throughput screening of compounds. Microfluidic platforms allow for miniaturized assays, efficient drug formulation optimization, and toxicity testing. They also facilitate the development of organ-on-a-chip systems that mimic the physiological environment for drug testing [5].
- **5. Chemical Synthesis and Analysis:** Microfluidics offers precise control over reactant mixing, reaction conditions, and reaction kinetics, making it valuable for chemical synthesis and analysis. It enables rapid synthesis of nanoparticles, microencapsulation of drugs, and controlled synthesis of complex chemical structures. Microfluidic platforms also enable on-chip chemical analysis, such as spectroscopy and chromatography [8]
- **6. Bioengineering and Tissue Engineering:** Microfluidics has contributed to advancements in bioengineering and tissue engineering. It enables the creation of microenvironments that mimic the structure and function of tissues and organs. Microfluidic devices are used to study cell-cell interactions, tissue development, and drug responses in a controlled and reproducible manner[9].

II. MAJOR INNOVATIONS CONTRIBUTED IN THE FIELD OF MICROFLUIDICS

Through the use of microfluidics, physical processes including osmotic movement, electrophoretic motility, and surface contacts may take place at a size (microns) where they are more advantageous. Sample sizes, test times, and procedure costs are decreased at the microscale. Microfluidic devices may interface with modern techniques and technology. It has been demonstrated that microfluidics can speed up the hybridization process for DNA microarray assays. The examination of picomole quantities of peptide in a controlled microenvironment is made possible by the integration of microfluidics with protein analysis technologies, such as mass spectrometry. Microfluidics' adaptability will make it easier to use it in the creation of assays for many biotechnological fields.

1. "Organ-on-a-chip" for Development of Drug: In terms of advancing biological research and diagnostics, microfluidics, a technique that allows for the engineered manipulation of fluids at the submillimetre scale, has shown a lot of promise. In order to reduce the expense of producing new drugs, experts in the field of microfluidics are tackling this issue by creating potentially game-changing technology. *In vivo* organ function is being attempted to be replicated on a microchip by a new class of microfluidic devices. This brand-new category of so-called "organ-on-a-chip" technology combines numerous well-understood microfluidic elements into a single in vitro device, enabling researchers to more accurately mimic in vivo function (both in healthy and pathological states) [5].

Source: https://link.springer.com/article/10.1007/s13534-022-00258-4 [10]

- 2. Genotoxicity Assessment of Biomaterials Using a Microfluidic System: Testing for genotoxicity is a crucial part of d determining the biocompatibility of novel materials being created for biomedical uses. However, because the traditional genotoxicity test (Ames test) is cumbersome and space-intensive, researchers studying biomaterials seldom employ it. One of the studyprovides a novel, more straightforward genotoxicity assay that operates on a microfluidic chip. Test is quicker and uses a lot less resources and room. Additionally, a microfluidics-based automation alternative with a control system has been developed. This enables scientists to assess genotoxicity early during the development of materials, increasing their patient safety. They created a novel method of testing for genotoxicity that substitutes fluorescent tagging of bacteria for the traditional Ames test's colony formation procedure. Unlike the traditional test, which requires 48 hours of incubation, this approach enables the identification of mutant bacteria after just 24 hours [11]. veloped. This enables
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- **3.** A Multi-Functional Microfluidic Injector for Droplet Identification: For droplet synthesis, microfluidics high throughput, accuracy, and affordability make it the best option. However, identifying and tracking droplets is still difficult.The study exploits pressure-controlled injection to achieve deterministic droplet coding for droplet pressure-controlled injection to achieve deterministic droplet coding for droplet
identification and tracking. For on-demand droplet injection and encoding, their system consists of a circular electrode and concentric fluidic channel with several injection ports. The scientists employed a color-coded method to further identify droplets using an image analysis algorithm and pressure pulse regulation to accurately inject droplet components. The developed platform is very adaptable, small, and simple to integrate, and it may be employed in applications that call for multiplexed compound screening, combinatorial synthesis, or other multistep assays. They created a flexible and multiplexable droplet injection, encoding, and tracking platform and integrated three injectors in a single device, all of which can be controlled individually to perform the on-demand injection of multiple targets or barcodes [12]. demand droplet injection and encoding, t
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- **4. Constructing Synthetic Cells' DNA-based Cytoskeletons:** Shape, internal organisation, cargo movement, and other crucial cell activities are all under the control of the cytoskeleton. Designing effective synthetic cells requires emulating such characteristics of natural cytoskeletons. Zhan and colleagues created a synthetic cell-sized system of a useful DNA-based cytoskeleton using DNA nanotechnology and microfluidics. The technology is made up of self-assembling filament networks made of DNA tiles that operate in a microfluidic compartment. Their synthetic cytoskeleton had traits shared with their natural counterparts, including ATP-triggered polymerization and reversible assembly. Their findings also pointed to autonomous cargo delivery via DNA strands. Overall, this work emphasises how DNA nanotechnology and microfluidics have enormous promise for creating synthetic cells from the ground up.This work illustrates the tremendous potential of DNA nanotechnology and microfluidics to duplicate biological systems and create synthetic cells from the bottom up[13].
- **5. Cyclic Preconcentration and Separation of Microorganisms on Chips:** Microfluidic analytical systems provide distinct benefits over conventional laboratory techniques, including mobility, operator-free operation, on-chip micro control capabilities, decrease in analysis time, and dramatically smaller sample volume. Microfluidic systems that incorporate immunochromatography, polymerase chain reaction, droplet reactions, electrophoresis, and other analytical techniques have proven to be more effective than conventional techniques. However, by first separating, concentrating, and purifying pathogens present in the sample from chemicals that obstruct the precise detection of pathogens and the estimation of their concentrations, the performance of many microfluidic assay methods may be markedly enhanced. Scientists demonstrate on-chip automatic efficient bacteria separation and preconcentration method with the use of pressure-driven flow-controlled microfluidics. This platform performs a fundamental yet crucial activity with the aid of pressure-driven flow-controlled microfluidics, which greatly expands its applicability and scalability [14].
- **6. Use of Microfluidics to Dynamically Screen and Print Single Cells:** The fundamental requirement to print single cells into separate chambers is present in single-cell clonal proliferation and sequencing using conventional equipment, as well as in other single-cell analyses. Their size depends on their types, operations, and even cell cycle stages. Individual cells must be printed within the necessary size range in order to do single-cell analysis. In order to dynamically screen and print single cells, a microfluidic chip with pneumatic microvalves is developed and the procedure included crucial instructions for single cell screening and printing (inoculating) into different cell culture wells. A dual microvalve mechanism is employed by the proposed microfluidic chip to screen cells for size. The dual microvalve system with pressure-driven flow control is used by the authors to dynamically regulate the size of the device.The efficiency of printing single endothelial cells was found to be 100%. This study shows the great potential of the microfluidic chip with dual microvalves in the field of single-cell inoculation for subsequent analyses [15].
- **7. In a Human Bladder-chip Model of Urinary Tract Infections, Uropathogenic Escherichia coli are being studied for its Dynamic Persistence:** A human bladder-chip model's operation is examined. In addition to employing pressure driven flow-controlled microfluidicsto provide a negative pressure within the bladder-chip, this model incorporates essential bladder physiology elements that are crucial to the early stages of UPEC infection. The bladder-chip model, which was developed utilising a platform with

a pressure-driven flow control mechanism, incorporates the essential bladder physiological characteristics that are crucial to the early stages of UPEC infection. In addition to administering antibiotics, this platform is appropriate for long-term live-cell imaging. The results unmistakably reveal that this model is applicable to immunological and drug delivery research and that IBCs are remarkably dynamic structures that offer adequate resistance to antibiotic clearance for a sizable period of time[16].

- **8. Development of a droplet-based microfluidic test for the assessment of the cytotoxicity of natural killer cells:** In order to develop a useful method for calculating and assessing the cytotoxic potential of NK-92 cells towards target cells, a technique known as a droplet-based microfluidicis investigated. With the aid of pressure-driven flow-controlled microfluidics, accurate droplet size control and pulseless flow control is made possible. The cytotoxic activity of NK-92 cells is assessed in this study after they are enclosed with haematological tumour cell lines in water-in-oil droplets of various diameters. This microfluidic cytotoxic test also assisted in identifying certain cells with multi-target killing capacity. The experiment may be readily repeated since the pressuredriven flow controller, Elveflow OB1 MK3+, makes controlling the droplet size a smooth operation. Cell-based immunotherapy and other contexts for biological and medical research are determined to be the real-world applications for this technology[17].
- **9. Generation of core-gap-shell microcapsules for stimuli responsive biomolecular sensing:** A variety of engineering applications, including sensors, actuators, medication delivery, and catalysis, might greatly benefit from the intricate design features of stimuliresponsive microparticles encapsulating important biomolecules. Thermoresponsive coregap-shell (TCGS) microcapsules constructed of poly N-isopropylacrylamide (PNIPAm) are used in this study to encapsulate hydrophilic payloads in a straightforward and reliable manner. A supersaturated aqueous solution of NIPAm is used to phase separate in a one-step microfluidic method to realise these facts. By individually controlling the swelling or by integrating pH-responsive co-monomers of the inner core and outer shell, a variety of microcapsule designs may be produced. Nanoparticles that resemble cargo can be placed in the gap, or the area between the inner core and outer shell. Smaller molecules from the external solution may be transported through the outer shell, which responds to stimuli. This study demonstrates the suitability of TCGS microcapsules as temperaturecontrolled glucose sensors and their potential to aid in the development of programmable enzymatic processes. The suggested platform offers a technique to create a new generation of microparticles with potential applications in several technical fields[18].
- **10. Microfluidics-based DNA Protection from Mechanical Stress:** A quite remarkable feature of our tissues and cells is their ability to endure mechanical stress in our daily lives. Any stretching, pulling, or compression could potentially lead to cell and tissue rupture and DNA damage, but this type of damage is rare. This raises the question: what are the mechanisms protecting our genome and cells from such external challenges? Recently, a team led by Sara Wickström from the University of Helsinki's Wihuri Research Institute and Helsinki Institute of Life Science conducted a thorough study of the mechanisms regulating DNA preservation in response to mechanical stress. The authors demonstrate how cells and tissues form two physiologically and temporally separate mechanisms for DNA protection and genomic integrity preservation using a variety of biomechanical and microfluidics reactors that can imitate stretching and compression *in vivo*. Together, results show that nuclei rapidly reduce lamina-associated

heterochromatin at the nuclear periphery near the nuclear envelope in response to mechanical stress in order to prevent DNA damage[19].

- **11. Dynamic Single Cell Screening:** Single cell screening helps us discover mechanisms which are not necessarily evident in bulk cell populations. Isolation and inoculation of single cells into separate culture chambers is a key requirement. Several highly sophisticated methods like fluorescence-activated cell sorting (FACS), Raman tweezers and laser capture are currently being used for single cell isolation. The authors offer a sophisticated method for screening single cells and inoculating them into different cell culture wells. A dual microvalve mechanism is used in the suggested microfluidic chip to select cells according to their size. To enable size-based cell screening, two nearby pneumatic microvalves were linked to a programmable pressure controller. Based on the pressure applied at each valve, the upper and lower limits for the cell screening may be dynamically regulated. The screening of individual beads and endothelial cells, as well as their subsequent printing into well plates, is shown to be 100% effective. In order to dynamically adjust the size limitations of the cells to be screened, the scientists used a pressure driven microfluidics with dual microvalve system. It is discovered that printing individual endothelium cells was 100% effective. Furthermore, the investigations on proven viability show that there is little cell damage[20].
- **12. Multivesicular Microfluidic Vesicles for the Creation of Synthetic Cells and Drug Delivery:** Multivesicular vesicles (MVVs) are vesicles-in-vesicle systems that are nonconcentrically organised and can be created from amphiphiles that form bilayers. Although MVVs can develop spontaneously, these hierarchical structures are often created on purpose via directed molecular assembly according to a number of wellestablished techniques. This succinct study addresses the microfluidic technology for producing MVVs and its present uses for medication administration or systems that mimic cells. Multivesicular microfluidic vesicles are potential instruments for the development of artificial-cell-like systems research and drug delivery systems. They offer additional protection for liposomes in the circulation during medication delivery while enabling fine-tuning of the system for improved release while taking into account the characteristics of the drug and administration route. They closely mimic biological cells in artificial cell-like systems, making it possible to investigate limited reaction systems and more intricate cellular behaviours like division and RNA transcription [21].
- **13. DNA-mediated Charge Amplification and the Electrokinetic Sandwich Test:** For the first time, an electrokinetic immuno-sandwich test using the streaming current approach of signal transduction is proposed by the authors in this work. Better target selectivity and a linear concentration-dependent response are provided for targets with concentrations between 0.2 and 100 nM. Additionally, the potential for signal enhancement by DNA conjugation is investigated on a theoretical and experimental level. DNA lengths are shown to clearly and consistently increase the detection signal. Target identification from a complicated *E. Coli* lysate medium served as a demonstration of how to use this technology. The authors demonstrate an electrical, label-free sandwich immunoassay that transmits signals via electrokinetic streaming current. Simulations suggested that by increasing the charge contrast between the target and the detecting probe, the signal may be improved. In an experimental setting, positively charged targets are found utilising detection probes coupled with negatively charged DNA. Additionally, it is demonstrated that as conjugated DNA length increased, the signal got stronger with time, up to 100% stronger with 30 nt of DNA. Furthermore, by detecting the target in a complex medium,

the *E. coli* cell lysate, the electrokinetic sandwich immunoassay's increased specificity is also established. Consequently, the research advances the development of highly selective and sensitive electrokinetic assays for possible application in clinical investigations [22].

Table 1: Microfluidics applications in Biotechnology

III. CONCLUSION

To sum up, the exact control and small-scale manipulation of fluids made possible by microfluidics has revolutionised the area of biotechnology. The ability to combine several operations on a single chip, fast throughput, cost-effectiveness, and miniaturisation are only a few of the benefits it has over conventional techniques. In several branches of biotechnology, microfluidic devices have found use, advancing research in genomics, proteomics, tissue engineering, drug discovery, and other fields as well as diagnostics and drug development.

Microfluidics has made it possible to create lab-on-a-chip devices, which combine many laboratory operations onto a single chip and give rise to portable and point-of-care diagnostic instruments. Additionally, it has made it easier to analyse individual cells, enabling researchers to study cellular heterogeneity and look into the behaviour of single cells. In addition, microfluidics has been significant in environmental monitoring, drug screening, organ-on-a-chip systems, DNA analysis, protein analysis, and protein analysis.

New insights and discoveries in the field of biotechnology have been made possible by the exact control and manipulation of fluids at the microscale in microfluidic devices. It has sped up research, made it more efficient, and opened up other directions for scientific investigation. Microfluidics is anticipated to contribute significantly to the advancement of biotechnology and to advancements in the fields of healthcare, pharmacology, and environmental sciences as technology develops.

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