ENHANCING VEGETABLE CROP BREEDING AND ADAPTATION TO CHANGING CLIMATES

Abstract

 The ability of the world to feed itself is under danger due to the global loss in agricultural output caused by climate change. Given that it is anticipated that by 2020, there will be more than 10 billion people on the planet, stakeholders and governments are concerned about food security. Crop improvement combining modern breeding techniques, efficient agronomic methods, advancements in microbiome applications, and using the natural variety in underutilised crops is a great step in the correct path for meeting future food needs. The breeding objectives concerning fruit quality, yield, and resilience to environment stress will be discussed, with a particular emphasis on how climate change could influence these goals. Next, we will discuss the utilization of traditional and molecular breeding strategies.

Keywords: Breeding, Climate change, Biotic stress, Abiotic stress tolerance, Molecular breeding, Genomics.

Authors

Shahaji R Hange

Department of Genetics and Plant Breeding Section, Vasantdada Sugar Institute, Manjari (Bk), Tal. Haveli, Maharashtra, India.

Shalaka R Sinhasane

Division of Horticulture, ICAR- Directorate of Onion and Garlic Research, Rajgurunagar, Pune, Maharashtra, India.

Ankush S Gadge

Forest College and Research Institute, Tamil Nadu Agricultural University, Mettupalayam, Coimbatore, Tamil Nadu, India.

Dhananjay V Shirsath

Division of Agricultural Entomology, ICAR -Directorate of Onion and Garlic Research, Rajgurunagar, Pune, Maharashtra, India

Pushpalatha M

Department of Entomology, Mahatma Phule Krishi Vidyapeeth, Rahuri, Maharashtra, India.

I. INTRODUCTION

Due to their potential contribution to a healthy diet for humans, especially for vegetarians who rely on them as a rich supply of vitamins, minerals, and dietary fibres, vegetable crops are referred to as protective foods. These crops' growth, productivity, and quality are threatened by a variety of biotic and abiotic stressors. In terms of reproduction, these crops might be classified as annual, biennial, or perennial. It is difficult to improve economic agricultural qualities using traditional breeding techniques [1]. According to [2], producing crops with a high yield and superior quality while using little inputs will eventually be extremely difficult for modern agriculture. Vegetable crops provide abundant nutrients like, minerals, vitamins, dietary fiber, and phytochemicals, making them protective foods essential for fulfilling people's vital nutritional requirements [3].

To lower the risk of cardiovascular diseases, a person should consume over 400 g of vegetables and fruits each day [4]. Vegetable crops, like all other food crops, are susceptible to a variety of biotic and abiotic challenges [5,6]. As a result, it is necessary to design crops of the next generation that can withstand severe environmental conditions [7]. To address the demands of a new era, turf vegetable breeding is continually evolving and developing new tactics. In recent years, the world has been primarily concerned with food quality to meet the demands of health-conscious consumers and food quantity to cater to the rapidly expanding global population. Additionally, challenges related to abiotic and biotic stresses have resulted in significant financial losses for farmers [8].

Due to climate change, we can expect a rise in global temperatures and $CO₂$ levels, along with rise in the frequency of unpredictable droughts, floods, and storms worldwide [9,10,11]. At the same time, the global population is rapidly expanding [12]. Crop yields are expected to be significantly impacted by rising temperatures [13], hence the speedy creation of new resistant varieties is necessary to mitigate this [14, 15].

Increasing crop yields is crucial, but if there is need to increase the areas where crops may be grown, we must create crop types that can withstand inferior soils, added variable precipitation, and/or salt [16]. In the future, crops may face the challenge of combating new sets of pests and pathogens, while also adapting to the direct abiotic burden of climate change and unfavourable growth conditions. These biotic pressures are thought to affect crop yields by 20–40% [17], and it is anticipated that many diseases' native geographic ranges may change as a result of climate change [18]. Although climate-tolerant varieties are created, this added (and currently somewhat unpredictable; [19] stress, new pests, and illnesses could cause a large decrease in production.

Climate prediction models forecast dramatic shifts in weather patterns that would lead to more frequent floods and droughts, a rise in global temperatures, and a reduction in the amount of fresh water available for agriculture. Therefore, preparing for future climates presents a significant difficulty. This chapter delves into the techniques used for breeding climate-resilient superior genotypes of vegetable crops, aiming to enhance agricultural productivity and address the challenges of future global food security.

II. CROPS RESPONSE TO CLIMATE CHANGE

The most probable scenario requiring the establishment of plant breeding targets includes:

- 1. Certain higher temperatures leading to reduced crop yield.
- 2. Certain rise in $CO₂$ concentration with both straight and unintended effects.
- 3. Highly probable growing occurrence of drought.
- 4. Likely upsurge in the areas affected by salinity.
- 5. Very probable rising incidence of biotic stress.

 Given this situation and the fact that crop breeding has been successful in enhancing the yield [20], it is possible that crop breeding has the ability to facilitate the development of innovative cultivars with improved traits, better suitable to adapt to climate change circumstances, by leveraging both traditional and genetic technologies [21]. These characteristics include resistance to salinity and water logging [22], pests and diseases that continue to result in crop losses [23], drought and heat stress resistance, and pest and disease resistance. Amongst most significant and typical goals of many breeding initiatives for each of the major food crops in almost all countries has historically been breeding for drought resistance [24,25]. The variations in phenology or improved responses to increasing $CO₂$ are potential opportunities for novel varieties with increased drought resistance. In terms of water, some studies have shown that key crop species (such as maize and soybeans) have undergone genetic alterations that have boosted their water-deficit tolerance [26, 27, 28], while it's possible that not all crops have undergone these adaptations. In general, there is currently insufficient knowledge regarding how genetically modified characteristics work in actual farming and forestry applications [29].

III. CHALLENGES, PRIORITIES, AND BREEDING OBJECTIVES

 Vegetable crops have a range of challenges that influence their breeding objectives. In line with the diverse range of growth conditions, breeders will concentrate on their key breeding objectives and use them either as-is or after processing. These objectives can be broken down into two groups: (1) adjustment to growth situations in terms of reaction to abiotic and biotic challenges; and (2) vegetable quality on both a sensory and dietary level. (3) Cold tolerance.

IV. APPROACHES TO DEVELOP CLIMATE-RESILIENT VEGETABLE CROP

- 1. The Abiotic Stress: Vegetables face numerous abiotic stresses, such as drought, temperature, salinity, and heat, which adversely affect their crop yield. Conventional breeding methods can mitigate pressures to some extent, but new cutting-edge technologies, like as CRISPR-Cas 9, have the potential to produce more resilient genotypes that can handle these stresses [30]. A significant stressor that slows the development and produce of vegetable crops is high temperatures.
	- Drought Tolerance: The majority of potato types have thin, shallow roots that are susceptible to a diversity of abiotic challenges, like excessive salt and dryness, which lowers tuber output and quality. Even brief bouts of drought stress have the potential to inflict significant harm and a sharp decline in tuber yield. Since drought was not

seen as a significant yield-limiting factor for potatoes for a very long time, research on drought tolerance in potatoes did not begin until the 1960s to 1980s. Over the past few years, the situation regarding drought in potato production has undergone a radical change, with a growing recognition of its significance and a notable interest in developing potato cultivars that can thrive in drought-prone areas [31].

Many scientists have tested the drought tolerance of different potato landraces. In Andean landraces, mainly in the species S. curtilobum (Juz. and Bukasov) in the cultivar families Stenotomum, Andigenum, and Chaucha, a many number of accessions merging drought tolerance through high irrigated production were discovered. S. chillonanum, S. okadae and S. jamesii were discovered by [32] through the screening of 44 accessions of wild species chosen based on their drought territories derived from geographic information system (GIS).

 Because of the fundamental and plastic nature of roots, they are typically implicated in both drought avoidance and tolerance during water shortages. Because of its tremendous plasticity, RSA can adapt quickly to environmental changes like a water shortage. According to [33], potato plants have significantly higher ABA concentrations in their xylem when water content in the substrate decreases.

 [34] Described the 83 tomato WRKY genes and showed how each one behaved differently to pathogen infection, salt, drought, heat, and cold stimuli. Some genes (SlWRKY3, SlWRKY3, and SlWRKY33) were shown to be impacted by a diversity of conditions, including salinity and drought stress, and they were recognised as possible study candidates. For a group of genes belonging to the ERFs family [35] and the Hsp20 gene family [36], the expression profiles of additional tomato stress-response genes were also studied. Single genes that are crucial for tomato tolerance to abiotic stress include SlJUB1, which enhances tolerance to drought, and ShDHN, MYB49, and SlWRKY39, which promote tolerance to a variety of stress conditions [37, 38, 39].

 Numerous investigations have concentrated on locating the genes in pepper that can withstand heat stress and testing them in transgenic model systems. For instance, CaHSL1, a protein kinase involved in shielding plants from hightemperature stress under high humidity, was described by [40]. [41] discovered a gene called BAX inhibitor-1 that confers transgenic plants through increased tolerance to a variety of stress stimuli and is linked to the regulation of programmed cell death. Despite the acknowledgment of genetic diversity for heat stress tolerance [4,43] and particular molecular investigations on the activity of heat shock proteins [44,45], research on the quantitative inheritance of heat tolerance in pepper is still lacking. Upcoming plant breeding initiatives are required to create cultivars that are better able to withstand the stress of high temperatures.

 Advanced cultivars with outstanding root characteristics, capable of absorbing water from deeper sections of the soil and under lower soil water strength, are being bred to enhance the utilization of soil water and contribute to the efficient use of water in vegetable production, whether from precipitation or irrigation.

• Salinity Tolerance: Salinity-sensitive potato leaves are brutally harmed by overhead watering with salinity-rich water. Toxic effects from chlorine and salt uptake by leaves can manifest as leaf burn around the edges. According to [46], salt stress had a deleterious impact on the cultivar Desiree's comparative water content, leaf stomata or conductance, and transpiration rate.

 The pH is raised by sodium carbonate, which is formed when the quantity of exchangeable sodium ions in the soil solution increases due to saline water. These alkaline circumstances make it harder for plants to get nutrients like phosphate, iron, zinc, and manganese. This destructive process is stopped in soils high in calcium carbonate, a fact that has been demonstrated in-vitro where extra calcium prevented salinity induced nuclear deterioration in root meristematic cells [47]. In a study cited as [48], it was found that adding 2% gypsum to salty soil enhanced the potato production in pots, improved their protein, potassium, and calcium content, and reduced their glycoalkaloid levels.

 The in vitro system was utilised to show that exogenously given proline offered some level of defense against salt stress and was deemed appropriate for testing salt tolerance [49]. [50] showed that salt-tolerant and sensitive potato cultivars had different antioxidant enzyme activities, indicating that the salt-tolerant varieties may be well protected compared to reactive oxygen species due to their capacity to increase antioxidant enzyme activity under salt stress. Using an in vitro microtuberization system, [51] discovered variations in salt sensitivity between two potato cultivars. According to [52], the effects of 5-aminolevulinic acid (ALA), a crucial precursor in the manufacture of porphyrins like chlorophyll and heine, encouraged potato microtuber formation and growth as well as improved protective activities against oxidative stressors.

 MYB49, ShDHN, and SlWRKY39 are single genes for multi-stress tolerance factors, and DREB1A and VP1.1 are single genes for salinity tolerance [43,38].

 The characterization of a few chosen tomato cultivars served as the basis for setting the threshold for saline tolerance described above. [53] observed a considerable genotypic heterogeneity in fresh-market tomato cultivars' responses to salinity. This demonstrates the potential for the crop to produce salt-tolerant cultivars.

 In Capsicum, there is a significant genotypic heterogeneity for tolerance to salt [53]. The sensitive types in the study accumulated noticeably more sodium ions in the shoot than in resistant varieties. According to [54], salt-tolerant cultivars had higher levels of enzymatic antioxidants and less relative water content loss.

 It has been discovered that a few mitigating strategies for dealing with high salinity are relatively helpful in the production of peppers. These cultural techniques include better nutrient management, the usage of protectants like catechin or glycine betaine, and better watering techniques. For example, calcium nutrition can be improved to lessen harm from sodium or chloride. Saline-tolerant grafted plants can be produced using rootstocks that have been found [55].

2. Biotic Stresses: Environmental circumstances are constantly changing, and by 2050, maintaining the production of food to feed the growing population will be difficult due to a lack of arable land [56]. A key strategy for addressing this problem is the creation of robust crops with high-stress tolerance.

 New pests and pathogens should enter production areas as a consequence of global climate change. Climate prediction models forecast abrupt changes in weather patterns that would lead to more frequent floods and droughts, a rise in global temperatures, and a reduction in the amount of fresh water available for agriculture. Therefore, strengthening plant resistance and tolerance to pests is a major task [57].

• Disease Resistance: At different stages of crop growth, crop plants are affected by many and distinct forms of diseases, which severely reduce production. There are several ways to avoid crop losses brought on by these dangerous diseases. Many diseases harm the pepper crop. Therefore, it's crucial to apply fungicides, miticides, and insecticides responsibly in order to effectively manage infections and pests, as well as to achieve the best possible production and fruit value. However, incorporating the usage of resistant types is a pest management strategy that is good for the environment. Every crop breeding programme now places a high priority on finding and using key genes for disease resistance in the crop. Breeders trying to create resistant types have a significant challenge as the disease races continuously evolve at various speeds to overcome host resistance. Pyramiding various resistance genes with various mechanisms of action collectively in one line is one solution to this issue. In pepper, polygenic resistance to Potato virus Y outperformed monogenic resistance, according to research by [58]. There is information available on pepper germplasm's high or moderate susceptibility to several of the major diseases, as well as the way the resistance trait is inherited.

Contrary to disease resistance features, little is known about the genetics of various pest resistance. Leaf position and ontogeny have an impact on thrips resistance, according to studies on the subject [59], and QTLs for this characteristic have been found in particular mapping populations [60]. Diterpenes and flavonoids may aid in thrips resistance, according to more recent metabolomic studies [61].

| Crop | Wild Species | Resistance Character transferred against | |
|----------------|---------------------|---|--|
| Okra | Abelmoschus caillei | YVMV (Bhendi yellow vein mosaic virus) | |
| Tomato | Solanum hirsutum | Fusarium wilt (Fusarium oxysporum f. sp. | |
| | | lycopersici) | |
| Potato | Solanum demissum | Late blight (Phytophthora infestans) | |
| Potato | Solanum demissum | Leaf roll (Potato leafroll virus) | |
| Brinjal | S. stenototum | Bacterial wilt (Pseudomonas solanacearum) | |
| Chilli | Capsicum chinense | Fruit rot (Colletotrichum capsici) | |
| Onion | Allium fistulosum | Purple blotch (Alternaria porri) | |
| French | P. flavescens | Rust (Uromyces appendiculatus) | |
| bean | | | |
| Cucumber | Cucumis Hardwiiki | Green-mottle mosaic (Cucumber green mottle | |
| | | mosaic virus) | |

Table 1: Resistance to Diseases in Various Vegetable Crops

• Pest Resistance: The global temperature is expected to rise, there will be more frequent droughts and floods, there will be less fresh water available for agriculture, and there will be dramatic changes in weather patterns. Therefore, strengthening plant resistance to pests and tolerance to them is a significant challenge.

 In wild species, a variety of pest resistances have been found. According to several research [62], insect resistance is attributed to glandular trichomes, glycoalkaloids, and other unidentified mechanisms. [63] tested 100 species of wild potatoes for insect resistance and found that the glycoalkaloid tomatine, thick hairs, and glandular trichomes were all associated with resistance. According to Jansky et al. [64], species with abundant glandular trichomes (S. polyadenium and S. tarijense) or high amounts of glycoalkaloids (S. chacoense) have proven resistance to the Colorado potato beetle. S. hougasii exhibited high levels of resistance to the Columbia root-knot nematode. Additionally, the Argentinian wild species S. vernei and S. acaule were found to possess resistance to cyst nematodes [65].

 Hypersensitive resistance and extreme resistance are two popular subtypes of single gene viral resistance in potato. It's common for viral strains to have unique hypersensitive resistance genes. When plants with these genes are exposed to viruses, they typically exhibit systemic necrosis or local necrotic lesions in the diseased tissue. In potatoes, a number of genes have been identified that code for hypersensitive resistance to potato viruses A, S, X, and Y [31]. The Solanaceae family exhibits a conserved location of genes imparting resistance to various diseases in addition to resistance to Phytophthora. The resistance gene hot spot on the long arm of chromosome 11 is home to three potato genes that code for resistance to PVY (Ryadg and Rysto) and PVA Naadg [66].

 Attacking plants, the onion thrips (Thrips tabaci L.) decreases photosynthetic activity, acts as a disease entry point, and spreads the IYSV [67]. To manage thrips, some onion cultivators employ biological and cultural controls, while the majority resort to insecticides. However, this practice may lead to the development of insecticide-resistant thrips and result in significant environmental problems. To address these concerns, efforts are underway to advance onion cultivars with inherent resistance to thrips. After analysing the germplasm from various countries, limited tolerant sources that can be exploited in breeding programs have been identified [68]. It has been shown that onion plants with glossy or semi glossy foliage are not more susceptible to thrips feeding damage [69]. It would be preferable to apply familybased assortment to increase genetic improvement because genetic studies reveal that thrips resistance is not extremely heritable [70].

• Root-Knot Nematodes Resistance: Temperatures over 30 °C impair plant defense mechanisms, frequently rendering important resistance genes useless. For instance, high temperatures inactivate the tomato Mi-1.2 resistance gene to the root-knot nematode and the Cf-4/Cf-9 genes to Cladosporium fulvum.

 Another intriguing strategy that requires little effort throughout the growing season and is good for the environment is the use of root-knot nematode-resistant Futuristic Trends in Agriculture Engineering & Food Sciences e-ISBN: 978-93-5747-368-2 IIP Series, Volume 3, Book 19, Part 2, Chapter 3 ENHANCING VEGETABLE CROP BREEDING AND ADAPTATION TO CHANGING CLIMATES

cultivars [71]. The study of pepper genetic resources against diseases, nematodes, and arthropods [72] lists excess than 40 accessions as tolerant or very resistant to several root-knot nematode species. In the Solanaceae family, resistance to root-knot nematodes is primarily dominant and regulated by a small number of crucial genes. Nine diverse dominant genes have been found to confer resistance to root-knot nematodes in several populations of peppers. Out of these, the resistance of M. incognita, M. javanica, M. arenaria, and M. haplanaria to N, Me1, and Me3 (= $Me7$) was assessed [73].

| Crop | Wild Species | The resistant Character transferred against |
|----------------|------------------------|--|
| Potato | Solanum verni | Nematode (Meloidogyne spp.) |
| Cucurbits | Cucumis trigonus | Fruit fly (Bactrocera cucurbitae) |
| Brinjal | S. incanum | Shoot & fruit borer (Leucinodes orbonalis) |
| Tomato | Solanum hirsutum | White fly (Bemicia tabaci) |
| Okra | Abelmoschus manihot | Shoot & fruit borer (Earias vitella, E. insulana) |

Table 2: Resistance in Various Vegetable Crops against Insects

Figure 1: A genomics-based breeding strategy for Allium [74]

Futuristic Trends in Agriculture Engineering & Food Sciences e-ISBN: 978-93-5747-368-2 IIP Series, Volume 3, Book 19, Part 2, Chapter 3 ENHANCING VEGETABLE CROP BREEDING AND ADAPTATION TO CHANGING CLIMATES

| Loci | Genotypes | Phenotypes | Clarifications | |
|------|------------------|----------------------|--|--|
| Foc1 | foc1foc1 | Susceptible | Fusarium basal plates rot tolerance locus 1 is partially dominating with Foc2 and exhibits additive resistance. | |
| | Foc1foc1 | Partial resistant | | |
| | Foc1Foc1 | Resistant | | |
| Foc2 | foc2foc2 | Susceptible | Resistance locus 2 for Fusarium basal plate rot. Foc1 is somewhat dominant and exhibits additive resistance. | |
| | Foc2foc2 | Partial resistant | | |
| | Foc2Foc2 | Resistant | | |
| Pr1 | prlprl | Resistant | | |
| | Prlprl | | Pink root tolerance locus. It has been shown that other modifier genes | |
| | Pr1Pr1 | Susceptible | may be implicated in resistance. | |
| Pd1 | pd1pd1 | Susceptible | | |
| | Pd1pd1 | Resistant | Dominant gene introduced from A. roylei for downy mildew tolerance locus 1. | |
| | Pd1Pd1 | | | |
| Oz | OZOZ | Susceptible | Ozone resistance gene. Individuals that have a recessive allele at the Oz gene are more vulnerable to ozone stress. | |
| | Ozoz | Tolerant | | |
| | OzOz | | | |

Table 3: Onion Morphological Marker Loci for Biotic and Abiotic Factors [75]

3. Vegetable Quality Improvement: Advanced post-harvest technologies are required for fruit and vegetable (F&V) storage constancy and prolonged shelf life because they are highly perishable food products [76]. T1 homozygous plants with long shelf lives were produced in tomato by replacing the allele of ALC with the alc gene via the homologydirected repair (HDR) mechanism. In many food applications, potato starch purity is crucial. In potatoes, full knockouts of the genes for the starch-branching enzymes (SBEs) SBE1 and SBE2 as well as the starch synthase gene (SS6) have been reported to improve starch quality [77, 78]. Similar to this, enzymatic browning in brinjal was associated with the three-polyphenol oxidase (PPO) genes SmelPPO4, SmelPPO5, and SmelPPO6. These three target PPO genes have been disabled utilising CRISPR-Cas9-based mutagenesis to prevent the browning of fruit flesh. [79].

 In particular, beta-carotene, the precursor to retinol (vitamin A), and lutein and zeaxanthin, which are significant antioxidants for eye health, are nutritionally valuable pepper carotenoids. Numerous research has examined entire carotenoid levels and composition in various pepper cultivars due to the nutritional importance of carotenoids. According to Brewster [80] and [81], the first-class of onion cultivars is attributed by the bulb colour (anthocyanin and flavonoid content), firmness, number of scales, number of developing points, neck thickness, Total soluble solids (TSS), pungency, and antioxidants.

| Crop | Wild Species | Character transferred |
|--------|----------------------|------------------------------|
| Tomato | Solanum hirsutum | Carotenoid content |
| Potato | Solanum acule | Starch content |
| Chilli | Capsicum frutescence | High capsaicin |
| Melons | Cucumis melo var. | Thick rind and good keeping |
| | cantaloupensis | quality |
| nion | Allium kurrat | Leaf flavour |

Table 4: Improvement of quality in various vegetable crops

4. Cold Tolerance: Cold is a main abiotic stress affecting agricultural productivity globally, impacting the growth and development of agronomic species. Given that approximately 64% of the earth's land area experiences a usual minimum temperature below 0° C and about 48% below 10 °C, comprehending the mechanism of frost damage and developing cold-tolerant varieties becomes highly essential. To boost output and stability in cold conditions that are getting worse with climate change, potato crops must adapt. In order to defend themselves from harm caused by below-freezing temperatures, plants have evolved two strategies. First, acclimated xylem parenchyma cells of moderately resistant woody plants are typically linked to supercooling, a low-temperature tolerance mechanism. Acclimatization is the second and most typical low-temperature response mechanism. According to [82], acclimatization is a slow progression that causes changes in almost every quantifiable physiological, morphological, and biochemical characteristic of the plant. The complicated interactions between genetics and environment are what cause these alterations.

 Peppers grown in tropical climates are vulnerable to frost. Numerous studies have examined how chilling stress negatively affects the various metabolic systems in peppers

[83]. The plants cultivated at the lesser night temperature exhibited superior chilling tolerance after being exposed for 4 nights at 6° C [84]. Low nighttime temperatures (14 °C or lower) have a detrimental effect on the flower's female reproductive system, pollen viability, and pollen production [85]. The work by [86] demonstrated the function of reactive oxygen species in the harm caused by cold stress. Low-temperature tolerance has a known genetic component [87], although no selections for this feature have been documented. Peppers are vulnerable to cold damage when the harvested fruit is left in the cold (7 \degree C) for an extended period of time. A sign of cooling harm is surface pitting. It is likely to improve the cold storability of food by combining low-temperature conditioning with the use of methyl jasmonate and UV-C treatments [88, 89]. A proteomic investigation found that the stress of freezing caused bell peppers to create more ethylene, modify the composition of their sugars and organic acids, and drastically alter the proteins involved in the regulation of redox homeostasis and the breakdown of carbohydrates.

- 5. Herbicide Resistance: Selective herbicides are commonly employed during cultivation to control weed growth and development, as weeds are a substantial stress factor impacting vegetable produce and its quality. To confer herbicide resistance in plants, the herbicide target gene acetolactate synthase (ALS) has been modified using CRISPR-Cas9 technology in crops such as watermelon, tomato, and potato [90,91].
- 6. Nutrient Use Efficiency: When it comes to boosting crop harvests in the face of escalating climate change and global warming, nutrient-efficient plants are crucial. At least 60% of the arable land on the planet has mineral or elemental toxicity issues. "The plant development, physiological action, yield, or harvested yield per unit of nutrient" is how the term "nutrient use efficiency" (NUE) is defined. The balance of nutrients and biological activity are the two main factors that affect a plant's productivity [31].

V. FUTURE PROSPECTS

To ensure global food security amid the continuing and projected climate change, which includes increasing temperatures and random climate events across large regions of the world, it is crucial to breed crop plants with enhanced yield potential and enhanced tolerance to such conditions. The goal of climate-resilient agriculture can only be accomplished with upgraded plant types that can withstand diseases and pests while effectively using less resources and demonstrating stable yields in stressful conditions in the near future. Research focus is essential for currently underutilised crop species if they are to contribute to climatic resilience. To address crop plants' sensitivity to climate change, smart breeding relies heavily on creating huge breeding populations, effective high throughput phenotyping, large data management technologies, and ensuing molecular approaches.

To prepare for the challenges of a warmer climate, characterized by increased vulnerability to droughts, floods, and potential threats from novel pests and viruses, it is crucial for current research to identify crop varieties and Crop Wild Relatives exhibiting adaptive tolerance to these stresses. Despite these efforts being carried out for vegetable crops, it seems that progress has been slow and, until recently, lagged behind developments in other crops.

VI. CONCLUSION

The continuing development of innovative crops and new kinds of plant-based foods will be essential for future food production. To help with climate adaptation, research will need to be focused on crop species that are now underutilised. One of the major and difficult areas of agriculture is vegetable breeding. Breeding in agricultural crops benefits from both traditional and molecular approaches. In order to prepare for a warmer world that will be more vulnerable to droughts and floods as well as the prospect of novel pests and viruses becoming a threat, it is imperative that current research find cultivars of crops with adaptive resilience to these stresses. Even if this is being done for the vegetable crop, it seems like progress is being made slowly and has lagged behind other crops up until very recently. Despite not being among the most essential vegetables internationally, many nations and cultures rely heavily on vegetables in their diets, therefore any reduction in production could have a negative impact on these populations. In this chapter, I've emphasised what is already known about vegetable crop tolerances that might be useful in a future climate, as well as several crucial research directions that ought to be given top priority.

Plant breeders can potentially design and produce the appropriate genotype by selecting the finest allele for each gene, enabling enhanced performance in the marked environment. The path to this approach is swiftly being defined by functional genomics.

REFERENCES

- [1] R. Devi, S. Chauhan, and T.S. Dhillon, ''Genome editing for vegetable crop improvement: Challenges and future prospects,''Frontiers in Genetics, 13, 1037091, 2022.
- [2] D. Tilman, C. Balzer, J. Hill, and B.L. Befort, ''Global food demand and the sustainable intensification of agriculture,'' Proc. Natl. Acad. Sci. U. S. A. 108, 20260–20264, 2011.
- [3] J. Dias, "Nutritional quality and health benefits of vegetables: A review," Food Nutr. Sci.Food Nutr. Sci. 3, 1354–1374, 2012.
- [4] L. A. Bazzano, J. He, L. G. Ogden, C.M. Loria, S. Vupputuri, and L. Myers, ''Fruit and vegetable intake and risk of cardiovascular disease in US adults: The first national health and nutrition examination survey epidemiologic follow-up study,'' Am. J. Clin. Nutr. 76, 93–99, 2002.
- [5] D. Jaganathan, K. Ramasamy, G. Sellamuthu, S. Jayabalan, and G. Venkataraman, ''CRISPR for crop improvement: An update review,'' Front. Plant Sci. 9, 985,2018.
- [6] M. Boscaiu, and A. Fita, ''Physiological and molecular characterization of crop resistance to abiotic stresses,'' Agronomy 10, 1308, 2020.
- [7] S.G. Karkute, A. K. Singh, O. P. Gupta, P. M. Singh, and B. Singh, ''CRISPR/Cas9 Mediated Genome Engineering for Improvement of Horticultural Crops,'' Front Plant Sci. 22, 1635,2017.
- [8] J.Von Braun, M.W. Rose grant, R. Pandya Lorch, M.J. Cohen, S.A. Cline, M.A Brown, and M.S. Bos ''New Risks and Opportunities for Food Security Scenario Analyses for IFPRI: Washington,'' DC, USA; 2015-2050, 2005.
- [9] D. Coumou, and S. Rahmstorf, ''A decade of weather extremes,'' Nat Clim Change 2:491, 2012.
- [10] K.E. Trenberth, A. Dai, G. van der Schrier, P.D. Jones, J. Barichivich, K.R. Briffa, and J. Sheffield ''Global warming and changes in drought,'' Nat Clim Change 4:17, 2013.
- [11] G.M. Poppy, S. Chiotha, F. Eigenbrod, C.A. Harvey, M. Honzak, M.D. Hudson, A. Jarvis, N.J. Madise, K. Schreckenberg, C.M. Shackleton, F. Villa, and T.P. Dawson. ''Food security in a perfect storm: using the ecosystem services framework to increase understanding,'' Phil Trans Ro Soc B-Biol Sci 369, 2014
- [12] H.C. Godfray, J.R. Beddington, I.R. Crute, L. Haddad, D. Lawrence, J.F. Muir, J.Pretty, S, Robinson, S.M. Thomas, and C. Toulmin, ''Food security: the challenge of feeding 9 billion people,'' Science 327(5967):812–818, 2010.
- [13] C. Zhao, B.Liu, S. Piao, X. Wang, D.B. Lobell, Y. Huang, M. Huang, Y. Yao, S. Bassu, P. J.L. Ciais, Durand, J. Elliott, F. Ewert, I.A. Janssens, T. Li, E. Lin, Q. Liu, P. Martre, C. Müller, S. Peng, J. Peñuelas, A.C. Ruane, D.Wallach, T. Wang, D. Wu, Z. Liu, Y. Zhu, Z.Zhu, and S. Asseng, ''Temperature increase

Futuristic Trends in Agriculture Engineering & Food Sciences e-ISBN: 978-93-5747-368-2 IIP Series, Volume 3, Book 19, Part 2, Chapter 3 ENHANCING VEGETABLE CROP BREEDING AND ADAPTATION TO CHANGING CLIMATES

reduces global yields of major crops in four independent estimates,'' Proc Natl Acad Sci USA 114(35):9326–9331, (2017)

- [14] A.J. Challinor, A.K. Koehler, J Ramirez-Villegas, S, Whitfield, and B. Das. ''Current warming will reduce yields unless maize breeding and seed systems adapt immediately,'' Nat Clim Change 6:954 , 2016.
- [15] [G.N. Atlin, J.E. Cairns, and B. Das, ''Rapid breeding and varietal replacement are critical to adaptation of cropping systems in the developing world to climate change,'' Glob Food Secur 12:31-37, 2017
- [16] I. Ahuja, R.C. de Vos, A.M. Bones, and R.D. Hall, ''Plant molecular stress responses face climate change,'' Trends Plant Sci 15(12):664–674, 2010
- [17] E.C.Oerke, H.W Dehne, F. Schonbeck, and A. Weber, ''Crop production and crop protection: estimated losses in major food and cash crops,'' Elsevier Science, Amsterdam,1994
- [18] S. Savary, A. Ficke, J.N. Aubertot, and C. Hollier, ''Crop losses due to diseases and their implications for global food production losses and food security,'' Food Secur 4(4):519–537, 2012.
- [19] M. Donatelli, R.D. Magarey, S. Bregaglio, L .Willocquet, J.P. Whish, and S. Savary, ''Modelling the impacts of pests and diseases on agricultural systems,'' Agri Syst 155:213–224, 2017
- [20] J. Dixon, L. Nalley, P. Kosina, R. La Rovere, J. Hellin, and P. Aquino, ''Adoption and economic impact of improved wheat varieties in the developing world,'' Journal of Agricultural Science, Cambridge 144, 489-502,2006.
- [21] D. Z. Habash, Z. Kehel, and M. Nachit, ''Genomic approaches for designing durum wheat ready for climate change with a focus on drought,''Journal of Experimental Botany 60, 2805–2815, 2009.
- [22] M.O. Humphreys, ''Genetic improvement of forage crops past, present and future,'' Journal of Agricultural Science, Cambridge 143, 441-448, 2005.
- [23] E.C. Oerke, ''Crop losses to pests,'' Journal of Agricultural Science, Cambridge 144, 3143, 2006.
- [24] S. Ceccarelli, S. Grando, M. Baum, and S.M. Udupa, ''Breeding for drought resistance in a changing climate,'' In Challenges and Strategies for Dryland Agriculture (Eds S. C. Rao & J. Ryan), pp. 167–190. 2004. CSSA Special Publication No. 32. Madison, WI: ASA and CSSA.
- [25] S. Ceccarelli, S. Grando, and M. Baum, "Participatory plant breeding in water-limited environments," Experimental Agriculture 43, 411–435, 2007.
- [26] P. M. Drennen, M. Smith, D. Goldsworthy, and J.Van Staten, ''The occurrence of trahaolose in the leaves of the desiccation tolerant angiosperm Myronthamnus flabellifoliius Welw,'' Journal of Plant Physiology 142, 493-496, 1993.
- [27] P. B. Kishor, Z. Hong, G. Miao, C. Hu, and D. Verma, ''Overexpression of Δ1-pyrroline-5-carboxylase synthase increases praline production and confers osmotolerance in transgenic plants,'' Journal of Plant Physiology 108, 1387-1394, 1995.
- [28] N. Cheikh, P.W. Miller, and G. Kishore, ''Role of biotechnology in crop productivity in a changing environment. In Climate Change and Global Crop Productivity (Eds K. R. Reddy & H. F. Hodges) ,'' pp. 425–436, 2000. New York, NY: CAB International.
- [29] T.R. Sinclair, and L.C. Purcell, ''Is a physiological perspective relevant in a 'genocentric' age? ,'' Journal of Experimental Botany 56, 2777–2782, 2005.
- [30] E. Haque, H. Taniguchi, M. Hassan, P. Bhowmik, M.R. Karim, and M. Smiech, ''Application of CRISPR/Cas9 genome editing technology for the improvement of crops cultivated in tropical climates: Recent progress, prospects, and challenges,'' Front. Plt. Sci. 9, 617,2018).
- [31] J.K. Tiwari, C. Challam, S.K. Chakrabarti, and S.E. Feingold, ''Climate-smart potato: an integrated breeding, genomics, and phenomics approach,'' Genomic Designing of Climate-Smart Vegetable Crops, 1-46, 2020.
- [32] K.N. Watanabe, A. Kikuchi, T. Shimazaki, and M. Asahina, ''Salt and drought stress tolerances in transgenic potatoes and wild species,'' Potato Res 54:319-324, 2011
- [33] F. Liu, C.R. Jensen, A. Shahanzari, M.N. Andersen, and S.E. Jacobsen, ''ABA regulated stomatal control and photosynthetic water use efficiency of potato (Solanum tuberosum L.) during progressive soil drying,'' Plant Sci 168(3):831-836, 2005.
- [34] Y.L. Bai, C. Kissoudis, Z. Yan, R.G. Visser, and G, van der Linden, ''Plant behaviour under combined stress: tomato responses to combined salinity and pathogen stress,'' Plant J 93:781–793, 2018.
- [35] I. Klay, S. Gouia, M. Liu, I. Mila, H. Khoudi, and A. Bernadac et al, ''Ethylene Response Factors (ERF) are differentially regulated by different abiotic stress types in tomato plants,'' Plant Sci 274:137-145-2018.
- [36] Y. Yu, T. Jia, and X. Chen, ''The 'how' and 'where' of plant microRNAs,'' New Phytol 216:1002–1017, 2017.
- [37] H. Liu, C.Yu, H. Li, B. Ouyang, T. Wang, and J. Zhang et al, ''Overexpression of SHDHN, a dehydrin gene from Solanum habrochaites enhances tolerance to multiple abiotic stresses in tomato,'' Plant Sci 231:198-211, 2015.
- [38] X. Sun, Y. Gao, H. Li, S. Yang, and Y. Liu. ''Over-expression of SlWRKY39 leads to enhanced resistance to multiple stress factors in tomato,'' J Plant Biol 58:52–60, 2015
- [39] J. Cui, N. Jiang, X. Zhou, X. Hou, G. Yang, J. Meng, and Y. Luan, ''Tomato MYB49 enhances resistance to Phytophthora infestans and tolerance to water deficit and salt stress,''Planta 248:1487-1503, 2018.
- [40] D.Y. Guan, F. Yang, X.Q. Xia, Y.Y. Shi, S.Yang, and W .Cheng, S.L. He, ''CaHSL1 acts as a positive regulator of pepper thermotolerance under high humidity and is transcriptionally modulated by CaWRKY40,''Front Plant Sci 9:1802,2018.
- [41] M. Isbat, N.Zeba, S.R. Kim, and C.B. Hong, ''A BAX inhibitor-1 gene inCapsicum annuum is induced under various abiotic stresses and endows multi-tolerance in transgenic tobacco,'' J Plant Physiol 166:1685-1693, 2009.
- [42] K.R. Reddy, and V.G. Kakani, ''Screening Capsicum species of different origins for high temperature tolerance by in vitro pollen germination and pollen tube length,'' Sci Hort 112:130–135, 2007
- [43] M. Guo, J.H Liu, J. P Lu, Y.F. Zhai, H. Wang, Z.H. Gong, S. B. Wang, and M.H. Lu, ''Genome-wide analysis of the CaHsp20 gene family in pepper: comprehensive sequence and expression profile analysis under heat stress,'' Front Plant Sci 6:806, 2015.
- [44] M. Guo, J.H. Liu, X. Ma, Y.F. Zhai, Z.H. Gong, and M.H. Lu, ''Genome-wide analysis of the Hsp70 family genes in pepper (Capsicum annuum L.) and functional identification of CaHsp70-2 involvement in heat stress,''Plant Sci 252:246-256, 2016.
- [45] J.T. Sun, G.X. Cheng, L.J. Huang, S. Liu, M. Ali, A. Khan, Q.H. Yu, S.B. Yang, D.X. Luo, and Z.H. Gong, ''Modified expression of a heat shock protein gene, CaHSP22.0, results in high sensitivity to heat and salt stress in pepper (Capsicum annuum L.),'' Sci Hort 249:364-373, 2019
- [46] F. Fidalgo, A. Santos, I. Santos, and R. Salema, ''Effects of long-term salt stress on antioxidant defence systems, leaf water relations and chloroplast ultrastructure of potato plants,''Ann Appl Biol 145:185-192, 2004.
- [47] K.V. Richardson, A.C. Wetten, and P.D. Caligari, ''Cell and nuclear degradation in root meristems following exposure of potatoes (Solanum tuberosum L.) to salinity,'' Potato Res 44:389-399, 2001
- [48] Z. Abdullah, and R. Ahmad, ''Salt tolerance of Solanum tuberosum L growing on saline soils amended with gypsum,'' Z Fuer Acker-Und PflanzBau 151:409-416, 1982
- [49] P.V. Prasad, and S.D. Potluri, ''Influence of proline and hydroxyproline on salt-stressed axillary bud cultures of two varieties of potato (Solanum tuberosum) ,'' Vitro Cell Devel Biol Plant 32:47–50, 1996
- [50] H. Rahnama, and H. Ebrahimzadeh, ''The effect of NaCl on antioxidant enzyme activities in potato seedlings,'' Biol Plant 49:93-97, 2005.
- [51] Z.J. Zhang, B.Z. Mao, H.Z. Li, W.J. Zhou, Y. Takeuchi, and K. Yoneyama, ''Effect of salinity on physiological characteristics, yield and quality of microtubers in vitro in potato,'' Acta Physiol Plant 27:481–489, 2005
- [52] Z.J. Zhang, H.Z. Li, W.J. Zhou, Y. Takeuchi, and K. Yoneyama, ''Effect of 5-aminolevulinic acid on development and salt tolerance of potato (Solanum tuberosum L.) microtubes in vitro,'' Plant Growth Regul 49:27–34, 2006.
- [53] H. Aktas, K. Abak, and I. Cakmak, ''Genotypic variation in the response of pepper to salinity,'' Sci Hort 110:260-266, 2006.
- [54] H. Aktas, K. Abak, and S. Eker, ''Anti-oxidative responses of salt-tolerant and salt-sensitive pepper (Capsicum annuum L.) genotypes grown under salt stress,'' J Hort Sci Biotechnol 87:360-366, 2012.
- [55] F. Guiffrida, C. Cassaniti, and C. Leonardi (2013) ''The influence of rootstock on growth and ion concentrations in pepper (Capsicum annuum L.) under saline conditions,'' J Hort Sci Biotechnol 88:110– 116
- [56] L. T. Hickey, H. Robinson, S. A. Jackson, S. C. Leal-Bertioli, and M. Tester, ''Breeding crops to feed 10 billion,''Nat. Biotechnol. 37, 744–754, 2019.
- [57] M. Causse, J. Zhao, I. Diouf, J. Wang, V. Lefebvre, B. Caromel, and N. Bertin,. ''Genomic designing for climate-smart tomato,'' Genomic designing of climate-smart vegetable crops, 47-159,2020.
- [58] A. Palloix, V. Ayme, and B. Moury, ''Durability of plant major resistance genes to pathogens depends on the genetic background, experimental evidence and consequences for breeding strategies,'' New Phytol 183:190–199, 2009.
- [59] I.G. Visschers, J.L. Peters, J.A. Van de Vondervoort, R.H. Hoogveld, and N.M. Van Dam, ''Thrips resistance screening is coming of age: leaf position and ontogeny are important determinants of leaf-based resistance in pepper,'' Front Plant Sci 10:510, 2019.
- [60] A. Maharijaya, B.Vosman, G. Steenhuis-Broers, K. Pelgrom, A. Purwito, R.G. Visser, and R.E. Voorrips, ''QTL mapping of thrips resistance in pepper,'' Theor Appl Genet 128:1945,1956
- [61] A. Maharijaya, B. Vosman, K. Pelgrom, Y. Wahyuni, R.C. De Vos, and R.E. Voorrips, ''Genetic variation in phytochemicals in leaves of pepper (Capsicum) in relation to thrips resistance,'' Arthropod Plant Interact 13:1-9,2019.
- [62] Y, Pelletier, F.G. Horgan, and J. Pompon, ''Potato resistance against insect herbivores: Resources and opportunities,'' In: Giordanengo P, Vincent C, Alyokhin A (eds) Insect Pests of Potato, Global Perspectives on Biology and Management. Academic Press, Oxford, UK, pp 439-462,2013.
- [63] K.L. Flanders, J.G. Hawkes, E.B. Radcliffe, and F.I. Lauer, ''Insect resistance in potatoes: sources, evolutionary relationships, morphological and chemical defenses, and ecogeographical associations,''Euphytica 61:83-111, 1992.
- [64] S.H. Jansky, R. Simon, and D.M. Spooner, ''A test of taxonomic predictivity: resistance to the Colorado potato beetle in wild relatives of cultivated potato,'' J Econ Entomol 102:422–431,2009.
- [65] J.G. Hawkes, ''Origins of cultivated potatoes and species relationships,'' In: Bradshaw JE,Mackay GR (eds) Potato Genetics. CAB International, Wallingford, pp 3–42,1994.
- [66] G. Brigneti, J. Garcia-Mas, and D.C. Baulcombe, ''Molecular mapping of the potato virus Y resistance gene Rysto in potato,'' Theor Appl Genet 94:198-203,1997.
- [67] S.H. Bag, C.S. Schwartz, M.J. Cramer, Havey and H.R. Pappu. ''Iris yellow spot virus (TospovirusBunyaviridae): from obscurity to research priority,'' Mol. Plant Pathol 16: 224-237, 2014
- [68] J.J. Diaz-Montano, M. Onatno, B.A. Fuchs, J, Nault, Fail, and A.M. Shelton, ''Onion thrips (Thysanoptera: Thripidae): a global pest of increasing concern in onion,'' J. Econ. Entomol 104: 1-13, 2011.
- [69] S. Damon and M.J. Havey, ''Quantitative trait loci controlling amounts and types of epicuticular waxes in onion,'' J. Am. Soc. Hortic. Sci 139: 597-602, 2014.
- [70] B.K. Hamilton, L.M. Pike, A.N. Sparks, D.A. Bender, R.W. Jones, J. Candeia and G.D. Franca, ''Heritability of thrips resistance in the 'IPA-3' onion cultivar in South Texas,'' Euphytica 109: 117- 122,1999.
- [71] V.L. Fuller, C.J. Lilley, and P.E. Urwin, ''Nematode resistance,'' New Phytol 180:27–44
- [72] B. Sarath Babu, S.R. Pandravada, R.D.V.J Prasada Rao, K. Anitha, S.K. Chakrabarty, and KS Varaprasad, ''Global sources of pepper genetic resources against arthropods, nematodes and pathogens,'' Crop Protec 30:389-400, 2008.
- [73] A. Hajihassani, W.B. Rutter, and X. Luo, ''Resistant pepper carrying N, Me1, and Me3 have different effects on penetration and reproduction of four major Meloidogyne species,'' J Nematol. jofnern-2019- 020, 2019.
- [74] A.F. Gökçe ''Classical genetics on gene mapping. In: The Allium Genomes. (Eds.) Shigyo, M., Khar, A., Abdelrahman, M. Compendium of Plant Genomes, '' Springer, Cham. pp. 53-66. DOI: https://doi.org/10.1007/978-3-319-95825-5_4, 2018.
- [75] J.S. Khosa, J. McCallum, A.S. Dhatt, R.C. Macknight, ''Enhancing onion breeding using molecular tools,'' Plant Breeding 135(1), 9-20. DOI: http://dx.doi.org/10.1111/pbr.12330, 2016.
- [76] M. J. Gallagher, and P. V. Mahajan, "Stability and shelf life of fruit and vegetables," in Food and beverage stability and shelf-life,'' Editors Kilcast and Subramaniam (Cambridge, UK: Woodhead Publishing Ltd), 641–656, 2011.
- [77] M. Andersson, H. Turesson, N. Olsson, A. Falt, S. P. Ohlsson, and M. N. Gonzalez, ''Genome editing in potato via CRISPR-Cas9 ribonucleoprotein delivery,'' Physiol. Plant. 164, 378–384, 2018.
- [78] X. Zhao, S. Jayarathna, H. Turesson, A.S. Falt, G. Nestor, and M. N. Gonzalez et al, ''Amylose starch with no detectable branching developed through DNA-free CRISPR-Cas9 mediated mutagenesis of two starch branching enzymes in potato,'' Sci. Rep. 11, 4311, 2021.
- [79] A. Maioli, S. Gianoglio, A. Moglia, A. Acquadro, D. Valentino, and A.M Milani, ''Simultaneous CRISPR/Cas9 editing of three PPO genes reduces fruit flesh browning in Solanum melongena L,'' Front. Plant Sci. 11, 607161, 2020.
- [80] J.L. Brewster, ''Onions and Other Vegetable Alliums,'' CABI, Oxfordshire, UK, 2008.
- [81] I.L. Goldman, ''Molecular breeding of healthy vegetables,'' EMBO Rep 12, 96-102, 2011
- [82] D. Takahashi, B. Li, T. Nakayama, Y. Kawamura, and M. Uemura, ''Plant plasma membrane proteomics for improving cold tolerance,'' Front Plant Sci 4:90, 2013.
- [83] L.M. Tijskens, E.C. Otma, and O. Van Kooten, ''Photosystem II quantum yield as a measure of radical scavengers in chilling injury in cucumber fruits and bell peppers,'' Planta 194:478-486,1994
- [84] J.A. Mercado, M.S. Reid, V. Valpuesta, and M.A. Quesada, ''Metabolic changes and susceptibility to chilling stress in Capsicum annuum plants grown at suboptimal temperature,'' Aust J Plant Physiol 24:759–767, 1997.
- [85] N. Cruz-Huerta, J.G. Williamson, and R.L. Darnell, ''Low night temperature increases ovary size in sweet pepper cultivars,'' HortScience 46:396–401, 2011.
- [86] M. Airaki, M. Leterrier, R.M. Mateos, R. Valderrama, M. Chaki, J.B. Barroso, L.A. Rio, J.M. Palma, and F.I. Corpas, ''Metabolism of reactive oxygen species and reactive nitrogen species in pepper (Capsicum annuum L.) plants under low temperature stress,'' Plant, Cell Environ 35:281-295, 2012
- [87] E. Pressman, H. Moshkovitch, K. Rosenfeld, R. Shaked, B. Gamliel, and B. Aloni, ''Influence of low night temperature on sweet pepper flower quality and the effect of repeated pollinations, with viable pollen, on fruit setting,'' J Hort Sci BioTechnol 73:131–136, 1998.
- [88] P. Wang, Q. Lu, Y. Ai, Y. Wang, T. Li, L. Wu, J. Liu, Q. Cheng, L. Sun, and H. Shen, ''Candidate gene selection for cytoplasmic male sterility in pepper (Capsicum annuum L.) through whole mitochondrial genome sequencing,'' Int J Mol Sci 20:578, 2019.
- [89] A.R. Vicente, C. Pineda, L. Lemoine, P.M. Civello, G.A. Martinez, and A.R. Chaves, ''UV-C treatments reduce decay, retain quality and alleviate chilling injury in pepper,'' Postharvest Biol Technol 35:69-78, 2005.
- [90] B. Danilo, L. Perrot, K. Mara, E. Botton, F. Nogue, and M. Mazier, ''Efficient and transgene-free gene targeting using Agrobacterium-mediated delivery of the CRISPR/Cas9 system in tomato,'' Plant Cell Rep. 38, 459–462, 2019.
- [91] Z. Li, Z. Liu, B. Xing, A. Moon, B. P. Koellhoffer, and J. P. L. Huang, et al, ''Cas9-Guide RNA directed genome editing in soybean. Plant Physiol,'' 169, 960-970, 2015.