

1 **Indoleamine 2, 3-dioxygenase (IDO) mediated Th1-Th2 shift, modulates the**
2 **maternal immune system and decipher pregnancy outcomes in dairy cattle**

3
4 Sunil Kumar Mohapatra^{1*}, Sameni Deepika¹, Dheeraj Chaudhary², Bibhudatta S.
5 K. Panda², Rajeev Kapila¹ and Ajay Kumar Dang²

6 ¹*Department of Animal Biochemistry, ICAR-National Dairy Research Institute, Karnal, Haryana*
7 *132001, India*

8 ²*Lactation and Immuno-Physiology Laboratory, ICAR-National Dairy Research Institute,*
9 *Karnal, Haryana 132001, India*

10
11

12 **Abstract:**

13 **Background**

14 In the current scenario, a decrease in the production potential of dairy cattle particularly
15 due to the reduced reproductive efficiency is a major concern for the farmers and field
16 veterinarians. Therefore, enhancement of reproductive efficiency is the need of the hour to raise
17 the production potential of dairy cattle. Reproductive failure in terms of infertility, subfertility
18 and dreadful embryonic mortality are some of the major substantial factors responsible for
19 lowering the reproductive efficiency in dairy cattle. Such serious issues can be tackled and
20 minimized by expanding the knowledge in the field of bovine pregnancy and unravel different
21 mechanisms of pregnancy establishment in bovines. This will enable and motivate researchers to
22 discover some possible and suitable interventions around these mechanisms in order to increase
23 the pregnancy outcomes in dairy cattle. Although, there are many mechanisms which have been
24 delineated so far for the pregnancy establishment in bovine but still many areas remain untapped
25 which needs to be taken into limelight. Exploring different mechanisms in early bovine
26 pregnancy may help to improve the reproductive efficiency in bovines and thereby production
27 potential in the dairy sector.

28 **Major advances**

29 This chapter provides an insight into different immune tolerance mechanisms associated with the
30 implantation of the blastocyst and early pregnancy establishment in bovines. During the early

31 pregnancy period (where a majority of embryonic loss occurs), it is imperative for the conceptus
32 not only to prevent the corpus luteal regression but also to modify the maternal immune system
33 from immune rejection. In this regard, interferon tau (IFN τ), a conceptus secreted factor, known
34 to have anti-luteolytic and immunomodulatory properties and is the key pregnancy recognition
35 agent in bovines, plays a preliminary and vital role in the establishment of bovine pregnancy.
36 Whether it is by paracrine or by endocrine manner, IFN τ modulates the maternal immune system
37 and helps in the establishment of pregnancy. Besides IFN τ , another crucial molecule recently
38 came into limelight is indoleamine 2, 3-dioxygenase (IDO), which is an interferon-induced
39 tryptophan catabolic enzyme and is the first and rate-limiting enzyme of the tryptophan
40 catabolism via the kynurenine pathway. It is known to modify the maternal immune system by
41 altering the T cell metabolism and thereby inhibiting the allo-reactive T cell proliferation and at
42 the same time promoting the proliferation of immunosuppressive T regulatory cells. Moreover,
43 to achieve successful implantation, IDO favors an anti-inflammatory cytokine milieu by inducing
44 a shift towards the Th2 cytokines. Although many mechanisms have been delineated so far for
45 the pregnancy establishment in bovine, the focus of this chapter resides on the role of IFN τ and
46 IDO associated Th1-Th2 shift for a successful pregnancy with major emphasis on the tolerogenic
47 mechanism of these molecules.

48 Keywords: Bovine, Pregnancy, IFN τ , IDO, Th1-Th2 shift.

49

50 * Address correspondence to this author at the Department of Animal Biochemistry, ICAR-
51 NDRI; E-mail: sunilium46@gmail.com; sunilium46@rediffmail.com

52

53

54 **Abbreviations:**

55 3-HAA: 3-hydroxyanthranilic acid

56 AhR: Aryl hydrocarbon receptor

57 AI: Artificial insemination

58 BoLA-1: Bovine leukocyte antigen-1

59 CL: Corpus luteum

60 GCN2: General control non-repressed 2
61 IDO: Indoleamine 2, 3-dioxygenase
62 IFN γ : Interferon gamma
63 IFN τ : Interferon tau
64 ISGs: Interferon stimulating genes
65 MHC: Major histocompatibility complex
66 MRP: Maternal recognition of pregnancy
67 PBL: Peripheral blood leucocytes
68 PBMC: Peripheral blood mononuclear cells
69 PDK1: Phosphoinositide-dependent kinase-1
70 VEGF: Vascular endothelial growth factor

71

72 **Introduction:**

73 Reproduction is an essential, yet critical phenomenon adopted by every species to
74 propagate from one generation to another. A successful reproduction comprises many imperative
75 factors out of which, proper gamete formation, timely gamete transfer & fertilization, maternal
76 recognition of pregnancy (MRP) and implantation of the semi-allogeneic fetus into the mother's
77 uterus are considered to be the most important factors for achieving success in pregnancy. In
78 bovines, as production is directly related to its reproduction, therefore, optimal reproductive
79 efficiency is of utmost importance so as to increase their production competency. In order to
80 achieve maximum reproductive efficiency, proper detection of heat, timely insemination and
81 identification of animals returning to heat post-artificial insemination (post-AI) is indispensable.
82 Moreover, successful establishment of pregnancy also plays a crucial factor in bovine
83 reproduction, as failure in this process leads to poor productivity of the animals ([Koot et al.,](#)
84 [2012](#); [Norwitz et al., 2001](#)). However, the perspective of pregnancy establishment switched into
85 a different angle, when immunologists intervened and hypothesized a new dimensional role of
86 various undiscovered agents such as enzymes, nucleic acid (fetal DNA) and several maternal
87 immune cells in achieving successful pregnancy establishment in bovines. This new approach of
88 understanding pregnancy has not only changed the traditional concept of hormonal role in
89 pregnancy establishment ([Spencer and Bazer, 2004](#)) but has also helped immunologist to

90 manipulate maternal immune cells at different stages of pregnancy to enhance the reproductive
91 efficiency. Various maternal immune cells play a balanced and orchestrated role in pregnancy
92 establishment and allow the allograft fetus to acquire a suitable niche in the maternal
93 endometrium without undergoing rejection (Than et al., 2019; Robillard et al., 2022). Besides the
94 cellular role, the impact of different humoral factors like cytokines is inevitable. In association
95 with different immune cells, these humoral factors play a paramount role in the successful
96 establishment of pregnancy. Controlled activation of the maternal immune response during
97 pregnancy is yet another vital process that renders simultaneous responsiveness towards
98 unforeseen infections and at the same time provides tolerance towards the paternal antigen and
99 semi-allogeneic fetus for a successful pregnancy (Mor and Cardenas, 2010). Understanding the
100 mechanisms involved in the process of implantation and early pregnancy establishment may
101 significantly improve the reproductive efficiency and thus productivity in the animal sector.
102 During the past decades, changes in the morphology and histology of endometrium and
103 molecular mechanisms behind the feto-maternal crosstalk during the implantation window have
104 been intensively documented in mouse and humans (Zenclussen and Hämmerling, 2015; Meyer
105 and Zenclussen, 2020; Das et al., 2021). However, limited literature is available explaining the
106 inter-relationship between the Th1-Th2 cytokine shift and immune tolerance mechanism by
107 indoleamine 2, 3-dioxygenase (IDO) during the implantation window in bovines. Hence, this
108 chapter provides an overview of these processes and discusses the immune tolerance
109 mechanisms associated with the implantation of the blastocyst and early pregnancy
110 establishment in bovines.

111 **Key elements of early pregnancy establishment:**

112 Existing works of literature in the public domain provides a plethora of information and
113 hypothesis related to early pregnancy establishment in bovines (Oliveira et al., 2012; Pohler et
114 al., 2020); however, this chapter focuses on three crucial aspects of early pregnancy
115 establishment viz. maternal recognition of pregnancy (MRP), Th1-Th2 cytokine shift and
116 immune tolerance mechanism by IDO. Each element has been discussed in detail with the main
117 focus on the immune tolerance mechanism of IDO behind the embryo survivability during early
118 pregnancy establishment in bovines.

119 **1. Maternal recognition of pregnancy (MRP) in bovines:**

120 **Priming of the maternal immune system:**

121 The foundation of every successful pregnancy depends upon the bidirectional
122 communication between the embryo and the mother and this process starts much before the
123 embryonic tissue becomes intimately attached to the uterine epithelium. [Dekker and Robillard,](#)
124 [2007](#) reported that maternal tolerance towards the paternal antigen is initiated much before the
125 actual conception. A study conducted in mice model suggests that the seminal plasma secreted
126 by the accessory glands i.e. seminal vesicles and prostate gland of the male reproductive system
127 contains various signaling agents including estrogen, testosterone, transforming growth factor β
128 (TGF- β), prostaglandins and glycoproteins ([Maegawa et al., 2002](#)) which interact with the
129 epithelial cells of cervix and uterus causing modulation of the maternal immune system
130 ([Moldenhauer et al., 2009](#); [Robertson et al., 2009](#)). The role of seminal plasma in modulating
131 different cellular and molecular adjustments of the maternal reproductive environment for a
132 successful pregnancy in different domestic species has been extensively reviewed by [Bromfield,](#)
133 [2016](#). Moreover, a recent study in cattle demonstrates that seminal plasma exposure to the female
134 reproductive tract through natural mating affects conceptus development ([Mateo-otero et al.,](#)
135 [2020](#)). This type of modulation of the maternal immune system occurs as a consequence of the
136 effect of these seminal factors and the process is known as the “priming of the maternal immune
137 system”. This process renders endometrial tissue remodeling and cause improvement in its
138 receptivity towards the implanting embryo ([Robertson, 2005](#)). Therefore, priming of the maternal
139 immune system is considered as pivotal preliminary step, absolutely necessary before the actual
140 process of implantation and pregnancy establishment.

141 **Process of maternal recognition of pregnancy (MRP)**

142 After establishment of the priming of the maternal immune system, the next critical stage
143 is the attachment of embryo into the mother’s uterus (implantation). Prior to implantation, the
144 developing embryo gives early pregnancy signal which is recognized by the maternal
145 endometrium and immune cells that allows the growing conceptus for implantation. This process
146 is known as maternal recognition of pregnancy (MRP). MRP is a physiological phenomenon in
147 which the embryo gives a signal to the mother about its whereabouts. It renders the abrogation of

148 the luteolytic mechanism and allows corpus luteum (CL) for the continuous production of
149 progesterone throughout the gestation cycle. There are different endocrine mechanisms involved
150 in different species that recognizes the presence of pre-attachment embryos, including the two
151 best-known endocrine mechanisms comprised of estrogens produced by the porcine trophoblast
152 (Perry et al., 1973) or interferon tau (IFN τ) produced by early conceptus in ruminants (Bazer et
153 al., 1991; Spencer and Bazer, 2004) (Table-1). The duration of the pre-implantation period also
154 varies among different species, for instance, in case of ewes the elongating conceptus adheres to
155 the uterine epithelial surface and placentation starts around day 19 of gestation (Guillomot et al.,
156 1981; Boshier, 1969), in cattle placentation is around day 22 (Guillomot and Guay, 1982),
157 whereas, in mares the implantation is delayed until around day 37 of gestation (Allen et al.,
158 1973). Whatsoever may be the agents of MRP in different species, without the involvement of
159 these fundamental molecules, it is impracticable to protect the semi-allogeneic fetus from
160 maternal immune rejection.

161 **Interferon tau (IFN τ), the key MRP agent in bovines:**

162 It is the IFN τ in ruminants which helps the conceptus to get implanted into the mother's
163 uterus without undergoing rejection. IFN τ is a type-I interferon made up of single chains of
164 amino acids unique to the ruminants. It is believed that 36 million years ago IFN τ was evolved
165 from interferon omega (IFN ω) by some rearrangement and/or insertion events that combined
166 IFN ω gene with the trophoblast specific promoter/enhancer (Ealy and Wooldridge, 2017). IFN τ
167 is secreted from the mononuclear trophoblast cells of a viable conceptus and possesses antiviral,
168 antiproliferative and immunomodulatory properties (Michael Roberts, 1989a). In bovines, the
169 secretion of IFN τ starts from day 8 and goes up to 21-25 days post fertilization after which its
170 synthesis ceases. The peak level is around 14-18 days post fertilization (Martal et al., 1979). The
171 concentration of IFN τ and the size of the embryo are directly proportional to each other and
172 report suggests that there is an increase in IFN τ concentration with the increase in embryo size
173 during elongation (Robinson et al., 2006).

174 The question regarding molecular mechanism behind MRP still remains elusive.? There
175 are many schools of thought which explains the role of IFN τ in MRP, but the common ground to
176 the various thoughts is that the episodic release of prostaglandin PGF2 α from the endometrium is
177 regulated by the hormone oxytocin which is mediated via the coupling of oxytocin with its

178 receptor. IFN τ acts locally in a paracrine manner and inhibits the expression of the estrogen
179 receptor which in turn abate oxytocin receptor expression in endometrium. Due to this
180 phenomenon, endometrium fails to produce the luteolytic agent PGF2 α , thus preventing lysis of
181 corpus luteum and there is a continuous production of progesterone (Spencer et al., 1995). Apart
182 from inhibiting luteolysis, IFN τ has also an impact on different intra and extra-uterine tissues in
183 bovine (Oliveira et al., 2008). Through a paracrine manner, IFN τ acts on the endometrial tissue
184 and induces several genes that directly or indirectly assist conceptus elongation and implantation
185 (Bazer et al., 2008). These induced genes are broadly called as interferon stimulating genes
186 (ISGs) which can also be induced by other type I interferons (Hansen et al., 2010; Pervolaraki et
187 al., 2018). Although IFN τ locally alters the expression of more than 750 ISGs in the
188 endometrium, some ISGs like interferon-stimulated gene 15 (ISG15), 2'-5'-oligoadenylate
189 synthetase 1 (OAS1) and myxovirus resistance (MX1 and MX2) showed higher and differential
190 expressions during early day 16 post-AI (Forde et al., 2011) and hypothesized to be used as early
191 pregnancy diagnosis markers. Besides stimulating the intrauterine tissues, IFN τ has also a
192 significant impact on extra uterine tissues (Endriß et al., 2021; Sheikh et al., 2018). An
193 undetectable level of IFN τ when enters local or peripheral circulation from the uterus causes
194 significant molecular changes in extra uterine tissues like CL (Shirasuna et al., 2015), peripheral
195 blood leucocytes (PBL) (Gifford et al., 2007), peripheral blood mononuclear cells (PBMCs)
196 (Matsuyama et al., 2012), neutrophils (Panda et al., 2020) and hepatocytes (Ruhmann et al.,
197 2017) through endocrine manner. It is now well known that, in order to carry out pregnancy
198 establishment under the expenses of a foreign fetus, IFN τ inhibits lymphocytes proliferation and
199 induces neutrophil chemotaxis toward uterus to start a non-specific immune response in the
200 uterus (Teixeira et al., 1997). Also, there are other reports which suggest a decrease in the
201 neutrophil activity during the peri-implantation period and are believed to be attributed towards
202 the IFN τ stimulation (Mohammed et al., 2017b). Moreover, IFN τ increases the expression of
203 bovine leukocyte antigen-1 (BoLA-1) in bovine endometrial luminal epithelial cells which is
204 known to have profound immunosuppressive effects during early embryonic development in
205 bovines. BoLA-1 inhibits the proliferation of cytotoxic T cell and natural killer cell (NK cell)
206 which ultimately induces feto-maternal tolerance in bovines (Zhu et al., 2017). According to
207 them, IFN τ up-regulates the expression of anti-inflammatory cytokine IL-10 while decreases the
208 expression of pro-inflammatory cytokine IL-6 in the endometrial tissue of bovines. After

209 unravelling so many different roles of IFN γ in bovine pregnancy, it is interesting to take a note
210 that this 24 kDa molecule has numerous potential and pivotal functions which renders the
211 process of pregnancy towards its success.

212 **2. Th1-Th2 cytokine balance during pregnancy:**

213 Pregnancy exhibits an immunological paradox state which is the most unique and
214 challenging task for the maternal immune system. A successful pregnancy requires significant
215 modulation of the maternal immune system that facilitates the acceptance of specific foreign
216 antigens to support reproductive function without compromising maternal immune protection
217 from pathogenic infections and inflammatory insults. Under such an immunological
218 contradiction state, an increase in innate immune activity may be expected to protect the uterus
219 from infection. But, this precise regulation of maternal immune modulation remains somewhat
220 elusive. Several mechanisms have been proposed by different scientists that render the embryo to
221 survive in the maternal environment. One of them includes the antigenic immaturity of the
222 conceptus during which the bovine trophoblast, like other mammalian species, doesn't express
223 classical polymorphic major histocompatibility complex (MHC) class 1 proteins in areas in touch
224 with the maternal endometrium during early pregnancy ([Bainbridge, 2000](#)) which gives maternal
225 immunological dormancy to the conceptus or localized immune tolerance to the growing
226 conceptus ([Medawar, 1953](#)). Besides priming of the maternal immune system, early embryonic
227 signal i.e. MRP and antigenic immaturity of the conceptus, a delicate harmony between pro and
228 anti-inflammatory cytokines is also needed for successful establishment and maintenance of
229 pregnancy.

230 **Classification of Th1 and Th2 cytokines:**

231 Cytokines are the soluble hormone-like proteins which are secreted by white blood cells
232 as well as many other cells like endothelial cells, epithelial cells, and fibroblast. Cytokines
233 include chemokines, monokines, lymphokines, interleukins, colony-stimulating factors, tumor
234 necrosis factors, interferons, etc. They mediate several functions like inflammation, immunity by
235 influencing cellular recruitment, proliferation, differentiation, activation, migration, suppression
236 and killing. T-lymphocytes are considered as the major source of cytokine production and their
237 cytokine secretion profile largely depends upon the availability of cytokines in their

238 microenvironment which helps the naive T helper (Th0) cells to mature and differentiate into
239 different subtypes. When Th0 cells are exposed to IFN γ they become Th1 cells and if they are
240 exposed to IL-4 they differentiate into Th2 type cells (Palmer and Van Severter, 1997; Asnagli
241 and Murphy, 2001). The cytokines produced by these two subsets are respectively known as Th1
242 and Th2 cytokines. Th1 cytokines are regarded as the pro-inflammatory cytokines which are
243 responsible for inflammatory response whereas, Th2 cytokines are known as anti-inflammatory
244 cytokines responsible for downregulation (suppression) of exaggerated and/or inappropriate Th1
245 response (Gianfranco Del Prete, 1998; Panda et al., 2020; Mohapatra et al., 2020). The major
246 Th1 cytokine is IFN γ which is regarded as the most potent inflammatory agent. Besides IFN γ ,
247 other cytokines like TNF α , IL-2, IL-8, IL-12 and IL-17 are also considered as potent
248 inflammatory agents, grouped under Th1 cytokines. In contrast, Th2 responses are characterized
249 by the production of various Th2 cytokines like IL-4, IL-5, IL-10 and IL-13 which are associated
250 with anti-inflammatory response (Garra, 2000; Kurt, 1987; Romagnani, 1992). Other T-helper
251 responses include T-regulatory (T-reg) cell responses, determined by the secretion of IL-10 and
252 TGF- β , and Th17 responses, defined by the secretion of IL-17 (Mucida, 2010).

253 **Potent bovine pregnancy-associated cytokines (Th1 and Th2)**

254 IL-1 family comprises 11 cytokines out of which IL-1 α and IL-1 β are the most studied
255 ones and are known to have pro-inflammatory nature. According to Oliveira et al. 2013, the
256 presence of embryo in the pregnant bovine endometrium lowered the mRNA expression of IL-1
257 β when compared to the cyclic cows. Additionally, non-pregnant cows exhibited higher plasma
258 IL-1 β concentration than pregnant cows (Manjari et al., 2018). However, *in vitro* studies reveal
259 that IL-1 modulates the production of luteotrophic PGE2 and luteolytic PGF2 α locally in bovine
260 CL as well as in endometrium (Nishimura et al., 2004; Tanikawa et al., 2005). IL-1 α is believed
261 to be more potent towards PGE2 production as compared to IL-1 β (Tanikawa et al., 2008) and
262 the luteotrophic action is modulated by IFN τ during early pregnancy which may support
263 implantation and early embryonic development in bovines (Majewska et al., 2010). PGE2 not
264 only sustains the progesterone production by its luteo-protective action but also down-regulates
265 IL-2 expression in bovine PBL to maintain the pregnancy (Emond et al., 1998). IL-2 is a pro-
266 inflammatory cytokine and is known to be a T cell growth and differentiation factor (Ross and
267 Cantrell, 2018). How IL-2 affects T cell population during bovine pregnancy is largely unknown.

268 However, higher plasma IL-2 concentration was noticed in cows which exhibited early
269 embryonic mortality and late embryonic mortality as compared to pregnant cows (Panda et al.,
270 2020). IL-4 being an anti-inflammatory cytokine and a key regulator of innate as well as adaptive
271 immune system causes differentiation of naive helper T cells (Th0 cells) to Th2 cells (Seder et
272 al., 1992). In addition to this IL-4 increases its expression in a positive feedback manner and
273 decreases the expression of IFN γ in a negative feedback loop. Higher expression of IL-4 during
274 pregnancy is necessary for embryo implantation which is modulated mainly by IFN τ (Tuo et al.,
275 1999) and progesterone (Maeda et al., 2013). IL-6 acts as both pro and anti-inflammatory
276 cytokine. It is known to increase the inner cell mass of bovine embryo (Wooldridge and Ealy,
277 2019) and along with leukemia inhibitory factor; IL-6 induces the proliferation and attachment of
278 porcine trophoblast cell during early pregnancy (Blitek et al., 2012). Contrary, around 18 days
279 post AI, non-pregnant cows displayed higher plasma IL-6 level as compared to the pregnant
280 cows (Mohammed et al., 2017). IL-8 is a pro-inflammatory cytokine and is known to be a potent
281 chemoattractant for neutrophil towards the inflammatory site. Besides their pro-inflammatory
282 activity, they also promote angiogenesis in the developing CL (Jiemtaweeboon et al., 2011).
283 Higher plasma IL-8 levels have been shown to increase the immune response towards the
284 implanting embryo thus resulting in implantation failure in bovines (Sheikh et al., 2019). Being
285 an anti-inflammatory cytokine, IL-10 puts a break on the innate and adaptive immune system
286 and limits tissue damage due to inflammation (Ouyang et al., 2011). Pregnant bovine
287 endometrium demonstrates an increase in IL-10 expression which is necessary for the
288 maintenance of the pregnancy (Oliveira et al. 2013). According to Shirasuna et al. (2012),
289 increased IL-10 expression in PBMCs is enhanced mainly due to IFN τ stimulation. Whatsoever
290 may be the mechanism behind the cytokines mediated maternal immune modulation, it is
291 imperative that without an orchestrated balance between pro-inflammatory and anti-
292 inflammatory cytokines, the success in pregnancy can never be achieved.

293 **Th2 bias during early pregnancy establishment:**

294 Wegmann and colleagues reported that there is biasness towards the Th2 cytokine
295 pathway during pregnancy which promotes tolerance towards the half foreign fetus (figure-1).
296 Moreover, Th1 cytokines are regarded as harmful for the tolerance of semi-allogeneic fetus
297 which is similar to allografts in transplant recipients (Wegmann et al., 1993; Erdmann et al.,

298 [2004](#)). It has been well documented that during the induction of tolerance to the allograft tissue
299 there is an increase in Th2 cytokines including IL-4 and IL-10 and a decrease in Th1 cytokines
300 such as IL-2 and IFN γ . Conversely, the allograft gets rejected when there is a high level of IL-2
301 and IFN γ ([Strom et al., 1996](#); [Li et al., 1998](#)). One of the interesting *in vitro* study conducted by
302 [Mao and co-workers \(2010\)](#), revealed that when T-reg cells purified from pregnant women were
303 cultured with paternal or unrelated PBMCs, T-reg cells caused the suppression of the
304 inflammatory cytokine IFN γ which ultimately inhibited the inflammatory type 1 (cell-mediated)
305 immune responses.

306 Many of the scientists believe that the Th2 bias in pregnancy is an oversimplified concept
307 and there is a dynamic modulation of pro and anti-inflammatory cytokines that exist during
308 various stages of pregnancy. In general, the pregnancy has been categorized into three important
309 stages according to the predominant cytokines milieu in different stages of pregnancy. The first
310 stage of pregnancy is the implantation of the blastocyst into the mother's uterus which is
311 predominantly a pro-inflammatory phase. In this stage, the mother's immune system tries to
312 accommodate the alien antigens. The second stage of pregnancy which is the phase of growth
313 and development of the fetus is predominantly an anti-inflammatory phase. Th2 cytokine bias
314 during this stage can be systemic or local at the feto-maternal interface. The last stage of
315 pregnancy is parturition in which expulsion of the fetus occurs by contraction of the uterus and
316 again the pro-inflammatory milieu is predominant at this stage. During all the stages of
317 pregnancy, the process of inflammation is tightly regulated by the maternal immune system and
318 failure in regulation during any stage will result in adverse pregnancy outcomes.

319 Microarray analysis performed by [Walker and colleagues \(2010\)](#), revealed 1,839 and
320 1,189 differentially expressed transcripts in the caruncular and intercaruncular endometrium
321 respectively between the pregnant and cyclic Holstein-Friesian dairy cattle. In their study, they
322 observed up-regulation of the chemokine receptor CXCR3 ligands (CXCL9, CXCL10 and
323 CXCL11) and downregulation of the CXCR3 receptor in pregnant animals. The CXCR3 receptor
324 is primarily expressed on Th1 cells ([Patel et al., 2001](#)) and its downregulation in pregnancy
325 suggests that either Th1 cells were not increased in the uterus or the Th1 cells are not expressing
326 CXCR3 receptors during the implantation window. It has been observed that the population of
327 uterine NK (uNK) cells dramatically increases in maternal decidua during early pregnancy and
328 continues to increase up to the end of the first trimester of pregnancy ([Bulmer et al., 1991](#); [Lee et](#)

329 [al., 2011](#)). These uNK cells also express CXCR3 receptors on their cell surface ([Sentman et al.,](#)
330 [2004](#)) and up-regulation of this receptor's ligand may have a role in the activation of uNK cells
331 in the uterus during pregnancy and thus initiating an innate immune response at the feto-maternal
332 interface so as to protect the mother from exogenous infections. Besides CXCs, there are several
333 other chemokines such as CCL11, CCL2, CCL8, etc. which attract immune tolerance promoting
334 immune cells like Th2 cells and NK cells during early pregnancy ([Walker et al., 2010](#)). Up-
335 regulation of chemokine CCL11 attracts CCR3 (Receptor of CCL11) expressing Th2 cells
336 ([Dominguez et al., 2003](#)) and CCL2 chemokine up-regulation attracts leucocytes which express
337 its receptor i.e., CCR2 ([Dominguez et al., 2003](#); [Meter et al., 2005](#)). Another chemokine CCL8
338 also known as monocyte chemo-attractant protein 2 (MCP2) is responsible for attracting
339 monocytes, NK cells, lymphocytes, eosinophils and basophils through bindings of its receptor
340 CCR1, CCR2, CCR3 and CCR5 during implantation ([Hannan and Salamonsen, 2007](#)). CCL8,
341 CCL2 along with chemokine receptor CX3CR1 are actively involved in leukocyte recruitment
342 and neovascularization during wound repair ([Ishida et al., 2008](#); [Balaji et al., 2015](#)). In addition
343 to chemokines, several interleukins such as IL-15, IL-7, and TGF β also play a significant role in
344 the establishment of pregnancy. IL-15 induces the proliferation of immune tolerance promoting
345 T-reg cells in the uterus of the pregnant animals which shifts the inflammatory response towards
346 the anti-inflammatory pathway ([Guerin and Robertson, 2009](#)). Likewise, the up-regulation of IL-
347 7 in the pregnant endometrium enhances growth and survival-promoting factor for T-reg cells
348 ([Harnaha et al., 2006](#)). Although, the role of different Th1 and Th2 cytokines in the context of
349 bovine pregnancy needs to be more precisely investigated, but, these preliminary ideas points
350 towards a Th2 bias during the establishment of bovine pregnancy.

351 **Role of interferon-stimulated genes (ISGs) during pregnancy:**

352 Another group of scientists ([Bauersachs et al., 2006](#); [Mitko et al., 2008](#); [Mansouri-Attia](#)
353 [et al., 2009](#)) also reported an up-regulation of interferon-stimulated genes (ISGs) in pregnant
354 animals. Among ISGs, the up-regulation of the OAS gene during early pregnancy is involved in
355 the regulation of osteopontin (SPP1) production ([Spencer et al., 1999](#); [McAveney et al., 2000](#)).
356 SPP1 up-regulation in pregnant animals may have several crucial functions which include the
357 promotion of trophoblast adhesion to the maternal endometrium, stimulation of morphological
358 changes in the trophoblast and regulation of the immune response ([Johnson et al., 2003](#)). Various

359 immune cells like NK cells, macrophages, monocytes and T-lymphocytes express SPP1 which is
360 believed to regulate the Th1/Th2 balance and apoptosis (Johnson et al., 2003). At this critical
361 immunosuppressive stage of pregnancy, the mother also needs to protect her from pathogenic
362 infections and this protection is facilitated by the up-regulation of some other ISGs like MX1 and
363 MX2. It has been hypothesized that to encounter the potential viral pathogens during the time of
364 local immune suppression mother's uterus express MX1 and MX2, suggesting that the innate
365 immune system is active during early pregnancy (Hicks et al., 2003). Beside uterus, it has been
366 reported that peripheral blood leucocytes also express MX2 during pregnancy (Gifford et al.,
367 2007).

368 In addition to ISGs, there are several additional ISGs like IFITM1, Peptide Transporters
369 (TAP-1 and TAP-2), MHC I A, and MHC I G which may have the function to provide localized
370 immune system suppression that allows the embryo to survive within the uterus (Walker et al.,
371 2010). IFITM1 acts as an immunomodulatory molecule and up-regulation of this gene is
372 believed to be linked with immune-modulation of NK cells (Yang et al., 2005) thus create an
373 immunosuppressive milieu during the implantation period. It has been observed that the local
374 immune suppression is also associated with the up-regulation of TAP proteins, as the up-
375 regulation of TAP-1 protein is involved in the transformation of natural killer cells to become
376 non-cytotoxic (Kalkunte et al., 2009). According to Kalkunte et al. (2009), this transformation of
377 NK cells is achieved through the expression of vascular endothelial growth factor-C (VEGF-C)
378 which with the help of induction of TAP-1 protein enhances the resistance of trophoblast and
379 endothelial cells to lysis. These findings demonstrate and highlight the necessity of maternal
380 immune-modulation during early pregnancy and the fundamental requirement of a delicate
381 balance between Th1/Th2 cytokines along with the pivotal importance of ISGs during early
382 pregnancy establishment in bovines.

383 **3. Role of indoleamine 2, 3-dioxygenase (IDO) in fetal immune-tolerance**

384 Pregnancy exhibits a complex phenomenon in which a delicate balance between pro and
385 anti-inflammatory cytokines is needed to maintain maternal immune system integrity while
386 preventing rejection of the embryo. In this critical process, IDO plays an important role in
387 shifting these cytokines according to the need of the maternal immune system. IDO is the first
388 and rate-limiting enzyme of the tryptophan catabolism via the kynurenine pathway which is the
389 major route of L-tryptophan catabolism and a mechanism for nicotinamide adenine dinucleotide

390 (NAD⁺) synthesis in mammalian tissues. IDO converts tryptophan into N-formyl-kynurenine
391 which is then converted into several biologically active metabolites like 3-hydroxykynurenic
392 acid, 3-hydroxyanthranilic acid and quinolinic acids (Stone and Darlington, 2002). IDO is an
393 intracellular enzyme and to date, none of its secreted or extracellular forms has been identified.
394 Evidence shows that IDO is expressed at the maternal-fetal interface, including trophoblast cells,
395 decidual stroma cells, decidual immune cells (e.g., natural killer cells, T cells, and macrophages),
396 and vascular endothelial cells of decidua and chorion (Kudo et al., 2004; Von Rango et al.,
397 2007).

398 A novel IDO paralogue known as indoleamine 2,3-dioxygenase-2 (IDO2), was discovered
399 independently by several groups (Ball et al., 2007, Metz et al., 2007) in 2007. It is not only
400 present in mammals but also found in lower vertebrates. It is named based on its structural
401 similarity and enzymatic activity to ‘conventional’ IDO (referred to as IDO1). The IDO1 and
402 IDO2 enzymes share a similar range of substrates (Ball et al., 2007), while they differ in some
403 biological functions, for instance, their selectivity for inhibitors. The selective inhibition of IDO2
404 is 1-methyl-D-tryptophan, while 1-methyl-L-tryptophan is a better inhibitor of IDO1 activity
405 (Ball et al., 2009). As most of the studies in the literature have not specialized the role of IDO1
406 and IDO2 separately, the term “IDO” will refer to the collective functional IDO enzyme activity,
407 unless otherwise specified. Apart from IDO, another tryptophan degrading enzyme is tryptophan
408 2,3-dioxygenase (TDO). TDO is widely distributed in both eukaryotes and bacteria whereas,
409 IDO has only been found in mammals and yeast.

410 The regulatory mechanism of IDO expression has been studied thoroughly by different
411 groups of scientists and substantial evidence suggests a complex counter-regulatory mechanism
412 of IDO expression at the feto-maternal interface. Researchers have revealed that IDO expression
413 is initially induced by certain pro-inflammatory molecules like IFN γ (most potent IDO
414 stimulator) produced from the T-cell activation (Miwa et al., 2005). This finding is well
415 correlated with the previous discussion in the review that the first phase of the pregnancy is a
416 pro-inflammatory phase. However, once the expression of IDO becomes significantly high, it
417 may counter-regulate the inflammatory molecules by suppressing the alloreactive T-cells and
418 skewing their differentiation towards immunosuppressive T-regulatory cells (Mezrich et al.,
419 2010). At this stage, the innate immunity of the mother takes over the charge and prevents the
420 mother from pathogenic infection (Hicks et al., 2003) with simultaneous protection of the

421 allogeneic fetus. The detailed mechanism behind the IFN γ mediated IDO up-regulation remains
422 unclear. However, it is reported that IFN- γ induces Janus-activated kinase (JAK)/STAT1 and
423 protein kinase C δ (PKC δ) signaling and blocking this pathway resulted in the suppression of
424 IDO expression in cancer cells (Jeong et al., 2009; Cheng et al., 2010). Nevertheless, these
425 findings provide us a clue about the possible mechanism behind IDO expression due to IFN γ
426 production in early pregnancy (Boyt et al., 2020).

427 In 1998 immunostaining technique performed by Munn and co-workers revealed that there is
428 a high level of IDO staining in the syncytiotrophoblasts of the placenta (Munn et al., 1998;
429 Mellor and Munn, 2001). They hypothesized that increased IDO expression in the placenta plays
430 a significant role in the prevention of the semi-allogeneic fetal rejection during early pregnancy.
431 In support of their hypothesis, they used IDO inhibitor (1-methyl-tryptophan) at the
432 pharmacological range and observed a rapid T-cell mediated rejection of allogeneic fetus (Munn
433 et al., 1998). Since then, the mechanism behind the immunosuppressive action of IDO in
434 successful pregnancy has been explored by several immunologists. The most accepted and
435 widely used theory behind the immunosuppressive action of IDO is based on the tryptophan
436 depletion theory (Mellor and Munn, 1999; Munn et al., 1999). According to this theory,
437 overexpression of IDO in certain types of cells will consume the available tryptophan in the
438 microenvironment; as a result, the neighboring maternal T-cell will starve of this essential amino
439 acid. Subsequently, the deficiency of tryptophan blocks the progression of the cell cycle in
440 activated T-cells which leads to T-cell arrest at the mid-G1 phase of the cell cycle and the cells
441 undergo apoptosis (Lee et al., 2002). This process leads to immunosuppression due to the
442 inhibition of locally activated T-cell proliferation. Schematic representation of IDO mediated
443 maternal immunosuppression in bovines has been depicted in [figure-2](#).

444 Tryptophan depletion theory has been supported by some experimental observations for
445 instance Mellor et al, reported that up-regulation of IDO expression in dendritic cells restricted
446 T-cell clonal expansion along with rapid death of activated T-cells (Mellor et al., 2004).
447 However, when tryptophan was supplied in excess the T-cell responses were restored. This
448 tryptophan starvation due to IDO expression also affects the neighboring cytotoxic T-cells and
449 reduces its cytotoxicity via downregulating the T-cell receptor (TCR) zeta-chain (Fallarino et al.,
450 2006). It has been studied that depletion of tryptophan activates certain kinases like general
451 control non-repressed 2 (GCN2) kinase which suppresses glycometabolism and proliferation of

452 alloreactive T-cells, thereby causing immunosuppression at the feto-maternal interface
453 (Eleftheriadis et al., 2013; Eleftheriadis et al., 2014).

454 With the increasing knowledge and research related to the IDO mechanism, the
455 immunologists explored yet another mechanism of IDO immune-tolerance which is mediated by
456 the biologically active metabolites of kynurenine produced during the process of tryptophan
457 catabolism (Terness et al., 2007; Badawy et al., 2016). Kynurenine is a tryptophan metabolite
458 that behaves like an endogenous ligand for the activation of aryl hydrocarbon receptor (AhR)
459 (Opitz et al., 2011; Liu et al., 2017). Upon activation of this receptor along with TGF β exposure
460 a cascade of signaling pathway is generated which skews the differentiation of alloreactive T-
461 cells to immunosuppressive Foxp3⁺ T regulatory cells (Nguyen et al., 2010; Mezrich et al.,
462 2010). Interestingly, this AhR signaling has also been shown to induce IDO expression and
463 activates 2 metabolites of kynurenine (kynurenic acid and xanthurenic acid) which ultimately
464 participate in the process of immune-suppression during early pregnancy (Jaronen and Quintana,
465 2014; Fazio et al., 2017; Nguyen et al., 2014). Other metabolites like 3-hydroxyanthranilic acid
466 (3-HAA) and quinolinic acids also affect the T-cell population. It has been reported that these
467 metabolites activate caspase 8 and releases cytochrome-c from the mitochondrial matrix which
468 can induce the selective apoptosis of Th1 cells rather than Th2 cells (Fallarino et al., 2002). 3-
469 HAA can also cause immunosuppression directly by inhibiting the activation of dendritic cells
470 and phosphoinositide-dependent kinase-1 (PDK1) along with depletion of intracellular
471 glutathione levels, ultimately leading to T-cell apoptosis (Hayashi et al., 2007; Lee et al., 2010).
472 Other tryptophan metabolites like 3-hydroxykynurenine (3-HK) and picolinic acid have also
473 been observed to cause immunosuppression during pregnancy; however, their mechanism of
474 action remains unclear (Zaher et al., 2011; Prodinger et al., 2016).

475 Beside IDO expressing immune cells, there are some non-immune cells like fetal
476 trophoblast cells, maternal endometrial/decidual stromal cells which also express IDO during
477 pregnancy and play a pivotal role in maternal-fetal tolerance. IDO expressing fetal trophoblast
478 cells remain in close proximity to the maternal immune system and suppresses the proliferation
479 of alloreactive T-cells (Dong et al., 2008). It also reduces the Th1/Th2 cytokine ratio at the feto-
480 maternal interface during pregnancy, thereby switching the cytokine profile towards the Th2
481 pathway (Li et al., 2007). Apart from immunological modulation by IDO expression in
482 trophoblast, IDO can directly influence fetal trophoblast in terms of its proliferation and

483 migration which has been observed to be mediated via the STAT3 signaling pathway (Zong et
484 al., 2016). Meanwhile, the maternal decidual stromal cells also express IDO which induces
485 macrophage tolerance via IL-33, thereby reducing the phagocytic ability of the macrophages
486 (Mei et al., 2014). These IDO expressing stromal cells also affect the proliferation, cytotoxicity
487 and cytokine production of NK cells and prepare the endometrium to become receptive for the
488 implantation of the semi-allogeneic fetus (Croxatto et al., 2014).

489 Numerous studies have also been conducted in relation to pathological pregnancy in
490 which the role of IDO has been implicated. In the case of recurrent spontaneous abortion (RSA),
491 it has been documented that the level of IDO expression at the maternal-fetal interface is
492 profoundly low as compared to normal pregnancy (Ban et al., 2013). Along with low IDO
493 expression, there is also a reduction in STAT3 phosphorylation and MMP9 expression observed
494 in RSA patients which indicates an imbalance in the IDO-STAT3 pathway (Zong et al., 2016).
495 As mentioned earlier that the most potent stimulator of IDO expression is IFN γ , the production
496 of this cytokine by decidual and peripheral blood mononuclear cells were surprisingly found to
497 be low in RSA patients resulting in a low level of IDO expression and activity (Miwa et al.,
498 2005) thereby leading to failure of pregnancy. A similar trend has been observed in the case of
499 preeclampsia in which there is a reduction in IDO expression and activity leading to diminished
500 numbers of Foxp3+ T-reg cells (Liu et al., 2011) and thereby leading to pregnancy failure. These
501 findings highlight the essential role of IDO expression at the feto-maternal interface to achieve
502 fetal immune tolerance during a successful pregnancy. However, there is still insufficiency of the
503 detailed mechanism behind IDO immunosuppression that exists in normal pregnancy and
504 extensive future study is needed to explore the immune-regulatory action of IDO at the maternal-
505 fetal interface.

506 **Conclusion:**

507 To meet the increasing demand of the growing population in terms of milk and beef
508 requirements, the production potential of cattle has to be improved. As production is directly
509 related to the reproduction, therefore improving reproductive efficiency of cattle is the need of
510 the hour. However, embryonic mortality, failure of successful implantation, fetal rejection due to
511 undue maternal immune system activation etc., are some of the major elements to derail its
512 success by production, reproduction and economic mean. Nutritional, genetical, climatic,

513 endocrine, inappropriate uterine environment, pathological agents are the major issues known to
514 be the predisposing factors for low reproductive efficiency in bovines. Moreover, improper
515 maternal immune activation might be another area which needs to be explored comprehensively
516 in bovines. Understanding the embryo-maternal bidirectional communication thorough different
517 approaches/ techniques might help us to decode it and some strategic measures like use of
518 immune-modulators, identification and selection of immune-competent breed, development of
519 some mathematical models to diagnose poor reproductive efficiency might be useful for
520 formulating as well as adopting different cattle breeding and management policy. Understanding
521 the different mechanisms behind bovine pregnancy establishment will not only help to prevent
522 the embryonic mortality but also will reduce the non-pregnancy period in the cattle. Although the
523 basic research on bovine reproductive immunology is expanding, the extent and intensity of
524 maternal immune modulation by different cellular and humoral factors during bovine pregnancy
525 remains elusive. Thus, further research is needed to ensure better immune management practices
526 during transition and post partum stages of bovine so as to improve reproductive efficiency and
527 thereby to achieve optimal production potential in bovines.

528 **Conflict of interest:**

529 None declare

530

531 **Acknowledgement:**

532 None declare

533 **References:**

- 534 1. Koot, Y.E.M., Teklenburg, G., Salker, M.S., Brosens, J.J. and Macklon, N.S., 2012.
535 Molecular aspects of implantation failure. *Biochimica et Biophysica Acta (BBA)-Molecular*
536 *Basis of Disease*, 1822(12), pp.1943-1950.
- 537 2. Norwitz, E.R., Schust, D.J. and Fisher, S.J., 2001. Implantation and the survival of early
538 pregnancy. *New England Journal of Medicine*, 345(19), pp.1400-1408.
- 539 3. Spencer, T.E. and Bazer, F.W., 2004. Conceptus signals for establishment and maintenance
540 of pregnancy. *Reproductive Biology and Endocrinology*, 2(1), p.49.
- 541 4. Than, N.G., Hahn, S., Rossi, S.W. and Szekeres-Bartho, J., 2019. Fetal-Maternal Immune
542 Interactions in Pregnancy. *Frontiers in Immunology*, 10, p.2729.

- 543 5. Robillard, P.Y., Dekker, G., Scioscia, M. and Saito, S., 2022. Progress in the understanding
544 of the pathophysiology of immunologic maladaptation related to early-onset preeclampsia
545 and metabolic syndrome related to late-onset preeclampsia. *American Journal of Obstetrics
546 and Gynecology*.
- 547 6. Mor, G. and Cardenas, I., 2010. The immune system in pregnancy: a unique complexity.
548 *American journal of reproductive immunology*, 63(6), pp.425-433.
- 549 7. Zenclussen, A.C. and Hämmerling, G.J., 2015. Cellular regulation of the uterine
550 microenvironment that enables embryo implantation. *Frontiers in immunology*, 6, p.321.
- 551 8. Meyer, N. and Zenclussen, A.C., 2020. Immune Cells in the Uterine Remodeling: Are They
552 the Target of Endocrine Disrupting Chemicals?. *Frontiers in immunology*, 11, p.246.
- 553 9. Das, D., Saikia, P.J., Gowala, U. and Sarma, H.N., 2021. Cell specific expression of vascular
554 endothelial growth factor receptor-2 (Flk-1/KDR) in developing mice embryo and supporting
555 maternal uterine tissue during early gestation (D4-D7). *International journal of fertility &
556 sterility*, 15(2), p.148.
- 557 10. Oliveira, L.J., Barreto, R.S.N., Perecin, F., Mansouri-Attia, N., Pereira, F.T.V. and Meirelles,
558 F.V., 2012. Modulation of maternal immune system during pregnancy in the cow.
559 *Reproduction in Domestic Animals*, 47, pp.384-393.
- 560 11. Pohler, K.G., Franco, G.A., Reese, S.T. and Smith, M.F., 2020. Physiology and pregnancy of
561 beef cattle. In *Animal Agriculture* (pp. 37-55). Academic Press.
- 562 12. Dekker, G. and Robillard, P.Y., 2007. Pre-eclampsia: is the immune maladaptation
563 hypothesis still standing?: an epidemiological update. *Journal of reproductive immunology*,
564 76(1-2), pp.8-16.
- 565 13. Maegawa, M., Kamada, M., Irahara, M., Yamamoto, S., Yoshikawa, S., Kasai, Y., Ohmoto,
566 Y., Gima, H., Thaler, C.J. and Aono, T., 2002. A repertoire of cytokines in human seminal
567 plasma. *Journal of reproductive immunology*, 54(1-2), pp.33-42.
- 568 14. Moldenhauer, L.M., Diener, K.R., Thring, D.M., Brown, M.P., Hayball, J.D. and Robertson,
569 S.A., 2009. Cross-presentation of male seminal fluid antigens elicits T cell activation to
570 initiate the female immune response to pregnancy. *The Journal of Immunology*, 182(12),
571 pp.8080-8093.
- 572 15. Robertson, S.A., Guerin, L.R., Bromfield, J.J., Branson, K.M., Ahlström, A.C. and Care,
573 A.S., 2009. Seminal fluid drives expansion of the CD4⁺ CD25⁺ T regulatory cell pool and

- 574 induces tolerance to paternal alloantigens in mice. *Biology of reproduction*, 80(5), pp.1036-
575 1045.
- 576 16. Bromfield, J.J., 2016. A role for seminal plasma in modulating pregnancy outcomes in
577 domestic species. *Reproduction*, 152(6), pp.R223-R232.
- 578 17. Mateo-Otero, Y., Sánchez, J.M., Recuero, S., Bagés-Arnal, S., McDonald, M., Kenny, D.A.,
579 Yeste, M., Lonergan, P. and Fernandez-Fuertes, B., 2020. Effect of Exposure to Seminal
580 Plasma Through Natural Mating in Cattle on Conceptus Length and Gene Expression.
581 *Frontiers in Cell and Developmental Biology*, 8, p.341.
- 582 18. Robertson, S.A., 2005. Seminal plasma and male factor signalling in the female reproductive
583 tract. *Cell and tissue research*, 322(1), pp.43-52.
- 584 19. Perry, J.S., Heap, R.B. and Amoroso, E.C., 1973. Steroid hormone production by pig
585 blastocysts. *Nature*, 245(5419), pp.45-47.
- 586 20. Bazer, F.W., Thatcher, W.W., Hansen, P.J., Mirando, M.A., Ott, T.L. and Plante, C., 1991.
587 Physiological mechanisms of pregnancy recognition in ruminants. *Journal of reproduction
588 and fertility. Supplement*, 43, pp.39-47.
- 589 21. Flint, A.P.F., Guesdon, F.M.J. and Stewart, H.J., 1994. Regulation of trophoblast interferon
590 gene expression. *Molecular and cellular endocrinology*, 100(1-2), pp.93-95.
- 591 22. Geisert, R.D., Zavy, M.T., Moffatt, R.J., Blair, R.M. and Yellin, T., 1990. Embryonic
592 steroids and the establishment of pregnancy in pigs. *J Reprod Fertil Suppl*, 40, pp.293-305.
- 593 23. Ticconi, C., Zicari, A., Belmonte, A., Realacci, M., Rao, C.V. and Piccione, E., 2007.
594 Pregnancy-promoting actions of HCG in human myometrium and fetal membranes. *Placenta*,
595 28, pp.S137-S143.
- 596 24. Ziecik, A.J., Waclawik, A., Kaczmarek, M.M., Blitek, A., Jalali, B.M. and Andronowska, A.,
597 2011. Mechanisms for the establishment of pregnancy in the pig. *Reproduction in domestic
598 animals*, 46, pp.31-41.
- 599 25. Pineda, M.H. and Dooley, M.P., 2003. *McDonald's veterinary endocrinology and
600 reproduction (No. Ed. 5)*. Iowa state press.
- 601 26. Guillomot, M., Flechon, J.E. and Wintenberger-Torres, S., 1981. Conceptus attachment in
602 the ewe: an ultrastructural study. *Placenta*, 2(2), pp.169-182.
- 603 27. Boshier, D.P., 1969. A histological and histochemical examination of implantation and early
604 placentome formation in sheep. *Reproduction*, 19(1), pp.51-61.

- 605 28. Guillomot, M. and Guay, P., 1982. Ultrastructural features of the cell surfaces of uterine and
606 trophoblastic epithelia during embryo attachment in the cow. *The Anatomical Record*,
607 204(4), pp.315-322.
- 608 29. Allen, W.R., Hamilton, D.W. and Moor, R.M., 1973. The origin of equine endometrial cups.
609 II. Invasion of the endometrium by trophoblast. *The Anatomical Record*, 177(4), pp.485-501.
- 610 30. Ealy, A.D. and Wooldridge, L.K., 2017. The evolution of interferon-tau. *Reproduction*,
611 154(5), pp.F1-F10.
- 612 31. Michael Roberts, R., 1989. Conceptus interferons and maternal recognition of pregnancy.
613 *Biology of reproduction*, 40(3), pp.449-452.
- 614 32. Martal, J., Lacroix, M.C., Loudes, C., Saunier, M. and Wintenberger-Torres, S., 1979.
615 Trophoblastin, an antiluteolytic protein present in early pregnancy in sheep. *Reproduction*,
616 56(1), pp.63-73.
- 617 33. Robinson, R.S., Fray, M.D., Wathes, D.C., Lamming, G.E. and Mann, G.E., 2006. In vivo
618 expression of interferon tau mRNA by the embryonic trophoblast and uterine concentrations
619 of interferon tau protein during early pregnancy in the cow. *Molecular reproduction and
620 development*, 73(4), pp.470-474.
- 621 34. Spencer, T.E., Becker, W.C., George, P., Mirando, M.A., Ogle, T.F. and Bazer, F.W., 1995.
622 Ovine interferon- τ regulates expression of endometrial receptors for estrogen and oxytocin
623 but not progesterone. *Biology of Reproduction*, 53(3), pp.732-745.
- 624 35. Oliveira, J.F., Henkes, L.E., Ashley, R.L., Purcell, S.H., Smirnova, N.P., Veeramachaneni,
625 D.R., Anthony, R.V. and Hansen, T.R., 2008. Expression of interferon (IFN)-stimulated
626 genes in extrauterine tissues during early pregnancy in sheep is the consequence of endocrine
627 IFN- τ release from the uterine vein. *Endocrinology*, 149(3), pp.1252-1259.
- 628 36. Bazer, F.W., Burghardt, R.C., Johnson, G.A., Spencer, T.E. and Wu, G., 2008. Interferons
629 and progesterone for establishment and maintenance of pregnancy: interactions among novel
630 cell signaling pathways. *Reproductive biology*, 8(3), pp.179-211.
- 631 37. Hansen, T.R., Henkes, L.K., Ashley, R.L., Bott, R.C., Antoniazzi, A.Q. and Han, H., 2010.
632 Endocrine actions of interferon-tau in ruminants.
- 633 38. Pervolaraki, K., Rastgou Talemi, S., Albrecht, D., Bormann, F., Bamford, C., Mendoza, J.L.,
634 Garcia, K.C., McLauchlan, J., Höfer, T., Stanifer, M.L. and Boulant, S., 2018. Differential

635 induction of interferon stimulated genes between type I and type III interferons is
636 independent of interferon receptor abundance. *PLoS pathogens*, 14(11), p.e1007420.

637 39. Forde, N., Beltman, M.E., Duffy, G.B., Duffy, P., Mehta, J.P., O'gaora, P., Roche, J.F.,
638 Lonergan, P. and Crowe, M.A., 2011. Changes in the endometrial transcriptome during the
639 bovine estrous cycle: effect of low circulating progesterone and consequences for conceptus
640 elongation. *Biology of reproduction*, 84(2), pp.266-278.

641 40. Endriß, K.J., Meyerholz, M.M., Fischbach, T., Brimmers, L., Pfarrer, C., Marth, C.D. and
642 Schmicke, M., 2021. In vitro effects of Type I interferons (IFN τ and IFN α) on bovine
643 hepatocytes cultured with or without Kupffer cells. *Reproduction, Fertility and Development*,
644 33(4), pp.305-317.

645 41. Sheikh, A.A., Hooda, O.K., Kalyan, A., Kamboj, A., Mohammed, S., Alhussien, M., Reddi,
646 S., Shimray, P.G., Rautela, A., Pandita, S. and Kapila, S., 2018. Interferon-tau stimulated
647 gene expression: A proxy to predict embryonic mortality in dairy cows. *Theriogenology*,
648 120, pp.61-67.

649 42. Shirasuna, K., Matsumoto, H., Matsuyama, S., Kimura, K., Bollwein, H. and Miyamoto, A.,
650 2015. Possible role of interferon tau on the bovine corpus luteum and neutrophils during the
651 early pregnancy. *Reproduction*, 150(3), pp.217-225.

652 43. Gifford, C.A., Racicot, K., Clark, D.S., Austin, K.J., Hansen, T.R., Lucy, M.C., Davies, C.J.
653 and Ott, T.L., 2007. Regulation of interferon-stimulated genes in peripheral blood leukocytes
654 in pregnant and bred, nonpregnant dairy cows. *Journal of Dairy Science*, 90(1), pp.274-280.

655 44. Matsuyama, S., Kojima, T., Kato, S. and Kimura, K., 2012. Relationship between quantity of
656 IFNT estimated by IFN-stimulated gene expression in peripheral blood mononuclear cells
657 and bovine embryonic mortality after AI or ET. *Reproductive Biology and Endocrinology*,
658 10(1), p.21.

659 45. Panda, B.S., Mohapatra, S.K., Chaudhary, D., Alhussien, M.N., Kapila, R. and Dang, A.K.,
660 2020. Proteomics and transcriptomics study reveals the utility of ISGs as novel molecules for
661 early pregnancy diagnosis in dairy cows. *Journal of Reproductive Immunology*, p.103148.
662 DOI: 10.1016/j.jri.2020.103148.

663 46. Ruhmann, B., Giller, K., Hankele, A.K., Ulbrich, S.E. and Schmicke, M., 2017. Interferon- τ
664 induced gene expression in bovine hepatocytes during early pregnancy. *Theriogenology*, 104,
665 pp.198-204.

- 666 47. Teixeira, M.G., Austin, K.J., Perry, D.J., Dooley, V.D., Johnson, G.A., Francis, B.R. and
667 Hansen, T.R., 1997. Bovine granulocyte chemotactic protein-2 is secreted by the
668 endometrium in response to interferon-tau (IFN- τ). *Endocrine*, 6(1), pp.31-37.
- 669 48. Mohammed, S., Alhussien, M.N., Ahmad Aljader, M., Kamboj, A., Gachuiwo Shimray, P.,
670 Ahmad Sheikh, A., Ial Yadav, M., Kumar Mohanty, A. and Kumar Dang, A., 2017.
671 Alteration in some pro and anti-inflammatory cytokines associated with complete and
672 incomplete gestation cycle of cows. *Biological Rhythm Research*, 48(6), pp.877-886.
- 673 49. Zhu, Z., Li, B., Wu, Y., Wang, X. and Deng, G., 2017. Interferon- τ increases BoLA-I for
674 implantation during early pregnancy in dairy cows. *Oncotarget*, 8(56), p.95095.
- 675 50. Bainbridge, D.R., 2000. Evolution of mammalian pregnancy in the presence of the maternal
676 immune system. *Reviews of Reproduction*, 5(2), pp.67-74.
- 677 51. Medawar, P.B., 1953. Some immunological and endocrinological problems raised by the
678 evolution of viviparity in vertebrates. In *Symp Soc Exp Biol* (Vol. 7, pp. 320-337).
- 679 52. Palmer, E.M. and Van Severen, G.A., 1997. Human T helper cell differentiation is regulated
680 by the combined action of cytokines and accessory cell-dependent costimulatory signals. *The*
681 *Journal of Immunology*, 158(6), pp.2654-2662.
- 682 53. Asnagli, H. and Murphy, K.M., 2001. Stability and commitment in T helper cell
683 development. *Current opinion in immunology*, 13(2), pp.242-247.
- 684 54. Del Prete, G., 1998. The concept of type-1 and type-2 helper T cells and their cytokines in
685 humans. *International reviews of immunology*, 16(3-4), pp.427-455.
- 686 55. Mohapatra, S.K., Panda, B.S., Verma, A.K., Kapila, R. and Dang, A.K., 2020. Implantation
687 associated changes in expression profile of indoleamine-2, 3-dioxygenase 1, Th1-Th2
688 cytokines and interferon-stimulated genes on neutrophils and peripheral blood mononuclear
689 cells of crossbred cows. *Journal of Reproductive Immunology*, p.103188.
- 690 56. O'Garra, A. and Arai, N., 2000. The molecular basis of T helper 1 and T helper 2 cell
691 differentiation. *Trends in cell biology*, 10(12), pp.542-550.
- 692 57. Kurt-Jones, E.A., Hamberg, S., Ohara, J., Paul, W.E. and Abbas, A.K., 1987. Heterogeneity
693 of helper/inducer T lymphocytes. I. Lymphokine production and lymphokine responsiveness.
694 *Journal of Experimental Medicine*, 166(6), pp.1774-1787.

- 695 58. Romagnani, S., 1992. Human TH1 and TH2 subsets: regulation of differentiation and role in
696 protection and immunopathology. *International archives of allergy and immunology*, 98(4),
697 pp.279-285.
- 698 59. Mucida, D. and Cheroutre, H., 2010. The many face-lifts of CD4 T helper cells. In *Advances*
699 *in immunology* (Vol. 107, pp. 139-152). Academic Press.
- 700 60. Oliveira, L.J., Mansourri-Attia, N., Fahey, A.G., Browne, J., Forde, N., Roche, J.F.,
701 Lonergan, P. and Fair, T., 2013. Characterization of the Th profile of the bovine
702 endometrium during the oestrous cycle and early pregnancy. *PLoS One*, 8(10), p.e75571.
- 703 61. Manjari, P., Kapoor, S., Hyder, I., De, S., Mohanty, A.K. and Dang, A.K., 2018. A study of
704 altered cytokine rhythms associated with successful implantation in cows. *Biological Rhythm*
705 *Research*, 49(2), pp.329-333.
- 706 62. Nishimura, R., Bowolaksono, A., Acosta, T.J., Murakami, S., Piotrowska, K., Skarzynski,
707 D.J. and Okuda, K., 2004. Possible role of interleukin-1 in the regulation of bovine corpus
708 luteum throughout the luteal phase. *Biology of reproduction*, 71(5), pp.1688-1693.
- 709 63. Tanikawa, M., Acosta, T.J., Fukui, T., Murakami, S., Korzekwa, A., Skarzynski, D.J.,
710 Piotrowska, K.K., Park, C.K. and Okuda, K., 2005. Regulation of prostaglandin synthesis by
711 interleukin-1 α in bovine endometrium during the estrous cycle. *Prostaglandins & Other Lipid*
712 *Mediators*, 78(1-4), pp.279-290.
- 713 64. Tanikawa, M., Lee, H.Y., Watanabe, K., Majewska, M., Skarzynski, D.J., Park, S.B., Lee,
714 D.S., Park, C.K., Acosta, T.J. and Okuda, K., 2008. Regulation of prostaglandin biosynthesis
715 by interleukin-1 in cultured bovine endometrial cells. *Journal of Endocrinology*, 199(3),
716 p.425.
- 717 65. Majewska, M., Woclawek-Potocka, I., Bah, M.M., Hapunik, J., Piotrowska, K.K., Tasaki, Y.,
718 Acosta, T.J., Okuda, K. and Skarzynski, D.J., 2010. Is interleukin-1? a luteotropic or
719 luteolytic agent in cattle?. *Reproduction*, 139(3), p.665.
- 720 66. Emond, V., Fortier, M.A., Murphy, B.D. and Lambert, R.D., 1998. Prostaglandin E2
721 regulates both interleukin-2 and granulocyte-macrophage colony-stimulating factor gene
722 expression in bovine lymphocytes. *Biology of reproduction*, 58(1), pp.143-151.
- 723 67. Ross, S.H. and Cantrell, D.A., 2018. Signaling and function of interleukin-2 in T
724 lymphocytes. *Annual review of immunology*, 36, pp.411-433.

- 725 68. Panda, B.S., Mohapatra, S.K., Verma, A.K., Kamboj, A., Alhussien, M.N. and Dang, A.K.,
726 2020. A comparative study on various immunological parameters influencing embryo
727 survivability in crossbred dairy cows. *Theriogenology*.
- 728 69. Seder, R.A., Paul, W.E., Davis, M.M. and Fazekas de St Groth, B., 1992. The presence of
729 interleukin 4 during in vitro priming determines the lymphokine-producing potential of
730 CD4+ T cells from T cell receptor transgenic mice. *The Journal of experimental medicine*,
731 176(4), pp.1091-1098.
- 732 70. Tuo, W., Macmillan, H., Gunter, N., Bazer, F.W. and Brown, W.C., 1999. Upregulation of
733 interleukin-4 and IFN-gamma expression by IFN-tau, a member of the type I IFN family.
734 *Journal of interferon & cytokine research*, 19(2), pp.179-187.
- 735 71. Maeda, Y., Ohtsuka, H., Tomioka, M. and Oikawa, M., 2013. Effect of progesterone on
736 Th1/Th2/Th17 and regulatory T cell-related genes in peripheral blood mononuclear cells
737 during pregnancy in cows. *Veterinary research communications*, 37(1), pp.43-49.
- 738 72. Wooldridge, L.K. and Ealy, A.D., 2019. Interleukin-6 increases inner cell mass numbers in
739 bovine embryos. *BMC developmental biology*, 19(1), p.2.
- 740 73. Blitek, A., Morawska, E. and Ziecik, A.J., 2012. Regulation of expression and role of
741 leukemia inhibitory factor and interleukin-6 in the uterus of early pregnant pigs.
742 *Theriogenology*, 78(5), pp.951-964.
- 743 74. Jiemtaweewoon, S., Shirasuna, K., Nitta, A., Kobayashi, A., Schuberth, H.J., Shimizu, T. and
744 Miyamoto, A., 2011. Evidence that polymorphonuclear neutrophils infiltrate into the
745 developing corpus luteum and promote angiogenesis with interleukin-8 in the cow.
746 *Reproductive Biology and Endocrinology*, 9(1), pp.1-10.
- 747 75. Sheikh, A.A., Hooda, O.K. and Dang, A.K., 2019. Interferon tau stimulated gene expression
748 and proinflammatory cytokine profile relative to insemination in dairy cows. *Biological
749 Rhythm Research*, 50(3), pp.335-345.
- 750 76. Ouyang, W., Rutz, S., Crellin, N.K., Valdez, P.A. and Hymowitz, S.G., 2011. Regulation and
751 functions of the IL-10 family of cytokines in inflammation and disease. *Annual review of
752 immunology*, 29, pp.71-109.
- 753 77. Shirasuna, K., Nitta, A., Sineenard, J., Shimizu, T., Bollwein, H. and Miyamoto, A., 2012.
754 Vascular and immune regulation of corpus luteum development, maintenance, and regression
755 in the cow. *Domestic Animal Endocrinology*, 43(2), pp.198-211.

- 756 78. Wegmann, T.G., Lin, H., Guilbert, L., Mosmann (pp. 353-356). TR. 1993. Bidirectional
757 cytokine interactions in the maternal-fetal relationship: is successful pregnancy a TH2
758 phenomenon.
- 759 79. Erdmann, A.A., Jung, U., Foley, J.E., Toda, Y. and Fowler, D.H., 2004. Co-stimulated/Tc2
760 cells abrogate murine marrow graft rejection. *Biology of Blood and Marrow Transplantation*,
761 10(9), pp.604-613.
- 762 80. Strom, T.B., Roy-Chaudhury, P., Manfro, R., Xheng, X.X., Nickerson, P.W., Wood, K. and
763 Bushell, A., 1996. The Th1/Th2 paradigm and the allograft response. *Current opinion in*
764 *immunology*, 8(5), pp.688-693.
- 765 81. Li, X.C., Zand, M.S., Li, Y., Zheng, X.X. and Strom, T.B., 1998. On histocompatibility
766 barriers, Th1 to Th2 immune deviation, and the nature of the allograft responses. *The Journal*
767 *of Immunology*, 161(5), pp.2241-2247.
- 768 82. Mao, G., Wang, J., Kang, Y., Tai, P., Wen, J., Zou, Q., Li, G., Ouyang, H., Xia, G. and
769 Wang, B., 2010. Progesterone increases systemic and local uterine proportions of CD4+
770 CD25+ Treg cells during midterm pregnancy in mice. *Endocrinology*, 151(11), pp.5477-
771 5488.
- 772 83. Walker, C.G., Meier, S., Littlejohn, M.D., Lehnert, K., Roche, J.R. and Mitchell, M.D., 2010.
773 Modulation of the maternal immune system by the pre-implantation embryo. *BMC*
774 *genomics*, 11(1), p.474.
- 775 84. Patel, D.D., Zachariah, J.P. and Whichard, L.P., 2001. CXCR3 and CCR5 ligands in
776 rheumatoid arthritis synovium. *Clinical immunology*, 98(1), pp.39-45.
- 777 85. Bulmer, J.N., Morrison, L., Longfellow, M., Ritson, A. and Pace, D., 1991. Granulated
778 lymphocytes in human endometrium: histochemical and immunohistochemical studies.
779 *Human reproduction*, 6(6), pp.791-798.
- 780 86. Lee, J.Y., Lee, M. and Lee, S.K., 2011. Role of endometrial immune cells in implantation.
781 *Clinical and experimental reproductive medicine*, 38(3), p.119.
- 782 87. Sentman, C.L., Meadows, S.K., Wira, C.R. and Eriksson, M., 2004. Recruitment of uterine
783 NK cells: induction of CXC chemokine ligands 10 and 11 in human endometrium by
784 estradiol and progesterone. *The Journal of Immunology*, 173(11), pp.6760-6766.
- 785 88. Dominguez, F., Galan, A., Martin, J.J.L., Remohi, J., Pellicer, A. and Simón, C., 2003.
786 Hormonal and embryonic regulation of chemokine receptors CXCR1, CXCR4, CCR5 and

787 CCR2B in the human endometrium and the human blastocyst. *Molecular human*
788 *reproduction*, 9(4), pp.189-198.

789 89. Meter, R.A., Wira, C.R. and Fahey, J.V., 2005. Secretion of monocyte chemotactic protein-1
790 by human uterine epithelium directs monocyte migration in culture. *Fertility and sterility*,
791 84(1), pp.191-201.

792 90. Hannan, N.J. and Salamonsen, L.A., 2007. Role of chemokines in the endometrium and in
793 embryo implantation. *Current Opinion in Obstetrics and Gynecology*, 19(3), pp.266-272.

794 91. Ishida, Y., Gao, J.L. and Murphy, P.M., 2008. Chemokine receptor CX3CR1 mediates skin
795 wound healing by promoting macrophage and fibroblast accumulation and function. *The*
796 *Journal of Immunology*, 180(1), pp.569-579.

797 92. Balaji, S., Watson, C.L., Ranjan, R., King, A., Bollyky, P.L. and Keswani, S.G., 2015.
798 Chemokine involvement in fetal and adult wound healing. *Advances in wound care*, 4(11),
799 pp.660-672.

800 93. Guerin, L.R., Prins, J.R. and Robertson, S.A., 2009. Regulatory T-cells and immune
801 tolerance in pregnancy: a new target for infertility treatment?. *Human reproduction update*,
802 15(5), pp.517-535.

803 94. Harnaha, J., Machen, J., Wright, M., Lakomy, R., Styche, A., Trucco, M., Makaroun, S. and
804 Giannoukakis, N., 2006. Interleukin-7 is a survival factor for CD4+ CD25+ T-cells and is
805 expressed by diabetes-suppressive dendritic cells. *Diabetes*, 55(1), pp.158-170.

806 95. Bauersachs, S., Ulbrich, S.E., Gross, K., Schmidt, S.E., Meyer, H.H., Wenigerkind, H.,
807 Vermehren, M., Sinowatz, F., Blum, H. and Wolf, E., 2006. Embryo-induced transcriptome
808 changes in bovine endometrium reveal species-specific and common molecular markers of
809 uterine receptivity. *Reproduction*, 132(2), pp.319-331.

810 96. Mitko, K., Ulbrich, S.E., Wenigerkind, H., Sinowatz, F., Blum, H., Wolf, E. and Bauersachs,
811 S., 2008. Focus on Mammalian Embryogenomics Dynamic changes in messenger RNA
812 profiles of bovine endometrium during the oestrous cycle. *Reproduction*, 135, pp.225-240.

813 97. Mansouri-Attia, N., Sandra, O., Aubert, J., Degrelle, S., Everts, R.E., Giraud-Delville, C.,
814 Heyman, Y., Galio, L., Hue, I., Yang, X. and Tian, X.C., 2009. Endometrium as an early
815 sensor of in vitro embryo manipulation technologies. *Proceedings of the National Academy*
816 *of Sciences*, 106(14), pp.5687-5692.

- 817 98. Spencer, T.E., Gray, A., Johnson, G.A., Taylor, K.M., Gertler, A., Gootwine, E., Ott, T.L.
818 and Bazer, F.W., 1999. Effects of recombinant ovine interferon tau, placental lactogen, and
819 growth hormone on the ovine uterus. *Biology of Reproduction*, 61(6), pp.1409-1418.
- 820 99. McAveney, K.M., Book, M.L., Ling, P., Chebath, J. and Yu-Lee, L.Y., 2000. Association of
821 2', 5'-oligoadenylate synthetase with the prolactin (PRL) receptor: alteration in PRL-
822 inducible stat1 (signal transducer and activator of transcription 1) signaling to the IRF-1
823 (interferon-regulatory factor 1) promoter. *Molecular Endocrinology*, 14(2), pp.295-306.
- 824 100. Johnson, G.A., Burghardt, R.C., Bazer, F.W. and Spencer, T.E., 2003. Osteopontin: roles
825 in implantation and placentation. *Biology of reproduction*, 69(5), pp.1458-1471.
- 826 101. Hicks, B.A., Etter, S.J., Carnahan, K.G., Joyce, M.M., Assiri, A.A., Carling, S.J., Kodali,
827 K., Johnson, G.A., Hansen, T.R., Mirando, M.A. and Woods, G.L., 2003. Expression of the
828 uterine Mx protein in cyclic and pregnant cows, gilts, and mares. *Journal of animal science*,
829 81(6), pp.1552-1561.
- 830 102. Yang, Y., Lee, J.H., Kim, K.Y., Song, H.K., Kim, J.K., Yoon, S.R., Cho, D., Song, K.S.,
831 Lee, Y.H. and Choi, I., 2005. The interferon-inducible 9-27 gene modulates the susceptibility
832 to natural killer cells and the invasiveness of gastric cancer cells. *Cancer letters*, 221(2),
833 pp.191-200.
- 834 103. Kalkunte, S.S., Mselle, T.F., Norris, W.E., Wira, C.R., Sentman, C.L. and Sharma, S.,
835 2009. Vascular endothelial growth factor C facilitates immune tolerance and endovascular
836 activity of human uterine NK cells at the maternal-fetal interface. *The Journal of*
837 *Immunology*, 182(7), pp.4085-4092.
- 838 104. Stone, T.W. and Darlington, L.G., 2002. Endogenous kynurenines as targets for drug
839 discovery and development. *Nature reviews Drug discovery*, 1(8), pp.609-620.
- 840 105. Kudo, Y., Boyd, C.A.R., Spyropoulou, I., Redman, C.W.G., Takikawa, O., Katsuki, T.,
841 Hara, T., Ohama, K. and Sargent, I.L., 2004. Indoleamine 2, 3-dioxygenase: distribution and
842 function in the developing human placenta. *Journal of reproductive immunology*, 61(2),
843 pp.87-98.
- 844 106. Von Rango, U., Krusche, C.A., Beier, H.M. and Classen-Linke, I., 2007. Indoleamine-
845 dioxygenase is expressed in human decidua at the time maternal tolerance is established.
846 *Journal of reproductive immunology*, 74(1-2), pp.34-45.

- 847 107. Ball, H.J., Sanchez-Perez, A., Weiser, S., Austin, C.J., Astelbauer, F., Miu, J.,
848 McQuillan, J.A., Stocker, R., Jermiin, L.S. and Hunt, N.H., 2007. Characterization of an
849 indoleamine 2, 3-dioxygenase-like protein found in humans and mice. *Gene*, 396(1), pp.203-
850 213.
- 851 108. Metz, R., DuHadaway, J.B., Kamasani, U., Laury-Kleintop, L., Muller, A.J. and
852 Prendergast, G.C., 2007. Novel tryptophan catabolic enzyme IDO2 is the preferred
853 biochemical target of the antitumor indoleamine 2, 3-dioxygenase inhibitory compound D-1-
854 methyl-tryptophan. *Cancer research*, 67(15), pp.7082-7087.
- 855 109. Ball, H.J., Yuasa, H.J., Austin, C.J., Weiser, S. and Hunt, N.H., 2009. Indoleamine 2, 3-
856 dioxygenase-2; a new enzyme in the kynurenine pathway. *The international journal of*
857 *biochemistry & cell biology*, 41(3), pp.467-471.
- 858 110. Miwa, N., Hayakawa, S., Miyazaki, S., Myojo, S., Sasaki, Y., Sakai, M., Takikawa, O.
859 and Saito, S., 2005. IDO expression on decidual and peripheral blood dendritic cells and
860 monocytes/macrophages after treatment with CTLA-4 or interferon- γ increase in normal
861 pregnancy but decrease in spontaneous abortion. *Molecular human reproduction*, 11(12),
862 pp.865-870.
- 863 111. Mezrich, J.D., Fechner, J.H., Zhang, X., Johnson, B.P., Burlingham, W.J. and Bradfield,
864 C.A., 2010. An interaction between kynurenine and the aryl hydrocarbon receptor can
865 generate regulatory T cells. *The Journal of Immunology*, 185(6), pp.3190-3198.
- 866 112. Jeong, Y.I., Kim, S.W., Jung, I.D., Lee, J.S., Chang, J.H., Lee, C.M., Chun, S.H., Yoon,
867 M.S., Kim, G.T., Ryu, S.W. and Kim, J.S., 2009. Curcumin suppresses the induction of
868 indoleamine 2, 3-dioxygenase by blocking the Janus-activated kinase-protein kinase C δ -
869 STAT1 signaling pathway in interferon- γ -stimulated murine dendritic cells. *Journal of*
870 *Biological Chemistry*, 284(6), pp.3700-3708.
- 871 113. Cheng, C.W., Shieh, P.C., Lin, Y.C., Chen, Y.J., Lin, Y.H., Kuo, D.H., Liu, J.Y., Kao,
872 J.Y., Kao, M.C. and Way, T.D., 2010. Indoleamine 2, 3-dioxygenase, an immunomodulatory
873 protein, is suppressed by (-)-epigallocatechin-3-gallate via blocking of γ -interferon-induced
874 JAK-PKC- δ -STAT1 signaling in human oral cancer cells. *Journal of agricultural and food*
875 *chemistry*, 58(2), pp.887-894.
- 876 114. Boyt, D.T., Boland, L.K., Burand Jr, A.J., Brown, A.J. and Ankrum, J.A., 2020. Dose and
877 duration of interferon γ pre-licensing interact with donor characteristics to influence the

878 expression and function of indoleamine-2, 3-dioxygenase in mesenchymal stromal cells.
879 Journal of the Royal Society Interface, 17(167), p.20190815.

880 115. Munn, D.H., Zhou, M., Attwood, J.T., Bondarev, I., Conway, S.J., Marshall, B., Brown,
881 C. and Mellor, A.L., 1998. Prevention of allogeneic fetal rejection by tryptophan catabolism.
882 Science, 281(5380), pp.1191-1193.

883 116. Mellor, A.L. and Munn, D.H., 2001. Tryptophan catabolism prevents maternal T cells
884 from activating lethal anti-fetal immune responses. Journal of reproductive immunology,
885 52(1-2), pp.5-13.

886 117. Mellor, A.L. and Munn, D.H., 1999. Tryptophan catabolism and T-cell tolerance:
887 immunosuppression by starvation?. Immunology today, 20(10), pp.469-473.

888 118. Munn, D.H., Shafizadeh, E., Attwood, J.T., Bondarev, I., Pashine, A. and Mellor, A.L.,
889 1999. Inhibition of T cell proliferation by macrophage tryptophan catabolism. The Journal of
890 experimental medicine, 189(9), pp.1363-1372.

891 119. Lee, G.K., Park, H.J., Macleod, M., Chandler, P., Munn, D.H. and Mellor, A.L., 2002.
892 Tryptophan deprivation sensitizes activated T cells to apoptosis prior to cell division.
893 Immunology, 107(4), pp.452-460.

894 120. Mellor, A.L., Chandler, P., Baban, B., Hansen, A.M., Marshall, B., Pihkala, J.,
895 Waldmann, H., Cobbold, S., Adams, E. and Munn, D.H., 2004. Specific subsets of murine
896 dendritic cells acquire potent T cell regulatory functions following CTLA4-mediated
897 induction of indoleamine 2, 3 dioxygenase. International immunology, 16(10), pp.1391-
898 1401.

899 121. Fallarino, F., Grohmann, U., You, S., McGrath, B.C., Cavener, D.R., Vacca, C., Orabona,
900 C., Bianchi, R., Belladonna, M.L., Volpi, C. and Santamaria, P., 2006. The combined effects
901 of tryptophan starvation and tryptophan catabolites down-regulate T cell receptor ζ -chain and
902 induce a regulatory phenotype in naive T cells. The Journal of Immunology, 176(11),
903 pp.6752-6761.

904 122. Eleftheriadis, T., Pissas, G., Yiannaki, E., Markala, D., Arampatzis, S., Antoniadis, G.,
905 Liakopoulos, V. and Stefanidis, I., 2013. Inhibition of indoleamine 2, 3-dioxygenase in
906 mixed lymphocyte reaction affects glucose influx and enzymes involved in aerobic
907 glycolysis and glutaminolysis in alloreactive T-cells. Human immunology, 74(12), pp.1501-
908 1509.

- 909 123. Eleftheriadis, T., Pissas, G., Antoniadi, G., Spanoulis, A., Liakopoulos, V. and Stefanidis,
910 I., 2014. Indoleamine 2, 3-dioxygenase increases p53 levels in alloreactive human T cells,
911 and both indoleamine 2, 3-dioxygenase and p53 suppress glucose uptake, glycolysis and
912 proliferation. *International immunology*, 26(12), pp.673-684.
- 913 124. Terness, P., Kallikourdis, M., Betz, A.G., Rabinovich, G.A., Saito, S. and Clark, D.A.,
914 2007. Tolerance signaling molecules and pregnancy: IDO, galectins, and the renaissance of
915 regulatory T cells. *American Journal of Reproductive Immunology*, 58(3), pp.238-254.
- 916 125. Badawy, A.A.B., Namboodiri, A.M. and Moffett, J.R., 2016. The end of the road for
917 the tryptophan depletion concept in pregnancy and infection. *Clinical Science*, 130(15),
918 pp.1327-1333.
- 919 126. Opitz, C.A., Litzenburger, U.M., Sahm, F., Ott, M., Tritschler, I., Trump, S.,
920 Schumacher, T., Jestaedt, L., Schrenk, D., Weller, M. and Jugold, M., 2011. An endogenous
921 tumour-promoting ligand of the human aryl hydrocarbon receptor. *Nature*, 478(7368),
922 pp.197-203.
- 923 127. Liu, Y., Liang, X., Yin, X., Lv, J., Tang, K., Ma, J., Ji, T., Zhang, H., Dong, W., Jin, X.
924 and Chen, D., 2017. Blockade of IDO-kynurenine-AhR metabolic circuitry abrogates IFN- γ -
925 induced immunologic dormancy of tumor-repopulating cells. *Nature communications*, 8(1),
926 pp.1-15.
- 927 128. Nguyen, N.T., Kimura, A., Nakahama, T., Chinen, I., Masuda, K., Nohara, K., Fujii-
928 Kuriyama, Y. and Kishimoto, T., 2010. Aryl hydrocarbon receptor negatively regulates
929 dendritic cell immunogenicity via a kynurenine-dependent mechanism. *Proceedings of the*
930 *National Academy of Sciences*, 107(46), pp.19961-19966.
- 931 129. Jaronen, M. and Quintana, F.J., 2014. Immunological relevance of the coevolution of
932 IDO1 and AHR. *Frontiers in immunology*, 5, p.521.
- 933 130. Fazio, F., Lionetto, L., Curto, M., Iacovelli, L., Copeland, C.S., Neale, S.A., Bruno, V.,
934 Battaglia, G., Salt, T.E. and Nicoletti, F., 2017. Cinnabarinic acid and xanthurenic acid: Two
935 kynurenine metabolites that interact with metabotropic glutamate receptors.
936 *Neuropharmacology*, 112, pp.365-372.
- 937 131. Nguyen, N.T., Nakahama, T., Le, D.H., Van Son, L., Chu, H.H. and Kishimoto, T.,
938 2014. Aryl hydrocarbon receptor and kynurenine: recent advances in autoimmune disease
939 research. *Frontiers in immunology*, 5, p.551.

- 940 132. Fallarino, F., Grohmann, U., Vacca, C., Bianchi, R., Orabona, C., Spreca, A., Fioretti,
941 M.C. and Puccetti, P., 2002. T cell apoptosis by tryptophan catabolism. *Cell Death &*
942 *Differentiation*, 9(10), pp.1069-1077.
- 943 133. Hayashi, T., Mo, J.H., Gong, X., Rossetto, C., Jang, A., Beck, L., Elliott, G.I.,
944 Kufareva, I., Abagyan, R., Broide, D.H. and Lee, J., 2007. 3-Hydroxyanthranilic acid inhibits
945 PDK1 activation and suppresses experimental asthma by inducing T cell apoptosis.
946 *Proceedings of the National Academy of Sciences*, 104(47), pp.18619-18624.
- 947 134. Lee, S.M., Lee, Y.S., Choi, J.H., Park, S.G., Choi, I.W., Joo, Y.D., Lee, W.S., Lee,
948 J.N., Choi, I. and Seo, S.K., 2010. Tryptophan metabolite 3-hydroxyanthranilic acid
949 selectively induces activated T cell death via intracellular GSH depletion. *Immunology*
950 *letters*, 132(1-2), pp.53-60.
- 951 135. Zaher, S.S., Germain, C., Fu, H., Larkin, D.F. and George, A.J., 2011. 3-
952 hydroxykynurenine suppresses CD4+ T-cell proliferation, induces T-regulatory-cell
953 development, and prolongs corneal allograft survival. *Investigative ophthalmology & visual*
954 *science*, 52(5), pp.2640-2648.
- 955 136. Prodinger, J., Loacker, L.J., Schmidt, R.L., Ratzinger, F., Greiner, G., Witzeneder, N.,
956 Hoermann, G., Jutz, S., Pickl, W.F., Steinberger, P. and Marculescu, R., 2016. The
957 tryptophan metabolite picolinic acid suppresses proliferation and metabolic activity of CD4+
958 T cells and inhibits c-Myc activation. *Journal of leukocyte biology*, 99(4), pp.583-594.
- 959 137. Dong, M., Ding, G., Zhou, J., Wang, H., Zhao, Y. and Huang, H., 2008. The effect of
960 trophoblasts on T lymphocytes: possible regulatory effector molecules-a proteomic analysis.
961 *Cellular Physiology and Biochemistry*, 21(5-6), pp.463-472.
- 962 138. Li, X., Gui, S. and Wang, H., 2007. Effect of Kidney-replenishing herb on the
963 indoleamine 2, 3-dioxygenase of human syncytiotrophoblasts cultured in vitro and the
964 balance of helper T-cell cytokines. *Gynecological Endocrinology*, 23(11), pp.653-661.
- 965 139. Zong, S., Li, C., Luo, C., Zhao, X., Liu, C., Wang, K., Jia, W., Bai, M., Yin, M., Bao,
966 S. and Guo, J., 2016. Dysregulated expression of IDO may cause unexplained recurrent
967 spontaneous abortion through suppression of trophoblast cell proliferation and migration.
968 *Scientific reports*, 6, p.19916.
- 969 140. Mei, J., Xie, X.X., Li, M.Q., Wei, C.Y., Jin, L.P., Li, D.J. and Zhu, X.Y., 2014.
970 Indoleamine 2, 3-dioxygenase-1 (IDO1) in human endometrial stromal cells induces

971 macrophage tolerance through interleukin-33 in the progression of endometriosis.
972 International journal of clinical and experimental pathology, 7(6), p.2743.

973 141. Croxatto, D., Vacca, P., Canegallo, F., Conte, R., Venturini, P.L., Moretta, L. and
974 Mingari, M.C., 2014. Stromal cells from human decidua exert a strong inhibitory effect on
975 NK cell function and dendritic cell differentiation. PloS one, 9(2).

976 142. Ban, Y., Chang, Y., Dong, B., Kong, B. and Qu, X., 2013. Indoleamine 2, 3-
977 dioxygenase levels at the normal and recurrent spontaneous abortion fetal–maternal interface.
978 Journal of international medical research, 41(4), pp.1135-1149.

979 143. Liu, X., Liu, Y., Ding, M. and Wang, X., 2011. Reduced expression of indoleamine 2,
980 3-dioxygenase participates in pathogenesis of preeclampsia via regulatory T cells. Molecular
981 medicine reports, 4(1), pp.53-58.

982

983

984 **Captions:**

985 **Figure-1:** Schematic representation of Th1-Th2 cytokine balance during pregnancy.

986

987 **Figure-2:** Schematic representation of indoleamine 2,3 dioxygenase (IDO) mediated maternal
988 immunosuppression in bovines.

989

990 **Table-1:** Gestation length, placentation days, agents of MRP and day of production of MRP
991 agents in different species

992