**A New Era in Agricultural Microbiology: SCAR Marker Validation**

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**Abstract**

The profound significance of agricultural microbiology in shaping sustainable and efficient farming systems has been increasingly recognized over the past few decades. Central to this evolution is the deployment of Sequence Characterized Amplified Region (SCAR) markers, ushering in transformative advancements in microbial authentication and validation processes. This chapter delves into the revolutionary role of SCAR markers, spotlighting their pivotal position in the modern agricultural microbiology landscape. SCAR markers, derived from Random Amplified Polymorphic DNA (RAPD) markers, are renowned for their reproducibility and specificity across varied conditions. This distinctive nature allows for precise identification of agriculturally significant microorganisms, a cornerstone in modern biotechnological applications. The significance extends to plant breeding, pathogen detection, and the authentication of beneficial microbes in biofertilizers and biopesticides. Technological strides have refined SCAR marker validation processes, integrating high-throughput sequencing platforms and advanced bioinformatics tools. These innovations accelerate the identification and validation of SCAR markers, expediting microbial diagnostics and enhancing the accuracy of molecular interventions. Moreover, the review underscores the intertwined progression of SCAR marker developments and the overarching evolution of digital agriculture. The fusion of SCAR marker data with emerging agricultural tools, such as remote sensing and data analytics, promises a future where farming decisions are underpinned by nuanced molecular insights. In summary, the validation of SCAR markers signifies a transformative phase in agricultural microbiology, providing unmatched accuracy in microbial identification. With the escalating challenges of worldwide agriculture, these molecular instruments become crucial in forging a path toward sustainability and enhanced productivity.

**Key words:** Bioinoculants, SCAR markers, strain validation, RAPD

1. **Introduction:**

Agricultural microbiology is a specialized branch of microbiology focused on understanding the intricate relationships between microorganisms and their environment in agricultural contexts. Since the inception of agriculture, microorganisms have been vital contributors to soil health, plant growth and overall crop productivity. These organisms facilitate key processes such as nutrient cycling and soil structure formation and they establish vital symbiotic relationships with plants. From nitrogen-fixing bacteria to mycorrhizal fungi, these interactions form the backbone of a supportive environment, enabling plants to flourish even under challenging conditions (Ray et al., 2020). Beneficial microorganisms can elevate plant disease resistance, augment soil fertility, and even enhance the nutritional profile of crops (Dixon & Tilston, 2010). However, the presence of pathogenic microorganisms poses threats, potentially leading to significant crop damage and consequent economic losses (Singh etal.,2021). Understanding these beneficial and harmful microorganisms has always been at the forefront of agricultural microbiology, ensuring better crop productivity, disease management, and sustainable farming practices.

In the realm of modern agriculture, the utilization of authenticated microbial strains is paramount for achieving targeted outcomes, be it soil enhancement, promoting plant growth, or fortifying disease resistance (Anand et al., 2022). However, the journey towards these outcomes isn't without challenges. Missteps in microbial identification or contamination issues can inadvertently usher harmful pathogens into the agricultural environment. This not only endangers crops and soil but can have broader implications on human health (Bintsis, 2018). From a regulatory standpoint, especially concerning commercial microbial products, authentication isn't just recommended; it's a requisite. Precise identification assures compliance with standards and bolsters consumer confidence in the product's reliability (Ratajczak etal., 2015). Scientifically, the consequences of misidentification can ripple across the research landscape, distorting findings and eroding trust in scholarly work. As the appetite for biologically-informed agricultural solutions grows, the industry acknowledges the indispensable role of robust authentication. Advanced tools, such as SCAR markers, exemplify the innovations propelling this new chapter in agricultural microbiology (Reddypriya etal., 2019).

1. **Overview of SCAR Markers**

In the constantly evolving field of molecular biology, specific tools and techniques emerge that revolutionize our understanding of organisms and their interactions. Among these tools, Sequence Characterized Amplified Region (SCAR) markers have surfaced as a pivotal component in various biological studies, most notably in the domain of plant genetics and agricultural microbiology. SCAR markers, distinct from random amplified polymorphic DNA (RAPD) markers, are DNA fragments from PCR-amplified RAPD that are genetically linked to a trait of interest. These markers are highly reproducible, unlike RAPD markers, which can sometimes exhibit inconsistency due to their sensitivity to PCR conditions. SCAR markers are unique because of their sequence-specific nature, meaning they are specific to particular sequences in the DNA, ensuring their reliability. A significant advantage of SCAR markers lies in their co-dominant inheritance, offering a clear distinction between homozygous and heterozygous states in genetic mapping (Kesawat et al., 2009). Such precision is vital when tracing specific traits within populations, making SCAR markers invaluable for breeding programs and trait-specific cultivar development. Moreover, SCAR markers have transformed the landscape of plant pathogen studies. As elucidated by Yang et al., (2018), they have been extensively used to detect plant pathogens, given their high specificity. By enabling early and accurate detection, these markers contribute enormously to disease management and preventive agricultural practices. Beyond plant sciences, SCAR markers have also found utility in the realm of agricultural microbiology. With the increasing emphasis on harnessing beneficial microorganisms for sustainable farming, the authentication of microbial strains becomes essential. As noted by Ambreetha & Balachandar (2023), SCAR markers have been employed for the precise validation of microbial compositions, ensuring that the right microbes are introduced to agricultural systems. With the progression of technology and the increasing needs of precision agriculture and molecular biology, SCAR markers are poised to consistently lead the way, steering research and methodologies with their steadfast precision.

1. **Mechanism of SCAR Marker Validation**
	1. **Genetic Basis of SCAR Marker Functionality**

The understanding of molecular genetics has made vast strides with the advent of specific tools like the Sequence Characterized Amplified Region (SCAR) markers. To fully appreciate the essence of SCAR markers, one must delve into their genetic foundation, which dictates their functionality and importance in molecular biology and agriculture (Smith & Jones, 2000). At its core, the SCAR marker's genesis is rooted in another well-known molecular marker: Random Amplified Polymorphic DNA (RAPD). RAPDs, while valuable, are prone to variability due to their non-specific nature and sensitivity to PCR conditions. Yet, it was observed that certain RAPD markers consistently correlated with specific traits. Such markers were sequenced and converted into SCAR markers, a process that transformed their inherent variability into a reproducible and specific genetic tool (Bhagyawant, 2016). The resulting SCAR marker retains the genetic link to the trait, but with the added benefit of sequence specificity. Functionally, SCAR markers have an edge due to their co-dominant nature. Co-dominance in genetic markers is a unique trait that allows the detection of both homozygous and heterozygous states within organisms (Amiteye, 2021). This quality is of utmost importance when mapping genes or identifying specific traits within populations. SCAR markers, given their origin from known sequences, exhibit this co-dominance with a high degree of accuracy.

The genetic basis of SCAR markers offers them a specificity that is unparalleled among many molecular markers. While other markers might give a broader view of an organism's genome, SCAR markers are like precision tools, targeting specific genes or gene regions (Kiran et al., 2010). This property is a direct consequence of their origin from known, sequenced DNA fragments. In practical applications, this specificity ensures that SCAR markers can reliably detect the presence or absence of particular genetic elements, be it for disease resistance in plants or specific traits in microbial communities. Additionally, their stable and consistent nature makes SCAR markers an invaluable asset in breeding programs. As breeders aim to incorporate specific traits into new plant varieties, SCAR markers can swiftly identify plants that possess the desired genetic elements, streamlining the breeding process and ensuring successful trait incorporation (Singh et al., 2014). Beyond that, their reliability eliminates much of the guesswork and reduces the chances of unwanted genetic elements being introduced.

Another notable aspect is the adaptability of SCAR markers. With advances in genomics and bioinformatics, SCAR markers can be designed and utilized for a plethora of organisms and purposes, reflecting their flexibility rooted in their genetic foundation (Sharma et al., 2014). Hence, the genetic foundation of SCAR markers ensures they serve as precise, reliable, and versatile tools in a myriad of applications, from basic research to practical breeding programs. As our understanding of molecular genetics continues to expand, the role of SCAR markers, underpinned by their unique genetic basis, will undoubtedly remain pivotal.

**3.2 Methodology and Procedures for SCAR Marker Validation**

The genesis of the SCAR marker starts with pinpointing a potential Random Amplified Polymorphic DNA (RAPD) marker. This marker should exhibit consistent correlation with a desired trait. Once identified, the RAPD marker undergoes sequencing that helps in understanding the specific DNA segments associated with the trait in question, which is the first crucial step in the validation process. Following sequencing, primers specific to the sequenced RAPD marker are designed. These primers, typically 18-25 base pairs in length, flank the identified sequence and are utilized in PCR amplification (Yang et al., 2013). The amplified product, now termed the SCAR marker, should consistently correlate with the trait, confirming its utility.

Validation further involves assessing the SCAR marker's performance across a diverse range of samples. This helps in ascertaining its robustness and reliability. Cross-referencing the SCAR marker's presence or absence with the desired trait across these samples serves as a confirmation of its validity (Gupta et al., 2006). In breeding programs, for instance, this would involve analyzing multiple plant samples to ensure the SCAR marker consistently identifies the desired trait. Moreover, external factors can influence SCAR marker performance, like PCR conditions or DNA quality (Shah et al., 2023). Therefore, part of the validation procedure is to establish standard protocols that stipulate ideal conditions for consistent SCAR marker amplification.

Lastly, a validated SCAR marker should be reproducible across different labs and under varying conditions, affirming its reliability. This often involves collaborative studies where multiple laboratories independently verify the SCAR marker's performance (Liu et al., 2020). Through a combination of sequencing, specific primer design, extensive cross-referencing, and collaborative verification, SCAR markers are rigorously validated, paving the way for their application in diverse genetic endeavors.

1. **Applications of SCAR Markers in Agriculture**

**4.1. SCAR Markers in Crop Protection and Disease Management**

At the heart of crop protection lies the need for early detection of pathogens and pests. SCAR markers, with their high specificity, are adept at identifying specific strains of pathogens at early infection stages, allowing for timely interventions (Paran & Michel, 1993). For instance, in the battle against fungal pathogens in wheat, SCAR markers have been used to detect the presence of harmful rust species even before visible symptoms manifest on the plant (Gupta et al., 2006). Beyond detection, SCAR markers play a crucial role in breeding programs focused on disease resistance. Once a disease-resistance trait is identified within a plant species or variety, SCAR markers associated with that trait can be used to expedite the breeding process. By swiftly identifying plants that carry the resistance genes, breeders can more efficiently develop disease-resistant crop varieties, ensuring crop protection from prevalent diseases (Kasai et al., 2000; Iruela etal.,2006). Additionally, in integrated pest management (IPM) strategies, SCAR markers offer a nuanced understanding of pest populations. By differentiating between pest strains, they can inform the deployment of targeted interventions, thereby minimizing the use of broad-spectrum pesticides and promoting environmentally friendly pest control (Garcia et al.,2007). In addition, SCAR markers have a potential role in monitoring disease evolution. As pathogens evolve, their genetic makeup may change, leading to new strains that might bypass current resistance mechanisms in crops. SCAR markers can track these genetic shifts, helping agriculturalists anticipate and prepare for potential future threats (Babu et al., 2020). As challenges in crop protection persist, SCAR markers will lead the way in crafting strategies for sustainable and robust agriculture.

**4.2 Validating Beneficial Microorganisms for Soil Health**

Soil health is a cornerstone of sustainable agriculture. Maintaining a balanced microbial ecosystem within the soil not only fosters plant growth but also fortifies the soil against pathogens and degradation. One of the primary challenges in harnessing beneficial microorganisms lies in their identification and validation. With an array of microbial species present in the soil, distinguishing beneficial microbes from potential pathogens or neutral microbes is paramount. SCAR markers, given their specificity, provide a precise means to achieve this (Paran & Michel, 1993). By targeting unique genetic sequences associated with known beneficial traits, SCAR markers can rapidly and reliably confirm the presence of beneficial microbes. The introduction of beneficial microbial formulations, such as biofertilizers and biopesticides, into the soil requires stringent validation to ensure safety and efficacy. SCAR markers, through their targeted identification mechanism, can validate these formulations, confirming the presence of intended beneficial microbes and the absence of contaminants (Reddypriya et al., 2016).

Additionally, monitoring the persistence and proliferation of introduced beneficial microbes in the soil is essential for understanding their long-term impact on soil health. SCAR markers offer a non-invasive method to track these microbes over time, shedding light on their interactions with native soil communities and their longevity in promoting soil health (Walker & Bais, 2018). Furthermore, in research settings, SCAR markers assist in deciphering the complex soil-microbe-plant interactions. As researchers probe the synergistic relationships that beneficial microbes have with plants, SCAR markers help pinpoint specific microbial strains involved, aiding in the development of improved microbial formulations for soil health (Holmberg et al., 2009; Aggarwal et al., 2011). By facilitating the validation, introduction, and monitoring of beneficial microorganisms, these markers ensure that we harness the full potential of microbial allies. For exploration into the soil's microbiome intensifies, SCAR markers are poised to be instrumental in forging a sustainable path for agriculture.

**4.3 Role in Authenticating Biofertilizers and Biopesticides**

In the pursuit of sustainable agriculture, biofertilizers and biopesticides have emerged as potent alternatives to traditional chemical inputs, promising both environmental benefits and enhanced crop yields. However, the successful deployment of these bio-based solutions hinges on their authenticity and effectiveness. In this context, SCAR markers have proven to be pivotal molecular tools that play a transformative role in authenticating biofertilizers and biopesticides, ensuring their reliability and impact (Nunes et al., 2008; Gotor‐Vila et al., 2016; Reddypriya & Gopalaswamy, 2017).

Biofertilizers, containing live cells of beneficial microorganisms, have redefined soil health management. They bolster plant growth by enhancing nutrient availability, promoting root development, and fostering symbiotic relationships. Authenticating the microbial strains within these preparations becomes paramount to ensure that they perform their intended roles effectively. SCAR markers, known for their sequence specificity, have provided a reliable means to confirm the identity of these beneficial microbes. By targeting unique genetic sequences associated with known beneficial traits, these markers ensure that biofertilizer products contain the appropriate strains, validating their potential in improving soil fertility. In Maize and Rice crops, nitrogen-fixing bacteria of the Azospirillum genus present in rhizoplane or as a endosymbiont, aiding in nitrogen fixation. Reddypriya eta al., (2016) conducted a study utilizing SCAR markers to authenticate Azosprillum strains in commercial biofertilizer products. The markers discerned between various Azospirillum species, confirming the presence of the appropriate strain in biofertilizer formulations. This SCAR-based authentication not only optimized nitrogen fixation but also upheld crop yield and soil fertility.

Derived from living entities, biopesticides have risen as sustainable solutions to combat a plethora of agricultural pests. By acting against pests either through direct toxicity, parasitism, or other mechanisms, they offer targeted pest management with minimal environmental impact. The complexity of these biological agents requires precise authentication mechanisms. SCAR markers, due to their high specificity, can discern between different strains of a microbial species. This precision ensures that the biopesticides contain the right microbial strain to target specific pests effectively. The fine-tuned identification afforded by SCAR markers ensures that biopesticides work optimally without inadvertently harming beneficial fauna or the environment (Perez et al., 2014).

Beyond the initial validation, SCAR markers have a role in monitoring and quality control. The agricultural market, awash with numerous bio-based products, necessitates consistent quality checks to maintain product integrity. SCAR markers offer a consistent method to ensure that these bio-products, over time and across batches, maintain their microbial consistency, meeting both regulatory standards and the expectations of farmers (Reddypriya et al, 2016). *Beauveria bassiana*, a bacterium which is widely used as biopesticides to combat Colorado potato beetle and other agricultural insect pests. Castrillo et al., (2003) employed SCAR markers to validate *B.bassiana* strains within commercial biopesticide products. By confirming the presence of the precise *B.bassiana* strain targeting the intended pest, SCAR markers ensured the efficacy of the biopesticide, minimizing collateral damage to non-target species and reducing environmental impact. Additionally, the global dialogue on sustainable agriculture often touches upon the environmental implications of introducing new microbial strains into diverse ecosystems. Here, SCAR markers provide a robust framework for monitoring the environmental behavior of introduced beneficial microorganisms, enabling researchers to understand their ecological interactions and long-term impact on native ecosystems.

**5.0 Challenges and Limitations of SCAR Markers**

SCAR markers, arising from specific DNA fragments, are known for their reproducibility and specificity. However, technical and methodological challenges necessitate careful consideration. The intricacy of their development involves transforming Random Amplified Polymorphic DNA (RAPD) markers into SCAR markers, which demands accurate sequencing prone to errors due to impurities . Primer design significantly impacts SCAR marker efficiency. Incorrect primers can lead to non-specific amplification or failed reactions, complicating diagnostic processes. Reproducibility relies on primer quality consistency (Arumingtyas et al., 2022). Despite robustness, SCAR markers are sensitive to PCR conditions, necessitating rigorous optimization for consistent results (Horejsi, 1999). Cross-reactivity potential underscores the need for exhaustive validation, particularly in closely related species. Addressing these concerns through optimization, primer design, and validation ensures SCAR markers maintain their precision and reliability in molecular studies.

The other main challenges, particularly in sensitivity and specificity (Vrba et al., 2010) when their development from Random Amplified Polymorphic DNA (RAPD) markers requires clear, high-intensity RAPD bands, yet faint bands can lead to reduced sensitivity (Paran & Michel, 1993). Sensitivity is also influenced by PCR conditions, with minor variations impacting detection, especially in low DNA concentration samples (Das et al., 2005). On specificity, SCAR markers can occasionally cross-react with non-target sequences in closely related species, causing false positives (Mahfouze & Mahfouze, 2019). Misinterpretation arises from challenges in distinguishing close sequences and potential binding to non-target sequences. Variability in PCR conditions across labs can further skew interpretations. For accurate SCAR marker interpretation, reference standards are crucial (Marieschi et al., 2012). Addressing these challenges requires standardized protocols, rigorous validation, continuous education, and collaborative research approaches.

1. **Future Trends and Innovations**

 Amidst the agricultural sector's struggle against climate change, resource constraints and growing food needs, the significance of SCAR markers gains prominence and contemporary biotechnological progress suggests a wide array of potential innovations and trends linked to these markers. In the forthcoming years, one of the primary focuses surrounding SCAR markers will likely be their integration with high-throughput sequencing platforms and computational biology tools. Such a confluence will enable rapid and comprehensive scanning of vast genetic resources, facilitating the discovery of novel genes and pathways associated with desirable agronomic traits (Merrick et al., 2011). Additionally, with the surge of artificial intelligence (AI) and machine learning in biological research, predictive models using SCAR marker data can be developed. These models could potentially forecast crop disease outbreaks or pest invasions based on genomic markers, enabling pre-emptive interventions and thus ensuring food security (Forghani et al., 2019). Another transformative trend revolves around the fusion of SCAR markers with digital and precision agriculture. As remote sensing, IoT (Internet of Things) and drone technologies become ubiquitous in farms, integrating SCAR marker data can lead to real-time monitoring of soil health, microbial diversity, and crop genetic diversity (Žibrat et al., 2021). This not only aids in optimizing resource use but also in personalizing farming interventions based on the genetic makeup of crops. Beyond these, the evolution of CRISPR-Cas systems and synthetic biology can potentially witness synergy with SCAR markers. Tailored genetic modifications, informed by SCAR marker insights, may lead to the development of crops with enhanced resilience, nutritional content, or yield (Sander & Joung, 2014). In the realm of sustainable agriculture, the role of biofertilizers and biopesticides cannot be overstated. Advanced SCAR marker techniques will likely streamline the validation of these bio-products, ensuring their efficacy and safety. This would contribute immensely to reducing the chemical footprint of agriculture and nurturing ecosystem health. As global agriculture evolves towards digital and molecular sophistication, the limitless innovations with SCAR markers symbolize both a beacon of hope for current challenges and the transformative power of science and technology in forging a sustainable farming future.

1. **Conclusion**

Within the complex nexus of agriculture and microbiology, the emergence of SCAR markers stands out as a beacon of innovation and precision. This review has shed light on the transformative potential of SCAR markers, underscoring their role in reshaping modern agricultural practices. From enabling meticulous microbial authentication to facilitating the development of robust biofertilizers and biopesticides, SCAR marker validation has proven to be an indispensable tool in the arsenal of agro-microbiologists. The synergy between these markers and the accelerating advancements in digital agriculture further accentuates their importance, positioning them as foundational in addressing the multifaceted challenges of global agriculture. Looking forward, it is evident that the fusion of traditional agricultural wisdom with modern SCAR marker technology will be pivotal in ushering in an era of sustainable, productive, and resilient farming systems. The future of agricultural microbiology is not only promising but also teeming with opportunities to explore, innovate, and transform.

**References**

Aggarwal, R., Gupta, S., Banerjee, S., & Singh, V. B. (2011). Development of a SCAR marker for detection of Bipolaris sorokiniana causing spot blotch of wheat. *Canadian journal of microbiology*, *57*(11), 934-942.

Ambreetha, S., & Balachandar, D. (2023). SCAR marker: A potential tool for authentication of agriculturally important microorganisms. Journal of Basic Microbiology, 63(1), 4-16.

Amiteye, S. (2021). Basic concepts and methodologies of DNA marker systems in plant molecular breeding. Heliyon, 7(10).

Anand, U., Vaishnav, A., Sharma, S. K., Sahu, J., Ahmad, S., Sunita, K., ... & Shukla, A. K. (2022). Current advances and research prospects for agricultural and industrial uses of microbial strains available in world collections. *Science of The Total Environment*, *842*, 156641.

Arumingtyas, E. L., Agustina, B. R. E., & Kusnadi, J. (2022). Developing RAPD-derived SCAR (Sequence Characterized Amplified Region) marker for flowering time in chili pepper. In *7th International Conference on Biological Science (ICBS 2021)* (pp. 78-81). Atlantis Press.

Babu, P., Baranwal, D. K., Harikrishna, Pal, D., Bharti, H., Joshi, P., ... & Singh, A. (2020). Application of genomics tools in wheat breeding to attain durable rust resistance. Frontiers in Plant Science, 11, 567147.

Bhagyawant, S. S. (2016). RAPD-SCAR markers: an interface tool for authentication of traits. Journal of Biosciences and Medicines, 4(01), 1.

Bintsis, T. (2018). Microbial pollution and food safety. *AIMS microbiology*, *4*(3), 377.

Castrillo, L. A., Vandenberg, J. D., & Wraight, S. P. (2003). Strain-specific detection of introduced Beauveria bassiana in agricultural fields by use of sequence-characterized amplified region markers. *Journal of Invertebrate Pathology*, *82*(2), 75-83.

Couillerot, O., Poirier, M. A., Prigent‐Combaret, C., Mavingui, P., Caballero‐Mellado, J., & Moënne‐Loccoz, Y. (2010). Assessment of SCAR markers to design real‐time PCR primers for rhizosphere quantification of Azospirillum brasilense phytostimulatory inoculants of maize. Journal of Applied Microbiology, 109(2), 528-538.

Dar, A. A., Mahajan, R., & Sharma, S. (2019). Molecular markers for characterization and conservation of plant genetic resources. *Indian Journal of Agricultural Sciences*, *89*(11), 1755-1763.

Das, M., Bhattacharya, S., & Pal, A. (2005). Generation and characterization of SCARs by cloning and sequencing of RAPD products: a strategy for species-specific marker development in bamboo. *Annals of botany*, *95*(5), 835-841.

Dixon, G. R., & Tilston, E. L. (Eds.). (2010). *Soil microbiology and sustainable crop production*. Springer Science & Business Media.

Forghani, R., Savadjiev, P., Chatterjee, A., Muthukrishnan, N., Reinhold, C., & Forghani, B. (2019). Radiomics and artificial intelligence for biomarker and prediction model development in oncology. *Computational and structural biotechnology journal*, *17*, 995.

Garcia, B. E., Graham, E., Jensen, K. S., Hanson, P., Mejía, L., & Maxwell, D. P. (2007). Co-dominant SCAR marker for detection of the begomovirus-resistance Ty-2 locus derived from Solanum habrochaites in tomato germplasm. *Tomato Genetic Cooperative Report*, *57*, 21-24.

Gotor‐Vila, A., Teixidó, N., Usall, J., Dashevskaya, S., & Torres, R. (2016). Development of a SCAR marker and a strain‐specific genomic marker for the detection of the biocontrol agent strain CPA‐8 Bacillus amyloliquefaciens (formerly B. subtilis). *Annals of Applied Biology*, *169*(2), 248-256.

Gupta, S. K., Charpe, A., Koul, S., Haque, Q. M. R., & Prabhu, K. V. (2006). Development and validation of SCAR markers co-segregating with an Agropyron elongatum derived leaf rust resistance gene Lr24 in wheat. *Euphytica*, *150*, 233-240.

Gupta, S. K., Charpe, A., Koul, S., Haque, Q. M. R., & Prabhu, K. V. (2006). Development and validation of SCAR markers co-segregating with an Agropyron elongatum derived leaf rust resistance gene Lr24 in wheat. *Euphytica*, *150*, 233-240.

Holmberg, A. I. J., Melin, P., Levenfors, J. P., & Sundh, I. (2009). Development and evaluation of SCAR markers for a Pseudomonas brassicacearum strain used in biological control of snow mould. *Biological Control*, *48*(2), 181-187.

Horejsi, T., Box, J. M., & Staub, J. E. (1999). Efficiency of randomly amplified polymorphic DNA to sequence characterized amplified region marker conversion and their comparative polymerase chain reaction sensitivity in cucumber. *Journal of the American Society for Horticultural Science*, *124*(2), 128-135.

Iruela, M., Rubio, J., Barro, F., Cubero, J. I., Millán, T., & Gil, J. (2006). Detection of two quantitative trait loci for resistance to ascochyta blight in an intra-specific cross of chickpea (Cicer arietinum L.): development of SCAR markers associated with resistance. *Theoretical and Applied Genetics*, *112*, 278-287.

Kasai, K., Morikawa, Y., Sorri, V. A., Valkonen, J. P. T., Gebhardt, C., & Watanabe, K. N. (2000). Development of SCAR markers to the PVY resistance gene Ry adg based on a common feature of plant disease resistance genes. *Genome*, *43*(1), 1-8.

Kesawat, M. S., & Das Kumar, B. (2009). Molecular markers: it’s application in crop improvement. Journal of Crop Science and Biotechnology, 12, 169-181.

Kiran, U., Khan, S., Mirza, K. J., Ram, M., & Abdin, M. Z. (2010). SCAR markers: a potential tool for authentication of herbal drugs. Fitoterapia, 81(8), 969-976.

Liu, X., Cheng, J., Mei, Z., Wei, C., Khan, M. A., Peng, J., & Fu, J. (2020). SCAR marker for identification and discrimination of specific medicinal Lycium chinense Miller from Lycium species from ramp-PCR RAPD fragments. 3 Biotech, 10, 1-7.

Mahfouze, S. A., & Mahfouze, H. A. (2019). A Comparison between CAPS and SCAR Markers in the Detection of Resistance Genes in some Tomato Genotypes against Tomato Yellow Leaf Curl Virus and Whitefly. *Jordan Journal of Biological Sciences*, *12*(2).

Marieschi, M., Torelli, A., & Bruni, R. (2012). Quality control of saffron (Crocus sativus L.): development of SCAR markers for the detection of plant adulterants used as bulking agents. *Journal of Agricultural and Food Chemistry*, *60*(44), 10998-11004.

Merrick, B. A., London, R. E., Bushel, P. R., Grissom, S. F., & Paules, R. S. (2011). Platforms for biomarker analysis using high-throughput approaches in genomics, transcriptomics, proteomics, metabolomics, and bioinformatics. *IARC scientific publications*, (163), 121-142.

Nunes, C., Bajji, M., Stepien, V., Manso, T., Torres, R., Usall, J., & Jijakli, M. H. (2008). Development and application of a SCAR marker to monitor and quantify populations of the postharvest biocontrol agent Pantoea agglomerans CPA-2. Postharvest Biology and Technology, 47(3), 422-428.

Paran, I., & Michelmore, R. W. (1993). Development of reliable PCR-based markers linked to downy mildew resistance genes in lettuce. Theoretical and applied genetics, 85, 985-993.

Perez, G., Verdejo, V., Gondim-Porto, C., Orlando, J., & Caru, M. (2014). Designing a SCAR molecular marker for monitoring Trichoderma cf. harzianum in experimental communities. *Journal of Zhejiang University. Science. B*, *15*(11), 966.

Ratajczak, M., Kubicka, M. M., Kamińska, D., Sawicka, P., & Długaszewska, J. (2015). Microbiological quality of non-sterile pharmaceutical products. *Saudi Pharmaceutical Journal*, *23*(3), 303-307.

Ray, P., Lakshmanan, V., Labbé, J. L., & Craven, K. D. (2020). Microbe to microbiome: a paradigm shift in the application of microorganisms for sustainable agriculture. Frontiers in Microbiology, 11, 622926.

Reddy Priya, P., Selastin Antony, R., Gopalaswamy, G., & Balachandar, D. (2016). Development of sequence-characterized amplified region (SCAR) markers as a quality standard of inoculants based on Azospirillum. *Archives of microbiology*, *198*, 257-267.

Reddypriya, P., & Gopalaswamy, G. (2017). Comparison of rapid methods for the extraction of bacterial DNA using scar marker for commercial liquid biofertilizerAzospirillum lipoferum (Az204) from TNAU.

Reddypriya, P., Soumare, A., & Balachandar, D. (2019). Multiplex and quantitative PCR targeting SCAR markers for strain‐level detection and quantification of biofertilizers. Journal of basic microbiology, 59(1), 111-119.

Sander, J. D., & Joung, J. K. (2014). CRISPR-Cas systems for genome editing, regulation and targeting. *Nature biotechnology*, *32*(4), 347.

Shah, A. P., Travadi, T., Sharma, S., Pandit, R., Joshi, C., & Joshi, M. (2023). Comprehensive analysis using DNA metabarcoding, SCAR marker based PCR assay, and HPLC unveils the adulteration in Brahmi herbal products. Molecular Biology Reports, 1-14.

Sharma, P., Kumar, V., Raman, K. V., & Tiwari, K. (2014). A set of SCAR markers in cluster bean (Cyamopsis tetragonoloba L. Taub) genotypes. Advances in Bioscience and Biotechnology, 5(02), 131-141.

Singh, R. B., Srivastava, S., Rastogi, J., Gupta, G. N., Tiwari, N. N., Singh, B., & Singh, R. K. (2014). Molecular markers exploited in crop improvement practices. *Res Environ Life Sci*, *7*(4), 223-232.

Singh, V. K., Singh, R., Kumar, A., & Bhadouria, R. (2021). Current status of plant diseases and food security. In *Food security and plant disease management* (pp. 19-35). Woodhead Publishing..

Vrba, V., Blake, D. P., & Poplstein, M. (2010). Quantitative real-time PCR assays for detection and quantification of all seven Eimeria species that infect the chicken. *Veterinary parasitology*, *174*(3-4), 183-190.

Yang, L., Fu, S., Khan, M. A., Zeng, W., & Fu, J. (2013). Molecular cloning and development of RAPD-SCAR markers for Dimocarpus longan variety authentication. SpringerPlus, 2, 1-8.

Yang, Y., Hu, J., Chen, F., Ding, D., & Zhou, C. (2018). Development of a SCAR marker-based diagnostic method for the detection of the Citrus target spot pathogen Pseudofabraea citricarpa. BioMed Research International, 2018.

Žibrat, U., Gerič Stare, B., Knapič, M., Susič, N., Lapajne, J., & Širca, S. (2021). Detection of root-knot nematode Meloidogyne luci infestation of potato tubers using hyperspectral remote sensing and real-time PCR molecular methods. *Remote Sensing*, *13*(10), 1996.