**SNAKE ENVENOMATION & NOVEL THERAPEUTICS**

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| **Abstract**Snakes are poikilothermic vertebrates. Snake venom is considered to be one of the most highly developed and complicated of all toxins produced by plants and animals. Snake venom is a complex mixture containing peptides, polypeptides, enzymes, glycoproteins and other substances, capable of several toxicological effects. Snakebite envenomation remains a significant global health issue, particularly in regions where venomous snake populations are prevalent. In recent years, the field of nanomedicine has emerged as a promising avenue for overcoming these challenges. This chapter explores the potential of nanoparticles as a novel therapeutic strategy for neutralizing snake venom in humans. The concept revolves around utilizing engineered nanoparticles as delivery vehicles for venom-neutralizing agents. These nanoparticles can be functionalized with specific targeting ligands to ensure efficient localization at the site of venom injection. Furthermore, the nanoparticles can be designed to encapsulate or conjugate venom-neutralizing compounds, such as small molecules, peptides, or antibodies, enhancing their stability, bioavailability, and therapeutic efficacy. This chapter outlines the current state of research in nanoparticle-based snakebite treatment, including various nanoparticle formulations, venom-neutralizing agents, and preclinical studies. It also highlights the potential challenges and considerations associated with this approach, such as nanoparticle toxicity, immunogenicity, and regulatory approval. In conclusion, the application of nanoparticles for neutralizing snake venom in humans holds great promise for addressing the limitations of current snakebite treatments. **Keywords:** nanoparticles, snake envenomation, experimental models | **Authors****Dr. Harish R** Assistant ProfessorDepartment of BiochemistryHaveri Institute of Medical Sciences Haveri, India-581110harishreddy1349@gmail.com **Dr. Gururaj Biradar** Assistant ProfessorDepartment of Forensic Medicine & ToxicologyHaveri Institute of Medical Sciences Haveri, India-581110rockguru18@gmail.com **Mrs. Vidhya K S**Data EngineerME Bioinformatics Bangalore, Indiavidhyaks1990@gmail.com **Dr. Ashakiran S**Professor and HoDDepartment of BiochemistryHaveri Institute of Medical Sciences Haveri, India-581110ashes27@rediffmail.com **Dr. Kotresh Doddamane**Professor and HoDDepartment of General MedicineHaveri Institute of Medical Sciences Haveri, India-581110kotresh\_doc@yahoo.co.in  |

1. **INTRODUCTION**

**A. Definition and Composition of Snake Venom:**

Snakes are poikilothermic vertebrates. Snake venom is a complex mixture of bioactive proteins and peptides that is produced and secreted by specialized glands in venomous snakes. It is primarily used as a means of predation and defense against potential threats. The composition of snake venom varies significantly among different species of venomous snakes [1]. Typically, venom contains enzymes, neurotoxins, cardiotoxins, cytotoxins, and other molecules that target specific physiological systems in prey or predators. These venom components work in synergy to immobilize or incapacitate the snake's prey, making it easier for the snake to consume its meal [2]. While the exact composition varies, most venomous snake venoms contain a combination of these toxic compounds tailored to their ecological niche.

**B. Overview of Venomous Snake Species:**

There are numerous species of venomous snakes found across the globe, belonging to different families and genera. Some of the most well-known venomous snake families include Elapidae (e.g., cobras, kraits, coral snakes) and Viperidae (e.g., vipers, pit vipers, rattlesnakes). Each family and species within it has evolved unique venom compositions and delivery mechanisms, making their study a diverse and intriguing field of research. Common venomous snakes found in India are cobra (Naja naja), common krait (Bungarus caeruleus), Russell’s viper (Daboia russeli) and saw scaled viper (Echis carinatus) [3, 4].

**C. Geographic Distribution of Venomous Snakes:**

Venomous snakes can be found in a wide range of habitats, including deserts, rainforests, grasslands, and even some aquatic environments. Their distribution is influenced by factors such as climate, prey availability, and territorial boundaries. Different venomous snake species are native to specific regions around the world, with some species confined to certain continents or countries, while others may have broader distributions [1].

**D. Importance of Studying Snake Venom and Its Potential Applications:**

The study of snake venom and its components has profound implications for biomedical research and drug development. Venom toxins have evolved to target specific molecular pathways, making them valuable tools for understanding cellular processes and signaling cascades. As a result, researchers have increasingly focused on the potential applications of snake venom in various fields of medicine. Some key areas of interest include:

Development of Antivenoms: Snakebite envenomation is a significant global health issue, affecting millions of people annually. The important enzymes in snake venom includes proteolytic enzymes, thrombin like enzymes, arginine ester hydrolase, collagenase, hyaluronidase, phospholipase A2, phospholipase B, phospholipase C, lactate dehydrogenase, phosphomonoesterase, phosphodiesterase, acetylcholinesterase, RNase, DNase, 5’- nucleotidase. It affects mainly the cardiovascular, nervous, renal and respiratory systems. Understanding the composition and action of snake venom is crucial for developing effective antivenom treatments to save lives and mitigate the impact of snakebites [5].

Venom from various sources offers exciting potential for medical applications. Components in venom possess analgesic properties, showing promise for novel pain-relieving drugs in chronic conditions. In cancer research, venom toxins display the ability to target cancer cells and hinder tumor growth, opening new avenues for drug development. Venom neurotoxins offer insights into neural signaling pathways, aiding neuroscience research and potential therapies for neurological disorders. Additionally, certain venom peptides have vasodilatory and antihypertensive effects, offering opportunities for developing cardiovascular drugs. Ongoing research on snake venom components continues to hold promise for novel drugs and therapies in various medical fields [5, 6].

**E. Detection of Snake Venom:**

The identification of snake venom holds significant forensic value, serving to establish the precise cause of death and prevent erroneous assertions. Various techniques have been devised for snake venom detection, including enzyme-linked immunosorbent assay (ELISA) and optical immune assay (OIA) [7]. In tropical countries the setting up of regional forensic science laboratories with snake venom detection facilities and capacity building is essential for addressing the mortality and morbidity due to snake envenomation.

**II. VENOMOUS SNAKE SPECIES AND THEIR VENOMS**

**A. Major Families of Venomous Snakes:**

Venomous snakes belong to different families, each with its distinct characteristics and geographical distribution [8]. Some of the major families of venomous snakes include:

* Elapidae: This family includes some of the most venomous snakes in the world. Examples include cobras, kraits, mambas, and coral snakes. Elapids typically have fixed front fangs and produce potent neurotoxic venoms.
* Viperidae: Vipers are known for their retractable hollow fangs, which allow for effective venom delivery. This family includes vipers, pit vipers (e.g., rattlesnakes, copperheads, and cottonmouths), and adders. Viper venoms often contain a combination of cytotoxic and hemotoxic components.
* Atractaspididae: This family includes burrowing asps, stiletto snakes, and mole vipers. They possess small fangs and produce cytotoxic venoms.
* Colubridae: Although the majority of colubrid snakes are non-venomous, some species within this family are mildly venomous. Examples include the boomslang and the twig snakes.

**B. Characteristics of Venomous Snake Venoms:**

Venomous snake venoms are highly diverse and comprise complex mixtures of bioactive proteins, peptides, enzymes, and other molecules. These venoms exhibit varying characteristics depending on the species. Common features include enzymatic components like proteases, phospholipases, hyaluronidases, and L-amino acid oxidases, which play roles in tissue breakdown, blood clotting interference, and cellular effects. Neurotoxic components in Elapidae family snake venoms target the nervous system, leading to paralysis or neuromuscular dysfunction. Viper venoms often contain hemotoxic and cytotoxic components causing damage to blood cells, blood vessels, cell death, and tissue necrosis. Additionally, some snake venoms have cardiotoxic effects impacting the cardiovascular system. Venom potency can differ significantly among species, with some causing severe envenomation and rapid effects while others are milder and cause less immediate harm [5, 6].

**C. Variation in Venom Composition among Species:**

The composition of venom can vary widely among different species of venomous snakes. This variation is due to the adaptation of venom to the ecological niche and specific prey of each snake species. Several factors influence the composition of venom in snakes. Prey preference plays a crucial role, with snakes that target small vertebrates possessing venoms rich in neurotoxins for swift immobilization, while those consuming larger animals may have cytotoxic and hemotoxic components for efficient prey digestion. Additionally, venom composition varies based on geographic distribution and the type of prey available in the region. Phylogenetic differences among snake families contribute to unique venom compositions. Even within the same species, inter-species variation can occur. Understanding these variations is vital for developing targeted antivenoms and exploring potential biomedical applications of venom components [8, 9].

**III. COMPONENTS OF SNAKE VENOM**

Snake venoms are complex mixtures containing a wide array of bioactive molecules, each with specific effects on the prey or potential predators [5, 10]. These components play critical roles in immobilizing or incapacitating the snake's target. Below, we'll explore the major components found in snake venom:

**A. Enzymes:**

*i. Proteases:*

Proteases are enzymes that break down proteins by cleaving peptide bonds. In snake venom, metalloproteases and serine proteases are common. Metalloproteases are involved in disrupting the extracellular matrix and basement membranes, leading to tissue damage and facilitating the spread of venom in the victim's body. Serine proteases interfere with blood clotting mechanisms, leading to the disruption of hemostasis [11]. Metalloproteinases are prominent constituents in the venoms of Crotalid and Viperid snakes. Snake Venom Metalloproteinases (SVMPs) exhibit diverse activities which encompass hemorrhagic, fibrinolytic, prothrombin activation, blood coagulation factor X activation, apoptotic, platelet aggregation inhibition, pro-inflammatory, and blood serine proteinase inhibitor inactivation. The SVMPs exhibit multiple functions beyond their renowned hemorrhagic activity [12].

*ii. Phospholipases:*

Phospholipases are enzymes that hydrolyze phospholipids present in cell membranes. Phospholipases A(2) (PLA(2)s) are abundant in snake venoms, serving both toxic and digestive functions. Snake venom PLA(2)s display a remarkable range of effects, including neurotoxic, myotoxic, hemolytic, edematogenic, hyperalgesic, pro-inflammatory, hypotensive, platelet-aggregation inhibitory, anticoagulant, cytotoxic, and bactericidal activities. [13]. Snake venoms from Colubridae, Elapidae, and Viperidae families are abundant in phospholipase A2s (PLA2s), also known as phosphatidylcholine 2-acylhydrolases. These enzymes primarily target phospholipids with unsaturated fatty acid tails at the sn-position, resulting in the formation of lysophospholipids and unsaturated fatty acids. The hydrolysis products alter cell membrane properties and trigger downstream signal transduction pathways, leading to widespread cellular pathology [14].

*iii. Hyaluronidases:*

Hyaluronidases are enzymes that degrade hyaluronic acid, an important component of the extracellular matrix. By breaking down this matrix, hyaluronidases facilitate the diffusion and spread of venom through tissues, increasing its local and systemic effects. Snake venom hyaluronidases (SVHYA) play a significant role in tissue destruction during envenomations and are known as spreading factors due to their ability to enhance venom toxin delivery. Interestingly, SVHYA are categorized in Enzyme Class 3.2.1.35, alongside mammalian hyaluronidases (HYAL). Both HYAL and SVHYA from Class 3.2.1.35 act on hyaluronic acid (HA), producing low molecular weight HA fragments (LMW-HA). LMW-HA generated by HYAL acts as a damage-associated molecular pattern, recognized by Toll-like receptors 2 and 4, triggering cell signaling cascades that lead to innate and adaptive immune responses. These responses involve the generation of lipid mediators, production of interleukins, upregulation of chemokines, activation of dendritic cells, and proliferation of T cells [15].

*iv. L-amino Acid Oxidases:*

L-amino acid oxidases (LAAO) catalyze the oxidative deamination of L-amino acids, producing hydrogen peroxide and ammonia. These enzymes are involved in various biological activities, such as promoting inflammation and apoptosis in target cells. Snake venoms contain significant concentrations of L-amino acid oxidases (LAAOs), which vary according to each snake species, potentially contributing to venom toxicity. LAAOs display catalytic specificity for long chain hydrophobic and aromatic amino acids and exhibit activity across a wide pH and temperature range. Their structures, molecular masses, and isoelectric points show considerable diversity. LAAOs have the ability to modulate platelet function, leading to local effects on plasma clotting disorders and other related effects. Additionally, LAAOs can induce apoptosis in various cell lines and demonstrate antimicrobial and antiparasitic activity [16].

**B. Neurotoxins:**

Snake venom toxins have traditionally been categorized into two types of neuromuscular blockade: pre-synaptic and post-synaptic. Pre-synaptically active neurotoxins, mainly neurotoxic phospholipase A2 toxins (PLA2s), bind to motor nerve terminals, causing depletion of synaptic ACh vesicles, impaired ACh release, and later, motor nerve terminal degeneration. This leads to a three-phase neuromuscular block, involving immediate ACh release depression, followed by enhanced release, and eventually complete inhibition of NMJ transmission. Post-synaptically active neurotoxins, known as alpha-neurotoxins, bind to post-synaptic muscle nAChRs and belong to the group of "three-finger toxins" (3FTXs) characterized by a shared toxin structure resembling three outstretched fingers. These alpha-neurotoxins are classified into three main groups: long-chain, short-chain, and non-conventional alpha-neurotoxins. They produce a reversible, non-depolarizing post-synaptic block by competitively inhibiting ACh binding to muscle nAChRs, leading to a curare-mimetic neurotoxic effect. However, recent insights into neuromuscular transmission and descriptions of different neurotoxicity patterns suggest that this view may be oversimplistic and requires reevaluation [17].

**C. Cytotoxins:**

Cytotoxins (CTs), also known as cardiotoxins, are toxins present in cobra venom that exhibit a three-finger (TF) fold. These CTs are approximately 60-residue-long peptides, containing up to 4 disulfide bonds. The β-strands originate from the hydrophobic core formed by the disulfides, taking the shape of the three loops, which gives the fold its name. In contrast to neurotoxins (NTs), another group of TF proteins from snake venom that exert their effects through specific interactions with protein receptors, CTs do not have a specific protein target identified. Unlike NTs, CTs are amphiphilic and possess cytotoxic properties against various cell types, including cancer cells [18].

**D. Other Bioactive Molecules:**

Snake venoms contain a diverse array of bioactive molecules with specific effects. Some examples include nucleotidases, enzymes that degrade nucleotides and release purine and pyrimidine derivatives with various biological activities. Metalloproteinases contribute to tissue damage and inflammation by degrading extracellular matrix proteins. Additionally, the venom may contain biogenic amines like serotonin and histamine, causing vasodilation, increased vascular permeability, and contributing to inflammatory responses. Furthermore, natriuretic peptides in the venom have diuretic and vasodilatory effects on the cardiovascular system [19].

**IV. MECHANISM OF ACTION OF SNAKE VENOM**

**A. Pathophysiology of Envenomation:**

The pathophysiology of snake envenomation involves the complex interactions of venom components with various physiological systems in the victim's body. Different venom components target specific molecular pathways, leading to a wide range of local and systemic effects [20]. Common pathophysiological mechanisms include:

**Local Tissue Damage:** Proteases, phospholipases, and cytotoxins in snake venom contribute to local tissue damage and inflammation at the site of the bite. These components degrade extracellular matrix proteins and cell membranes, leading to swelling, pain, and tissue necrosis [21]. Venom metalloproteinases exhibit diverse domain structures, including metalloproteinase domains, disintegrin-like and high cysteine domains, and lectin-like subunits. These zinc-dependent enzymes share similar zinc binding environments. Some directly induce hemorrhage by selectively cleaving basement membrane bonds, disrupting endothelial cell interactions and causing cellular alterations linked to hemodynamic factors. This results in endothelial gaps and extravasation. Beyond hemorrhage, these metalloproteinases cause skeletal muscle damage and myonecrosis due to ischemia from bleeding and reduced perfusion. Microvessel disruption hampers skeletal muscle regeneration, leading to fibrosis and persistent tissue loss post-snakebites. Furthermore, they degrade extracellular matrix components, contribute to local inflammation by inducing edema, activating matrix metalloproteinases, and releasing TNF-alpha. Due to their pivotal role in tissue damage, they're valuable targets for natural and synthetic inhibitors, which could complement antivenom treatments to neutralize their effects [22].

**Hemostatic Disturbances:** Snake venoms often contain enzymes that interfere with blood clotting mechanisms. Proteases can activate or inhibit blood clotting factors, leading to coagulopathies, which can result in both bleeding and thrombosis. Venomous snake procoagulant toxins categorically induce consumption coagulopathy, potentially leading to spontaneous or uncontrolled bleeding as clotting cascade factors are depleted. The impact on clotting factors varies among different snake venoms. These toxins, like metalloproteinases, play a pivotal role in activating prothrombin, factor V, factor X, or thrombin-like enzymes (fibrinogenases), making them significant procoagulant agents [23]. Thrombotic microangiopathy, often accompanying venom-induced consumption coagulopathy, manifests as thrombocytopenia, microangiopathic hemolytic anemia, and acute kidney injury [24]. The nature and extent of consumption coagulopathy vary based on the specific procoagulant toxin. These toxins typically activate clotting factors, resulting in diminished fibrinogen levels post-envenoming. Thrombin-like enzymes or fibrinogenases typically cleave the α-chain or β-chain of fibrinogen, yielding fibrinopeptide A or B. This leads to fibrinogen consumption without fibrin formation [25].

**Neurotoxic Effects:** Neurotoxins, particularly α-neurotoxins in Elapidae venoms, target acetylcholine receptors at neuromuscular junctions resulting in the blockade of neuromuscular transmission and paralysis of muscles, including those involved in respiration. Snake venoms often contain neurotoxins inducing a descending flaccid neuromuscular paralysis, which can involve dangerous bulbar and respiratory muscle blockade. Two primary neurotoxin types are α-neurotoxins and β-neurotoxins.

α-Neurotoxins, part of the three-finger toxin family, act postsynaptically at neuromuscular junctions, binding to cholinergic receptors and causing flaccid paralysis by inhibiting acetylcholine binding [26]. Conversely, β-neurotoxins are usually PLA2s acting presynaptically. For instance, β-bungarotoxin from kraits binds voltage-gated potassium channels, leading to enzymatic phospholipid hydrolysis, neurotoxicity, vesicle fusion, and calcium influx, ultimately destroying nerve terminals and causing prolonged paralysis. Some neurotoxic PLA2s also act intracellularly, exacerbating degenerative effects [27].

Other venom neurotoxins include dendrotoxins and fasciculins in African mamba venoms, blocking potassium channels and inhibiting acetylcholinesterase, respectively. Combined actions lead to excitatory effects and muscle contractions. Additionally, certain cysteine-rich secretory proteins induce smooth muscle paralysis [28].

**Cardiovascular Effects:** Cardiotoxins and certain venom enzymes can affect the cardiovascular system, leading to changes in heart rate, blood pressure, and cardiac function. These effects can range from mild disturbances to severe cardiac complications. Venom proteins and peptides can manifest diverse actions, with potential cardiotoxic or cardioprotective effects. The key categories of these compounds encompass cobra cardiotoxins, phospholipases A2, natriuretic peptides, and bradykinin-potentiating peptides. Another group includes proteins that stimulate angiogenesis, like vascular endothelial growth factors which possess hypotensive and cardioprotective properties. These venomic elements demonstrating cardiotropic and vasoactive influences hold promise as candidates for novel drug design aimed at preventing or mitigating the onset of pathological processes in cardiovascular ailments, a leading global cause of mortality [29].

**Systemic Toxicity:** Some venom components, such as metalloproteinases and other cytotoxic molecules, can cause systemic effects by entering the bloodstream and affecting multiple organs and tissues. Systemic manifestations include generalized, hematologic, and neuromuscular symptoms. General indications encompass weakness, fatigue, anxiety, tachycardia, weak or bradycardic pulse, tachypnea, diaphoresis, nausea, vomiting, diarrhea, abdominal pain, hypothermia, fever, lymphadenopathy, lymphangitis (within 1–2 days), metallic taste, headache, thirst, pulmonary edema, heart failure, hypotension, collapse, shock, cerebral anoxia (resulting in sleepiness, slurred speech, disorientation, delirium, unconsciousness), and anaphylactic reactions due to venom proteins (causing facial, tongue, and epiglottic angioedema and respiratory tract obstruction). Rapid-onset hypotension within 2 hours signifies significant intoxication. Hypotension may manifest briefly (resolving spontaneously within 2 hours), persistently, or progressively, leading to fatality. Renal impairment presents as proteinuria, hemoglobinuria, myoglobinuria, azotemia, and anuria. Convulsions can arise from cerebral anoxia due to hypotension and anemia rather than direct venom neurotoxicity. Electrocardiograms (ECGs) may exhibit nonspecific ST segment and T wave changes, along with atrial fibrillation episodes. Hematological changes span coagulopathy and hemorrhagic syndrome ranging from localized bleeding to substantial blood loss, including bleeding from gums, nose, hematemesis, melena, hematuria, petechiae, and ecchymoses around the snakebite site. Bleeding may arise from kidneys, lungs, peritoneum, rectum, vagina, endometrium, and pathological locations like peptic ulcers. Slight icterus might be discernible. Laboratory assessments display extended prothrombin and activated partial thromboplastin times (APTT), decreased fibrinogen, plasminogen, factor XIII, factor V, antithrombin III, protein C levels, thrombocytopenia, elevated D-dimer levels, morphological erythrocyte changes, anemia due to blood extravasation, and neutrophilic leukocytosis. Neurological signs are distinct in Elapidae family snakebites, emerging within 8 hours of a latent period [30, 31].

**B. Interactions with Cellular Components:**

Snake venom components engage with various cellular elements in the victim's body, yielding diverse effects. Phospholipases in venom target cell membranes, causing membrane disruption and lysis. Neurotoxins bind to specific nerve cell receptors, altering signal transmission and causing paralysis. Hemotoxic elements, including proteases, influence platelets and blood cells, inducing coagulation disruptions and hemolysis. Venom enzymes, like metalloproteinases, degrade extracellular matrix constituents, contributing to tissue damage and venom propagation [32].

**V. NEUTRALIZATION OF SNAKE VENOM**

**A. Antivenom Development:**

**Production and Purification of Antivenoms:**

Snake antivenoms, crucial for treating snakebite envenomation, have remained largely consistent in their production approach since their inception over a century ago, albeit with the integration of technological advancements in the manufacturing process. These medications are required to adhere to the contemporary standards of identity, purity, safety, and efficacy outlined by the current Good Manufacturing Practices (GMPs) governing modern biopharmaceutical drugs. The industrial production of snake antivenoms involves multiple key phases: 1) creating reference venom pools, 2) generating hyperimmune plasma, 3) purifying antivenom immunoglobulins, 4) formulating the antivenom, 5) stabilizing the formulation, and 6) conducting quality control assessments for both in-process and final products. This review provides an overview of the prevalent technology employed in the industrial manufacturing of snake antivenoms [33].

Antivenoms are developed by immunizing large animals, such as horses or sheep, with small, controlled amounts of snake venom. The immune response in these animals results in the production of polyclonal antibodies against venom toxins. These antibodies are then harvested from the animal's blood, purified, and formulated into antivenom products. The production and purification process are complex and require careful consideration of factors such as antigen selection, animal welfare, and immunogenicity. Furthermore, antivenoms must be thoroughly tested for safety, potency, and efficacy to ensure their effectiveness in neutralizing venom toxins in snakebite victims [34]. Antivenoms are produced through the plasma fractionation of immunized animals, resulting in products containing complete IgG or immunoglobulin fragments such as F(ab')2 or Fab. Antivenom formulations exhibit distinct physicochemical attributes across different laboratories, encompassing variations in total protein content, levels of protein aggregates and non-IgG plasma proteins, and the inclusion of Fc fragments in whole IgGs [35].

To mitigate the drawbacks inherent in traditional antivenom manufacturing, a number of researchers have explored alternative approaches using existing production methods. They have investigated the use of recombinant or synthetic toxins, peptides, and DNA vaccination strategies to generate antibodies with therapeutic relevance through immunization techniques. This advancement not only eliminates the current necessity for venom in antivenom production, thereby obviating the need for maintaining collections of venomous animals, but also significantly reduces the labor-intensive and potentially hazardous aspects associated with animal handling for personnel [36]. Nonetheless, venom remains essential for quality control and research validation of antivenom, ensuring the efficacy of newly manufactured antivenom products.

**Challenges and Advancements in Antivenom Development:**

Despite their life-saving potential, antivenom development faces several challenges. These challenges include the complexity of venom composition, regional variations in venom components among different snake populations, and the limited availability of antivenoms for some species. Additionally, the high cost of production, the need for proper storage, and issues related to adverse reactions to antivenoms pose significant obstacles. Researchers are exploring novel approaches, such as the use of recombinant antibodies or synthetic peptides, to overcome these challenges and develop safer and more effective antivenoms [35].

The initial antibody formats explored for creating recombinant antivenoms involved monoclonal IgGs and single-chain variable fragments. Extensive in vivo studies have highlighted the effectiveness of monoclonal IgGs in counteracting the myotoxic, hemorrhagic, and proteolytic effects of snake toxins. These specifically engineered monoclonal IgGs have been designed for medically significant snake species such as Naja spp., Crotalus spp., Echis spp., Laticauda spp., and Bothrops spp. Additionally, monoclonal antibodies that target a phospholipase A2, a metalloproteinase, and a thrombin-like component have been successfully generated through hybridoma technology. The combined use of these antibodies proved capable of preventing the in vivo lethality induced by Bothrops atrox venom [37].

Chinese Hamster Ovary (CHO) cell-based expression systems are widely used for large-scale production of monoclonal recombinant antibodies, in part due to their capacity to generate glycosylation patterns similar to those in humans. The conventional method for CHO cell expression is the fed-batch process, where nutrients are supplied to the CHO cells for the entire manufacturing cycle, followed by batch harvest. The antibodies obtained from this batch are subsequently purified through one or more chromatographic purification steps. Emerging cost-effective production methods include hybrid and continuous perfusion processes. In the hybrid approach, cultivation is carried out in a fed-batch reactor, followed by continuous or semi-continuous purification of the produced antibodies. In the continuous perfusion process, cultivated cells are retained while the antibody-containing growth medium is continuously replaced with fresh medium in a perfusion reactor. The used medium undergoes continuous or semi-continuous purification to isolate the antibodies. Notably, several companies in the industry are utilizing continuous chromatographic techniques, such as Simulated Moving Bed Chromatography (SMBC), which offers efficient use of chromatographic media and yields higher output of purified product [38].

**B. Nanotechnology for neutralizing snake venom**

Nanoparticles serve as effective carriers for antivenom, enhancing stability and bioavailability. They optimize sustained release, improving antivenom's binding to toxins, increasing potency, and reducing dosage. Portable nanoparticle-based formulations enable rapid on-site treatment in remote areas. They also stabilize antivenom during storage and transportation, extending shelf life and ensuring efficacy [39].

Earlier research has demonstrated the remarkable stability of metal nanoparticles and their suitability for snake venom inhibition [40, 41]. Silver nanoparticles (SNPs) are synthesized by chemical reduction method and characterized using UV-Visible spectroscopy, Dynamic Light Scattering (DLS) and Transmission electron microscope (TEM) [42]. DLS study showed the formation of complex between SNPs and crude viper venom with decrease in hydrodynamic size of complex compared to the size of native viper venom. Venom components might have adsorbed on the surface of SNPs and making Viper venom proteins more compact in nature results in modification of its activity [42]. Scientists from the University of Calcutta's toxicology laboratory attached gold nanoparticles (GNP) to HMBA (2-hydroxy-4-methoxy-benzoic acid), a compound derived from the root of the Anantamul (*Hemidesmus indicus*) herb, known for its anti-venom properties. Through animal trials, it was demonstrated that the GNP-HMBA pairing effectively counteracted various forms of toxicity (including kidney, muscle, and liver toxicity) in mice exposed to Russell's viper venom, which is among the most potent snake venoms [40].

Synthetic polymer nanoparticles have also been used as broad spectrum antivenom. This versatile antivenom consists of polymer nanoparticles (NPs) specially designed to capture the primary protein toxins found in elapid snake venoms. These durable, economically efficient NPs can be subcutaneously administered directly at the bite site immediately after envenomation, effectively curbing or minimizing local tissue damage and curtailing the systemic dissemination of toxins following envenomation [43].

The potential immunoadjuvant properties of chitosan nanoparticles (CNPs) loaded with venoms from B. jararaca and B. erythromelas in the production of sera targeting these venoms has been explored. Stable CNPs were created through ionic gelation, and mice were subcutaneously immunized over six weeks using 100 µL of each snake venom at concentrations of 5.0% or 10.0% (w/w), either encapsulated in CNPs or combined with aluminum hydroxide (AH). Assessment of protein interactions with CNPs demonstrated their capacity to induce antibody levels comparable to those induced by AH, even with lower antigen doses. Furthermore, CNPs exhibited reduced inflammatory effects due to their controlled protein release. This suggests that CNPs offer a promising avenue for delivering peptides and proteins from snake venoms and hold potential for the development of novel vaccines [44].

The increasing adoption of environmentally friendly approaches has spurred the creation of silver nanoparticles (AgNPs) through various sources, including bacteria, fungi, algae, and plants, enabling extensive production while minimizing contamination referred to as ‘Green Synthesis’. Green synthesis represents an eco-conscious and biologically compatible method, typically involving the utilization of a capping agent or stabilizer (to regulate size and prevent clustering), plant extracts, yeast, or bacteria to achieve these objectives [45]. Rhizomes of the Indian male fern, *Dryopteris cochleata*, were employed for the green synthesis of silver nanoparticles with the goal of enhancing the bioactivity of the plant extract and assessing the inhibitory effect on *N. naja* snake venom. The findings from the neutralization experiments demonstrated that the silver nanoparticles synthesized from *D. cochleata* effectively reduced tissue damage, leading to a substantial inhibition of phospholipase A2 and the venom of *N. naja* snakes [46].

Recent approaches in snake venom neutralization with a focus on nanoparticles hold immense promise for revolutionizing snakebite management. These innovations aim to enhance the efficacy, accessibility, and safety of antivenom therapy, ultimately improving the prognosis for snakebite victims. While challenges remain, such as regulatory approval and scalability, the integration of nanotechnology into snakebite treatment represents a significant stride towards more effective and accessible care for those affected by venomous snakebites.

**C. Small Molecule Inhibitors:**

Small molecules are being investigated as inhibitors of venom toxins. These compounds can interfere with the toxic mechanisms of snake venom and provide a novel approach to managing snake envenomation. Compound 5d, a small molecule derived from apigenin, has demonstrated its efficacy against SVMPs both in computational simulations and live experiments. Apigenin based small molecules with a flexible substitution at second position of the chroman moiety are prepared by multi-component Ugi reaction using various aromatic amines, t-butyl-isocyanide and halo acetic acids. A library of various apigenin analogues was synthesized, and among these, compound 5d exhibited a significant dose-dependent reduction in Echis carinatus (EC) venom-induced local hemorrhage, tissue necrosis, and myotoxicity [47].

**D. Gene Therapy:**

Gene-based therapies, including DNA vaccines, are being explored to prompt the production of specific antivenom antibodies within the victim's body, offering rapid, on-demand protection against venom toxins. This approach leverages the cost-effective and swift insights gained from random gene sequencing, known as expressed sequence tags (ESTs), into the intricate biochemical processes underlying various biological phenomena. This is exemplified in identifying clinically significant toxins in snake venoms, aiding immunotherapeutic treatments for snakebites. Snake venoms typically contain over a hundred proteins with significant inter- and intraspecific diversity in toxicity, posing challenges for purification and characterization. A systematic method for selecting immunoprotective sequences based on molecular sequence data alone can lead to the development of innovative, highly avid polyspecific antivenoms, enhancing dosages' effectiveness and minimizing toxicity in snakebite victims. DNA immunization offers a rational approach to toxin-specific immunotherapies, as previously demonstrated, inducing antibody levels and protective responses suitable for antivenom production and cross-reactivity with venoms from diverse viper species and genera [48].

**VI. FUTURE PERSPECTIVES**

* *Personalized Medicine:* Advances in genomics and proteomics may enable personalized snake venom therapeutics tailored to an individual's venom sensitivity, maximizing efficacy.
* *Multi-Omics Approaches:* Integrating genomics, transcriptomics, proteomics, and metabolomics can provide a comprehensive understanding of venom composition and action, guiding therapeutic design.
* *Nanotechnology:* Continued exploration of nanoparticles as carriers for antivenom components may lead to more efficient and targeted delivery systems, reducing dosages and side effects.
* *Global Collaboration:* Collaborative efforts involving researchers, healthcare providers, governments, and pharmaceutical companies can expedite the development, affordability, and global distribution of snake venom therapeutics.
* *Regulatory Streamlining:* Simplifying and streamlining regulatory processes for innovative therapeutics can accelerate their development and approval.
* *Community Engagement:* Engaging local communities in snakebite prevention, awareness, and treatment, including novel therapeutics, can improve outcomes and save lives.
* *Education and Training:* Investing in education and training programs for healthcare professionals in snakebite-prone regions is vital to enhance the timely and effective administration of snake venom therapeutics.

**VII. CONCLUSION**

In conclusion, the field of snake venom research has made significant strides in recent years, offering novel perspectives on the treatment and mitigation of snakebite envenomation. From the development of innovative antivenom delivery systems, such as nanoparticles, to the utilization of molecular sequencing data to design toxin-specific immunotherapies, these advancements hold great promise in improving the outcomes for snakebite victims. By leveraging green synthesis methods and exploring natural compounds like apigenin derivatives, researchers are expanding the toolkit for combating venomous snakebites. These approaches not only enhance antivenom efficacy but also contribute to reducing the burden of snakebite-related morbidity and mortality, particularly in regions where access to conventional antivenom may be limited. As the global scientific community continues to collaborate and innovate, the prospects for more effective and accessible snakebite treatments are brighter than ever, offering hope to those at risk of snake envenomation worldwide.

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