

# **DNA METHYLATION AND HISTONE MODIFICATION AS A DRIVING FORCE OF CANCER**

Mansa Pandey, Neha Srivastava, Richa Mishra, Somya Singh, Toshi Chandra, Deepika Delsa  
Dean\*

Department of Biotechnology, Isabella Thoburn College, Lucknow, Uttar Pradesh, India .

## **ABSTRACT**

Cancer is a major human health problem worldwide and is largely characterized by its abnormal increase in cell number and cell proliferation, decreased cell death or apoptosis, and its ability to invade surrounding tissues. A lot of factors are responsible for causing cancer one of which include epigenetic changes.

Epigenetic changes are concerned with genetic alterations to drive the cancer phenotype. Epigenetic changes result due to DNA methylation, histone modifications, chromatin remodelling, microRNAs, and other components of chromatin [3]. Histone modifications include histone acetylation, histone methylation, histone phosphorylation and histone ubiquitination. Epigenetic changes can also be induced by exposure to various environmental stimuli. These stimuli include aging that was shown with increased levels of DNA methylation; physical agents such as X – rays, UV light; infectious agents such as bacteria and viruses; similarly chronic inflammation triggered by various factors was also involved in inducing aberrant DNA methylation [1].

The sections in this chapter provide us with the controlled information about role of epigenetics in causing cancer and how it can be controlled with the help of epigenetic therapy .

**KEYWORDS:** Cancer, Epigenetics, Histones Modifications, DNA methylation, Acetylation, Deacetylation, tumor suppressor genes, oncogenes.

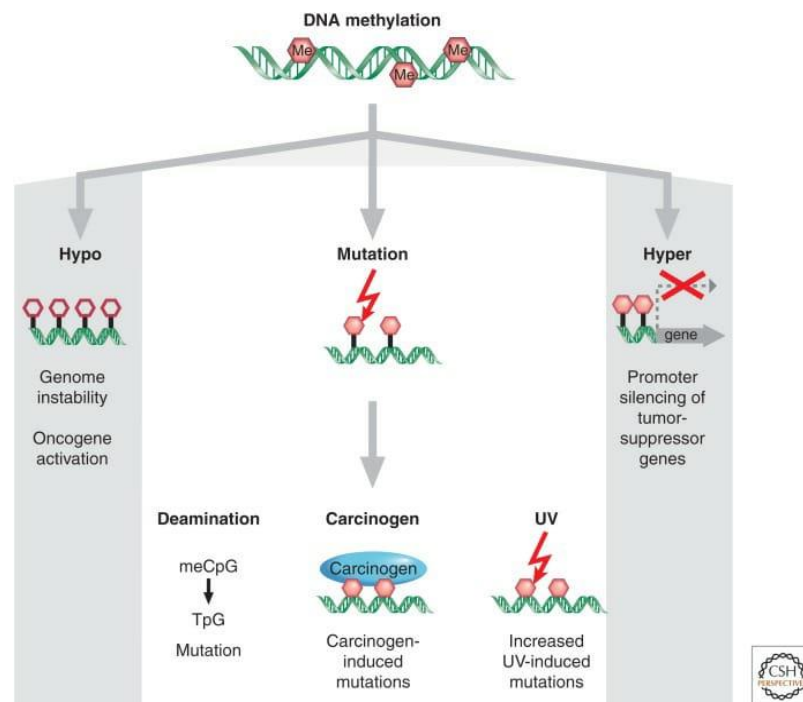
## **1. INTRODUCTION**

In 1940 C.H. Waddington defined the term “epigenetics” which means “the branch of biology which studies the causal interactions between genes and their products, which bring the phenotype into being” [4]. Accumulation of genetic mutations with epigenetic alterations together with environmental factors are the main root cause of cancer. Epigenetic alterations are reversible. These epigenetic mechanisms are regulated by four classes of epigenetic

regulators in a coordinated manner. Those which add the epigenetic marks are known as ‘writers’; which remove the epigenetic marks are known as ‘erasers’; which recognize specific epigenetic marks to mediate proximate effects are known as ‘readers’; and which modify chromatin structure are known as ‘remodelers’. Mammals contain about ~ 1000 epigenetic regulators forming the largest protein group.

## 2. CANCER EPIGENETICS

Cancer is caused by changes to the DNA within cells either by somatic mutations or epigenetic mutations. Mutation causing loss of function in tumor suppressor genes or gain of function in oncogenes and abnormal expression eventually leads the path to cancer. Whereas, the epigenetic pathway to cancer is determined by DNA methylation, histone modifications, nucleosome remodelling as well as miRNAs (Sharma et al., 2010). The figure below describes the various mechanisms involving DNA methylation that can lead to cancer [3]. During tumor initiation and progression, the epigenome goes through multiple alterations, including loss of DNA methylation (hypomethylation), frequent increases in promoter methylation of CpG islands, changes in nucleosome occupancy, and modification profiles.



## 3. DNA METHYLATION

Gene expression is controlled in eukaryotes, but methylation of DNA is a common epigenetic signalling tool that cells use to lock genes in the "off" position. DNA methylation is an important regulator of gene transcription, and its role in carcinogenesis has been a topic of

considerable interest in the last few years. Alterations in DNA methylation are common in a variety of tumors as well as in development. Of all epigenetic modifications, hypermethylation, which represses transcription of the promoter regions of tumor suppressor genes leading to gene silencing, has been most extensively studied. However, global hypomethylation has also been recognized as a cause of oncogenesis. New information concerning the mechanism of methylation and its control has led to the discovery of many regulatory proteins and enzymes. The contribution of dietary folate and methylene tetrahydrofolate reductase polymorphisms to methylation patterns in normal and cancer tissues is under intense investigation.

As methylation occurs early and can be detected in body fluids, it may be of potential use in early detection of tumors and for determining the prognosis. Because DNA methylation is reversible, drugs like 5'-azacytidine, decitabine, and histone deacetylase inhibitors are being used to treat a variety of tumors. Novel demethylating agents such as antisense DNA methyltransferase and small interference RNA are being developed, making the field of DNA methylation wider and more exciting.

### **3.1 Hypermethylation and Silencing in driving tumorigenesis**

In tumor suppressor genes hypermethylation of promoter region is commonly observed in cancers, a phenomenon that has been implicated with driving tumorigenesis (Baylin and Jones, 2011). Cell cycle and DNA repair genes, such as *RB*, *BRCA1/2*, and *PTEN*, have all been shown to be hypermethylated or mutated/deleted in cancer (Hatziaopostolou and Iliopoulos, 2011). Epigenetic silencing also induces cancers by abnormal activation of signalling pathway, further promoting the expression of several genes whose products are responsible for cell proliferation. As a result of survival and proliferation, these cells accumulate genetic mutations in other components of the several signalling pathway. Such as in normal cells secreted frizzled- related proteins (SFRPs) antagonize WNT signalling, silencing of these SFRPs accumulate in WNT signalling pathway [5].

### **3.2 DNA Methyltransferase Family and Establishment of DNA Methylation Profiles**

The first identified DNA methyltransferase DNMT1 is known as a maintenance DNMT. It is responsible for the exact copying of the DNA methylation pattern on the neo-synthesized strand during DNA replication. Therefore, it principally localizes to the DNA replication fork. Due to its importance in DNA replication, DNMT1 expression is tightly regulated during the cell cycle by several mechanisms and maximal expression occurs during S phase.

Since its role is to ensure the inheritance of the DNA methylation pattern through cell division, DNMT1 expression is maintained after development. From a transcriptional point of view, two transcript variants of DNMT1 mRNA were identified: a full-length form of 1,616 amino acids, and an oocyte-specific variant that lacks the N-terminal 118 amino acids of the full-length form (DNMT1o) but both are enzymatically active.

DNMT1 is capable of *de novo* methylation but its affinity for unmethylated DNA is far lower than for hemi-methylated DNA. As an illustration of the crucial role of DNMT1, the genetic loss of *DNMT1* gene in the mouse model is embryonic lethal. The *de novo* DNA methyltransferases DNMT3a and DNMT3b are responsible for the establishment of DNA methylation patterns during development. They are highly expressed during embryogenesis. Similarly, to DNMT1, DNMT3a and 3b expression is increased in S phase but they do not localize at the DNA replication fork. Immuno-fluorescence studies show that both *de novo* DNMTs localize to heterochromatin 6, and further experiments demonstrate that DNMT3a and DNMT3b are strongly associated to nucleosomes containing methylated DNA, and promote propagation of DNA methylation through stabilization of those enzymes.

### **3.3 DNA Methylation Alterations in Cancers**

Alteration of DNA methylation patterns is a hallmark of cancer. Numerous studies describe repression of tumor suppressor genes (TSG) involved in various cellular pathways (cell cycle, apoptosis or genome maintenance) during carcinogenesis by DNA hypermethylation of their promoters. Paradoxically, cancer cells exhibit a global genome hypomethylation that leads to genomic instability and re-expression of silenced genes.

Mechanisms underlying this paradox are still not clearly explained. Wild and Flanagan depict current knowledge on genome wide DNA hypomethylation associated with cancer. Briefly, two competing theories of “passive” vs. “active” demethylation processes could explain this phenomenon. The former implies a disruption of the link between histone modifications and DNA methylation establishment, an aberrant localization of DNMT1 to DNA damage sites or a metabolic imbalance favoring a decrease in the methyl group donor, *S*-adenosyl-methionine.

Conversely, the latter theory relies on a class of enzymes harboring a demethylase activity. The TET protein family (Ten Eleven Translocation proteins) is described to actively demethylate methyl-cytosines by their oxidization and elimination through different mechanisms in physiological conditions. Briefly, the TET enzyme family facilitates passive

DNA demethylation by oxidizing methyl-cytosines to 5-hydroxyl-methylcytosines (5 hmC) leading to a considerable reductions in UHRF1 binding (ubiquitin-like containing PHD and RING finger domains) and in DNMT1 methyltransferase activity at the replication fork.

A second mechanism involves the DNA repair pathway. Hydroxy-methyl cytosines are converted either by further oxidization or by deamination that leads to a nucleotide mismatch, which will be excised and replaced by a cytosine. Last, DNMT3a demonstrates methyltransferase activity in reducing conditions and conversely, dihydroxy methylation in oxidizing conditions that converts 5 hmC in cytosines.

Recent studies report that the induction of TET suppresses breast tumor growth, invasion and metastasis in mouse xenografts. Moreover, TET down-regulation in hepatocellular carcinoma correlates with a decreased level of 5 hmC and is associated with tumor size and poor overall survival. Taken together, these observations are controversial, with a pro-oncogenic effect of TET mediated-DNA demethylation.

#### **4. HISTONE MODIFICATION**

Along with DNA Methylation, histone modification after translation does not alter the nucleotide sequence of the DNA enhances it efficiency for transcriptional machinery. Epigenetic machinery is classically represented by DNA methylation and histone modification. Histone modification includes acetylation phosphorylation, ubiquitination, methylation, etc. Histone modification has role in allergic reaction that can be manifested at two level, first regulating these cells that cause allergic inflammation, like T cells and macrophage and the participants of airway. Second, the direct connection between allergic phenotype and histone modification drugs that could inhibit the histone modification enzymes. They usually act at the end terminal of histone with amino acids such as arginine or lysine as well as threonine, serine tyrosine etc. Histone methylation has effective role in gene expression. Methylation of histone could be either transcriptionally permeable or impermeable based on the position of the amino acid in tail or number of the group added that are modified.

##### **4.1 Histone Acetylation**

Acetylation of histone is modulated by two analogously acting enzymes histone deacetylases (HDACs) and histone acetyltransferases (HATs). The acetyl group transfer from acetyl CoA is mediated by HAT to the amino acid group of lysine residues targeted at the tail. Acetylation

removes histone positivity resulting in weaker interaction between DNA and histone. Weak interaction between DNA and histone lessens the compaction of chromatin and hence enhances its availability to the machinery of transcription. HDACs have a role in repressing gene expression by removing the acetyl group from the tail of histone where lysine residue is present. Five families of HATs are known GNAT family, MYST family, p300/CBP family, steroid receptors co-activators family, and cytoplasmic HATs. GNAT family comprises KAT2A and KAT2B enzymes that regulate cell cycle, DNA repair, and replication and are also important for centrosome function. The MYST family comprises KAT6A/MYST3/MOZ, KAT6B/MYST4/MORF, KAT7/MYST2/HBOI, KAT8/hMOF/MYST and KAT5/Tip60 that regulate DNA repair and transcription. The MYST family enzymes have unique characteristics of autoacetylation. p300/CBP family comprises of KAT3A and KAT3B enzymes and KAT4/TBP/TAF1 and TIF 3C90/KAT12 are contributors of general transcriptional factor related HAT family. KAT13A/SRC1, KAT13B/SRC/AIB1/ACTR, KAT13C/p600, and KAT13D/ CLOCK are members of the steroid receptor coordinators family. Cytoplasmic HATs have Kat1/HAT1 and HAT4/NAA 60 [29].

#### **4.2 Histone Methylation**

Histone methylation (HMTs) mediate histone methylation. These include lysine and arginine methyltransferase along with histone demethylases (HDMs) for demethylation. Histone methylation of lysine or Arginine influences the binding of different regulatory proteins to the chromatin indirectly. Three methyl groups can be transferred by HMTs from cofactor S-adenosyl-L-methionine SAM to the histone lysine or Arginine residues. The specificity of KMTs is higher than HATs because of the target-specific lysine residues. Methylation of different histone residues is mediated by different KMTs. H3K4 residue is methylated by KMT2A/MLL1, KMT2A/MLL2, KMT2F/hSET1A, KMT2G/hSET1B or KMT2H/ASH1. H3K9 is methylated by KMT1A/SUV39H1, KMT1B/SUV39H2, KMT1C/G9a or KMT1D/EuHMTase/GLP. methylation of H3K36 is catalyzed by KMT3B/NSD1, KMT3C/SMYD2 OR KMT3A/SETD2. Others like H3K27 is methylated by KMT6A/EZH2 and H3K79 by KMT4/DOT1L etc. based on mechanism of catalysis and sequence homology HDMs are of two types: 1) amine oxidase type lysine-specific demethylase (LSDs or KDM-1s) 2) Jumonji C (JMJC) domain-containing HDMs. The LSDs include KDM1A/LSD1/AOF2 and KDM1B/LSD2/AOF1 which remove the methyl groups and demethylated H3K4. The JMJC domain containing HDMs acts on mono, di, and trimethylated lysine residues and catalyzes their demethylation at histone [25,44,45,46,47].

### **4.3 Histone Phosphorylation**

Two antagonistic enzymes kinases and phosphatases remove it [19,21]. Phosphorylated histones are known for their three special functions: 1) chromatin compaction control, 2) DNA damage repair, 3) transcriptional activity repair. Histone phosphorylation creates a platform for the other histone modifications to interact. Histone H3 phosphorylation directly affects the two amine acid residues of the same histone (H3K9ac and H3K14ac) at acetylation levels [19,21,48].

### **4.4 Histone Ubiquitination**

Histone ubiquitination has a role in every aspect of cellular function in cell signaling pathways, especially in eukaryotes. Ubiquitination is regulated by a protein ubiquitin of 8.5 kD conjugated with substrate protein by the ubiquitin proteasome system, hence regulating the stability and turnover of target protein. Ubiquitination and DE ubiquitination are catalyzed by histone ubiquitin ligases and ubiquitin-specific peptidases (ubiquitinating enzymes) (DUBs) respectively [51,52,53]. The number of ubiquitin-associated results in distinct functions. Mono ubiquitination regulates protein translocation, transcriptional regulation, and DNA damage signaling. Histone 2A Mono ubiquitination deals with gene silencing. Histone 2B Mono ubiquitination (H2Bub) is associated with transcription activation. Activation or degradation of a certain protein is marked by polyubiquitination in cell signaling pathways. Histone ubiquitination also provides a platform for crosstalk between other histone modifications, for example: histone H3 Mono ubiquitination induces acetylation of the same histone [55].

## **5. EPIGENETIC THERAPY**

A number of ways have been developed in treating cancer, some which include chemotherapy, radiotherapy, surgery for the removal of local tumors, cytotoxic treatments, etc. But due to some major drawback of these treatments like damaging or killing normal cells or limitations in their effectiveness, other treatments like immunotherapy and epigenetic therapy are also considered.

The bottom of epigenetic treatments rely on treating cancer cells by reversing its abnormal modifications with the help of drugs known as “epi-drugs”. Generally, the enzymes involved in epigenetic modifications are the main targets of epigenetic therapy. These enzymes are Histone deacetylase (HDACs), DNA methyltransferases (DNMTs) and histone demethylases (HDMs). Therefore, the epi-drugs being considered are inhibitors of these enzymes such as HDAC inhibitors and DNMT inhibitors.

## **5.1 Histone Deacetylase (HDAC) Inhibitors**

Acetylation is the process involved in histone modification and its main function is to add acetyl groups to histones. An antagonistic effect to acetylation is caused by histone deacetylation by an enzyme HDAC which eventually leads to gene silencing of tumor suppressor genes and DNA repair genes. Therefore, HDAC inhibitors decrease Histone deacetylase activity and indirectly increase the activity of tumor suppressors genes. Example of HDAC inhibitors for clinical use includes vorinostat mefniyh{ (Zolinza®), romidepsin (Istodax®).

## **5.2 DNA Methyltransferase (DNMT) Inhibitors**

One of the major occurrence of cancer are often result of heavy methylation and silencing of tumor suppressor genes. During DNA methylation enzymes called DNA methyltransferases add methyl groups to bases in DNA. When these epi drugs comes in contact with DNA they inhibit any DNMTs that come along and ultimately destroyed them preventing them from further methylation. Treatment of acute myeloid leukemia and myelodysplastic syndrome have been shown to be effective with the help of two epi drugs : azacytidine (Vidaza®) and decitabine (Dacogen®).

Although epi-drugs are widely used in clinical treatment, they have several disadvantages associated with it including fatigue and diarrhea. Chemical instability related with these drugs rapidly broke down in less than an hour and are changed into inactive compounds that can cause DNA damage and lower immune function. Also due to their lack of specificity they are also toxic to bone marrow, and can reduce blood cell counts.

## **CONCLUSION**

In conclusion , cancer is caused when cells of the body start multiplying at a fast rate without any definite differentiation and formation of a tumour which spreads to other body organs and tissue. Cancer epigenetics is the study on how your environment and behaviour can affect the work of genes. Epigenetics changes are regulated by the genes which are turned on or turned off. These changes do not change the DNA building blocks sequences . Epigenetic changes include modifications of histone that lead to cancer, DNA hypermethylation and other environmental stimuli. The study of these epigenetic changes are useful in treating various cancers through epi drugs and the advancement in this area is much needed as well as important to meet the competitive evolving rate of cancer .



## **AUTHORS CONTRIBUTION**

The authors confirm contribution to the paper as follows: M.P., N.S., R.M., S.S., T.C.: DATA COLLECTION AND MANUSCRIPT PREPARATION; D.D.Dean: STUDY CONCEPTION AND DESIGN, AND CRITICAL ANALYSIS. All authors reviewed the results and approved the final version of the manuscript.

## **ACKNOWLEDGMENT**

We express our gratitude to DEPARTMENT OF BIOTECHNOLOGY, ISABELLA THOBURN COLLEGE, for providing invaluable resources and facilities during the preparation of this research paper. Special thanks are extended to our Head of Department: Dr (Maj.) Neerja Masih and other faculty members Dr Deepika Delsa Dean, Dr Vidya Meenakshi and Dr Madhurima Tiwari for their valuable insights and assistance.

## **REFERENCES**

- [1]. Martinez J, Gerner E, Ignatenko NA, Molecular biology of cancer, 2003, 2-3.
- [2]. Alberts B, Johnson A, Lewis J, Morgan D, Raff M, Roberts K, Walter P, Molecular biology of cell, chapter 20: cancer, 1092.
- [3]. Baylin SB, and Jones PA, Epigenetic Determinants of cancer, Cold Spring Harb Perspect Biol. 2016 Sep;8(9).
- [4]. Lu Y, Chan YT, Tan HT, Li S, Feng Y, and Wang S, , Epigenetic regulation in human cancer: the potential role of epi-drug in cancer therapy, Molecular cancer volume 19, Article number: 79; 27 april (2020).
- [5]. J.S. You, P.A. Jones, Cancer Genetics and Epigenetics: Two Sides of the coin? , volume 22,10 July (2012), pages 9-20.
- [6]. Takeshima H, and Ushijima T. Accumulation of genetic and epigenetic alterations in normal cells and cancer risk, *npj Precision Oncology* volume 3, Article number: 7 , 6 March (2019).
- [7]. Holgate ST, Wenzel S, Postma DS, Weiss ST, Renz H, Sly PD. Asthma. Nat Rev Dis Primers. 2015 Sep 10;1(1):15025.
- [8]. Krämer U, Oppermann H, Ranft U, Schäfer T, Ring J, Behrendt H. Differences in allergy trends between East and West Germany and possible explanations. Clin Exp Allergy. 2010 Feb;40: 289-98.

- [9]. Krämer U, Schmitz R, Ring J, Behrendt H. What can reunification of East and West Germany tell us about the cause of the allergy epidemic?, *Clin Exp Allergy*. 2015 Jan;45:94–107.
- [10]. Renz H, Conrad M, Brand S, Teich R, Garn H, Pfefferle PI. Allergic diseases, gene-environment interactions. *Allergy*. 2011 Jul;66(Suppl 95):10–2.
- [11]. Harb H, Alashkar Alhamwe B, Garn H, Renz H, Potaczek DP. Recent developments in epigenetics of pediatric asthma. *Curr Opin Pediatr*. 2016 Dec;28:754–63.
- [12]. Potaczek DP, Harb H, Michel S, Alaskhar Alhamwe B, Renz H, Tost J. Epigenetics and allergy: from basic mechanisms to clinical applications. *Epigenomics*. 2017 Apr;9:539–571.
- [13]. Kabesch M. Early origins of asthma (and allergy). *Mol Cell Pediatr*. 2016 Dec ;3(1):31.
- [14]. Yang IV, Lozupone CA, Schwartz DA. The environment, epigenome, and asthma. *J Allergy Clin Immunol*. 2017 Jul ;140:14–23.
- [15]. Martino D, Prescott S. Epigenetics and prenatal influences on asthma and allergic airways disease. *Chest*. 2011 Mar ;139(3):640–647.
- [16]. Schieck M, Sharma V, Michel S, Toncheva AA, Worth L, Potaczek DP, et al. A polymorphism in the TH2 locus control region is associated with changes in DNA methylation and gene expression. *Allergy*. 2014 Sep ;69(9):1171–80.
- [17]. Healy S, Khan P, He S, Davie JR. Histone H3 phosphorylation, immediate-early gene expression, and the nucleosomal response: a historical perspective. *Biochem Cell Biol*. 2012 Feb; 90:39–54.
- [18]. Sawicka A, Seiser C. Histone H3 phosphorylation—a versatile chromatin modification for different occasions. *Biochimie*. 2012 Nov;94:2193–201.
- [19]. Rossetto D, Avvakumov N, Côté J. Histone phosphorylation: a chromatin modification involved in diverse nuclear events. *Epigenetics*. 2012 Oct ;7(10):1098–108.
- [20]. Swygert SG, Peterson CL. Chromatin dynamics: interplay between remodeling enzymes and histone modifications. *Biochim Biophys Acta*. 2014 Aug ;1839(8):728–36.
- [21]. Bannister AJ, Kouzarides T. Regulation of chromatin by histone modifications. *Cell Res*. 2011 Mar ;21(3):381–95.
- [22]. Gansen A, Tóth K, Schwarz N, Langowski J. Opposing roles of H3- and H4-acetylation in the regulation of nucleosome structure—a FRET study. *Nucleic Acids Res*. 2015 Feb 18 ;43(3):1433–43.
- [23]. Kurdistani SK, Tavazoie S, Grunstein M. Mapping global histone acetylation patterns to gene expression. *Cell*. 2004 Jun 11;117(6):721–33.

- [24]. Agricola E, Verdone L, Di Mauro E, Caserta M. H4 acetylation does not replace H3 acetylation in chromatin remodelling and transcription activation of Adr1-dependent genes. *Mol Microbiol*. 2006 Dec;62(5):1433–46.
- [25]. Morera L, Lübbert M, Jung M. Targeting histone methyltransferases and demethylases in clinical trials for cancer therapy. *Clin Epigenetics*. 2016 May 24;8:57.
- [26]. Sawicka A, Hartl D, Goiser M, Pusch O, Stocsits RR, Tamir IM, et al. H3S28 phosphorylation is a hallmark of the transcriptional response to cellular stress. *Genome Res*. 2014 Nov;24 (11):1808–20.
- [27]. Fierz B, Muir TW. Chromatin as an expansive canvas for chemical biology. *Nat Chem Biol*. 2012 Apr 17;8 (5):417–27.
- [28]. Angiolilli C, Baeten DL, Radstake TR, Reedquist KA. The acetyl code in rheumatoid arthritis and other rheumatic diseases. *Epigenomics*. 2017;9(4):447–461.
- [29]. Wapenaar H, Dekker FJ. Histone acetyltransferases: challenges in targeting bi-substrate enzymes. *Clin Epigenetics*. 2016 May 26; 8:59.
- [30]. Ceccacci E, Minucci S. Inhibition of histone deacetylases in cancer therapy: lessons from leukaemia. *Br J Cancer*. 2016 Mar 15 ;114(6):605–11.
- [31]. Nagy Z, Tora L. Distinct GCN5/PCAF-containing complexes function as co-activators and are involved in transcription factor and global histone acetylation. *Oncogene*. 2007 Aug 13 ;26(37):5341–57.
- [32]. Cieniewicz AM, Moreland L, Ringel AE, Mackintosh SG, Raman A, Gilbert TM, et al. The bromodomain of Gcn5 regulates site specificity of lysine acetylation on histone H3. *Mol Cell Proteom*. 2014 Nov ;13(11):2896–910.
- [33]. Fournier M, Tora L. KAT2-mediated PLK4 acetylation contributes to genomic stability by preserving centrosome number. *Mol Cell Oncol*. 2017;4(2):e1270391.
- [34]. Allis CD, Berger SL, Cote J, Dent S, Jenuwien T, Kouzarides T, et al. New nomenclature for chromatin-modifying enzymes. *Cell*. 2007 Nov 16;131(4):633–6.
- [35]. Klein BJ, Lalonde M-E, Côté J, Yang X-J, Kutateladze TG. Crosstalk between epigenetic readers regulates the MOZ/MORF HAT complexes. *Epigenetics*. 2014 Feb ;9(2):186–93.
- [36]. Lalonde M-E, Avvakumov N, Glass KC, Joncas F-H, Saksouk N, Holliday M, et al. Exchange of associated factors directs a switch in HBO1 acetyltransferase histone tail specificity. *Genes Dev*. 2013 Sep 15 ;27(18):2009–24.
- [37]. McCullough CE, Song S, Shin MH, Johnson FB, Marmorstein R. Structural and functional role of acetyltransferase hMOF K274 autoacetylation. *J Biol Chem*. 2016 Aug 26 ;291(35):18190–8.

- [38]. Yuan H, Rossetto D, Mellert H, Dang W, Srinivasan M, Johnson J, et al. MYST protein acetyltransferase activity requires active site lysine autoacetylation. *EMBO J*. 2012 Jan 4;31(1):58–70.
- [39]. Marmorstein R, Trievel RC. Histone modifying enzymes: structures, mechanisms, and specificities. *Biochim Biophys Acta*. 2009 Jan ;1789(1):58–68.
- [40]. Sun X-J, Man N, Tan Y, Nimer SD, Wang L. The role of histone acetyltransferases in normal and malignant hematopoiesis. *Front Oncol*. 2015 May 26 ;5:108.
- [41]. Brook PO, Perry MM, Adcock IM, Durham AL. Epigenome-modifying tools in asthma. *Epigenomics*. 2015;7(6):1017–32.
- [42]. Hull EE, Montgomery MR, Leyva KJ. HDAC inhibitors as epigenetic regulators of the immune system: impacts on cancer therapy and inflammatory diseases. *Biomed Res Int*. 2016 July 31;2016:8797206.
- [43]. Lombardi PM, Cole KE, Dowling DP, Christianson DW. Structure, mechanism, and inhibition of histone deacetylases and related metalloenzymes. *Curr Opin Struct Biol*. 2011 Dec;21(6):735–43.
- [44]. Kaniskan HÜ, Martini ML, Jin J. Inhibitors of protein methyltransferases and demethylases. *Chem Rev*. 2017, 2018 Feb 14;118(3):989-1068.
- [45]. Hyun K, Jeon J, Park K, Kim J. Writing, erasing and reading histone lysine methylations. *Exp Mol Med*. 2017 Apr 28;49(4):e324.
- [46]. Bennani-Baiti IM. Integration of ER $\alpha$ -PELP1-HER2 signaling by LSD1 (KDM1A/AOF2) offers combinatorial therapeutic opportunities to circumventing hormone resistance in breast cancer. *Breast Cancer Res*. 2012 Sep 17;14(5):112.
- [47]. Fang R, Barbera AJ, Xu Y, Rutenberg M, Leonor T, Bi Q, et al. Human LSD2/KDM1b/AOF1 regulates gene transcription by modulating intragenic H3K4me2 methylation. *Mol Cell*. 2010 Jul 30;39(2):222–33.
- [48]. Zippo A, Serafini R, Rocchigiani M, Pennacchini S, Krepelova A, Oliviero S. Histone crosstalk between H3S10ph and H4K16ac generates a histone code that mediates transcription elongation. *Cell*. 2009 Sep 18 ;138(6):1122–36.
- [49]. Edmondson DG, Davie JK, Zhou J, Mirnikjoo B, Tatchell K, Dent SYR. Site-specific loss of acetylation upon phosphorylation of histone H3. *J Biol Chem*. 2002 Aug 16;277(33):29496–502.

- [50]. Lo WS, Trievel RC, Rojas JR, Duggan L, Hsu JY, Allis CD, et al. Phosphorylation of serine 10 in histone H3 is functionally linked in vitro and in vivo to Gcn5-mediated acetylation at lysine 14. *Mol Cell*. 2000 Jun;5(6):917–26.
- [51]. Ravid T, Hochstrasser M. Diversity of degradation signals in the ubiquitin-proteasome system. *Nat Rev Mol Cell Biol*. 2008 Sep;9(9):679–90.
- [52]. Schwertman P, Bekker-Jensen S, Mailand N. Regulation of DNA double-strand break repair by ubiquitin and ubiquitin-like modifiers. *Nat Rev Mol Cell Biol*. 2016 May 23;17(6):379–94.
- [53]. Cao J, Yan Q. Histone ubiquitination and deubiquitination in transcription, DNA damage response, and cancer. *Front Oncol*. 2012 Mar 12;2 :26.
- [54]. Weake VM, Workman JL. Histone ubiquitination: triggering gene activity. *Mol Cell*. 2008 Mar 28 ;29(6):653–63.
- [55]. Zhang X, Li B, Rezaeian AH, Xu X, Chou P-C, Jin G, et al. H3 ubiquitination by NEDD4 regulates H3 acetylation and tumorigenesis. *Nat Commun*. 2017 Mar 16;8:14799.
- [56]. Falkenberg KJ, Johnstone RW. Histone deacetylases and their inhibitors in cancer, neurological diseases and immune disorders. *Nat Rev Drug Discov*. 2014 Sep ;13(9):673–91.
- [57]. Fujisawa T, Filippakopoulos P. Functions of bromodomain-containing proteins and their roles in homeostasis and cancer. *Nat Rev Mol Cell Biol*. 2017 Apr ;18(4):246–62.
- [58]. Tripathi SK, Lahesmaa R. Transcriptional and epigenetic regulation of T-helper lineage specification. *Immunol Rev*. 2014 Sep;261(1):62–83.
- [59]. Suarez-Alvarez B, Rodriguez RM, Fraga MF, López-Larrea C. DNA methylation: a promising landscape for immune system-related diseases. *Trends Genet*. 2012 Oct ;28(10):506–14.
- [60]. Clifford RL, Patel JK, John AE, Tatler AL, Mazengarb L, Brightling CE, Knox AJ. CXCL8 histone H3 acetylation is dysfunctional in airway smooth muscle in asthma: regulation by BET. *Am J Physiol Lung Cell Mol Physiol*. 2015 May 1;308(9):L962–72.
- [61]. Perry MM, Durham AL, Austin PJ, Adcock IM, Chung KF. BET bromodomains regulate transforming growth factor- $\beta$ -induced proliferation and cytokine release in asthmatic airway smooth muscle. *J Biol Chem*. 2015 Apr 3;290(14):9111–21.
- [62]. Comer BS, Camoretti-Mercado B, Kogut PC, Halayko AJ, Solway J, Gerthoffer WT. Cyclooxygenase-2 and microRNA-155 expression are elevated in asthmatic airway smooth muscle cells. *Am J Respir Cell Mol Biol*. 2015 Apr ;52(4):438–47.

- [63]. Clifford RL, John AE, Brightling CE, Knox AJ. Abnormal histone methylation is responsible for increased vascular endothelial growth factor 165a secretion from airway smooth muscle cells in asthma. *J Immunol.* 2012 Jul 15;189(2):819–31.
- [64]. Coward WR, Feghali-Bostwick CA, Jenkins G, Knox AJ, Pang L. A central role for G9a and EZH2 in the epigenetic silencing of cyclooxygenase-2 in idiopathic pulmonary fibrosis. *FASEB J.* 2014 Jul ;28(7):3183–96.
- [65]. Sanders YY, Hagood JS, Liu H, Zhang W, Ambalavanan N, Thannickal VJ. Histone deacetylase inhibition promotes fibroblast apoptosis and ameliorates pulmonary fibrosis in mice. *Eur Respir J.* 2014 May ;43(5):1448–58.
- [66]. Zhang X, Liu H, Hock T, Thannickal VJ, Sanders YY. Histone deacetylase inhibition downregulates collagen 3A1 in fibrotic lung fibroblasts. *Int J Mol Sci.* 2013 Oct ;14(10):19605–17.
- [67]. Liu T, Ullenbruch M, Young Choi Y, Yu H, Ding L, Xaubet A, et al. Telomerase and telomere length in pulmonary fibrosis. *Am J Respir Cell Mol Biol.* 2013 Aug ;49(2):260–8.
- [68]. Cahill KN, Raby BA, Zhou X, Guo F, Thibault D, Baccarelli A, et al. Impaired E prostanoind2 expression and resistance to prostaglandin E2 in nasal polyp fibroblasts from subjects with aspirin-exacerbated respiratory disease. *Am J Respir Cell Mol Biol.* 2016 Jan;54(1):34–40.
- [69]. Cho JS, Moon YM, Park IH, Um JY, Kang JH, Kim TH, et al. Effects of histone deacetylase inhibitor on extracellular matrix production in human nasal polyp organ cultures. *Am J Rhinol Allergy.* 2013 Jan ;27(1):18–23.
- [70]. Cho JS, Moon YM, Park IH, Um JY, Moon JH, Park SJ, et al. Epigenetic regulation of myofibroblast differentiation and extracellular matrix production in nasal polyp-derived fibroblasts. *Clin Exp Allergy.* 2012 Jun ;42(6):872–82.
- [71]. Yang Y, Wicks J, Haitchi HM, Powell RM, Manuyakorn W, Howarth PH, et al. Regulation of a disintegrin and metalloprotease-33 expression by transforming growth factor- $\beta$ . *Am J Respir Cell Mol Biol.* 2012 May ;46(5):633–40.
- [72]. Stefanowicz D, Lee JY, Lee K, Shaheen F, Koo H-K, Booth S, et al. Elevated H3K18 acetylation in airway epithelial cells of asthmatic subjects. *Respir Res.* 2015 Aug 5;16(1):95.
- [73]. Zhang Y, Leung DYM, Goleva E. Anti-inflammatory and corticosteroid-enhancing actions of vitamin D in monocytes of patients with steroid-resistant and those with steroid-sensitive asthma. *J Allergy Clin Immunol.* 2014;133:1744.e1–1752.e1.

- [74]. Borriello F, Longo M, Spinelli R, Pecoraro A, Granata F, Staiano RI, et al. IL-3 synergises with basophil-derived IL-4 and IL-13 to promote the alternative activation of human monocytes. *Eur J Immunol*. 2015 Jul;45(7):2042–51.
- [75]. Marwick JA, Tudor C, Khorasani N, Michaeloudes C, Bhavsar PK, Chung KF. Oxidants induce a corticosteroid-insensitive phosphorylation of histone 3 at serine 10 in monocytes. *PLoS ONE*. 2015 Apr 23;10(4):e0124961.
- [76]. Castellucci M, Rossato M, Calzetti F, Tamassia N, Zeminian S, Cassatella MA, Bazzoni F. IL-10 disrupts the Brd4-docking sites to inhibit LPS-induced CXCL8 and TNF- $\alpha$  expression in monocytes: implications for chronic obstructive pulmonary disease. *J Allergy Clin Immunol*. 2015 Jun 1 ;136(781–791):e9.
- [77]. Hsieh CC, Kuo CH, Kuo HF, Chen YS, Wang SL, Chao D, et al. Sesamin suppresses macrophage-derived chemokine expression in human monocytes via epigenetic regulation. *Food Funct*. 2014;5:2494–500.
- [78]. Harb H, Raedler D, Ballenberger N, Böck A, Kesper DA, Renz H, Schaub B. Childhood allergic asthma is associated with increased IL-13 and FOXP3 histone acetylation. *J Allergy Clin Immunol*. 2015 Jul;136(1):200–2.
- [79]. Seumois G, Chavez L, Gerasimova A, Lienhard M, Omran N, Kalinke L, et al. Epigenomic analysis of primary human T cells reveals enhancers associated with TH2 memory cell differentiation and asthma susceptibility. *Nat Immunol*. 2014 Aug;15(8):777–88.
- [80]. Huber JP, Gonzales-van Horn SR, Roybal KT, Gill MA, Farrar JD. IFN- $\alpha$  suppresses GATA3 transcription from a distal exon and promotes H3K27 trimethylation of the CNS-1 enhancer in human Th2 cells. *J Immunol*. 2014 Jun 15;192(12):5687–94.
- [81]. Gerasimova A, Chavez L, Li B, Seumois G, Greenbaum J, Rao A, et al. Predicting cell types and genetic variations contributing to disease by combining GWAS and epigenetic data. *PLoS ONE*. 2013;8:e54359.
- [82]. C-y Li, Peng J, L-p Ren, Gan L-x Lu, X-j Liu Q, et al. Roles of histone hypoacetylation in LAT expression on T cells and Th2 polarization in allergic asthma. *J Transl Med*. 2013 Jan 30;11:26.
- [83]. Luo S, Liang G, Zhang P, Zhao M, Lu Q. Aberrant histone modifications in peripheral blood mononuclear cells from patients with Henoch-Schönlein purpura. *Clin Immunol*. 2013 Mar ;146(3):165–75.
- [84]. Han J, Park S-G, Bae J-B, Choi J, Lyu J-M, Park SH, et al. The characteristics of genome-wide DNA methylation in naïve CD4<sup>+</sup> T cells of patients with psoriasis or atopic dermatitis. *Biochem Biophys Res Commun*. 2012 May 25;422(1):157–63.

- [85]. Dunstan JA, West C, McCarthy S, Metcalfe J, Meldrum S, Oddy WH, et al. The relationship between maternal folate status in pregnancy, cord blood folate levels, and allergic outcomes in early childhood. *Allergy*. 2012 Jan ;67(1):50–7.
- [86]. McStay CL, Prescott SL, Bower C, Palmer DJ. Maternal folic acid supplementation during pregnancy and childhood allergic disease outcomes: a question of timing? *Nutrients*. 2017.
- [87]. D’Vaz N, Meldrum SJ, Dunstan JA, Lee-Pullen TF, Metcalfe J, Holt BJ, et al. Fish oil supplementation in early infancy modulates developing infant immune responses. *Clin Exp Allergy*. 2012 Aug ;42(8):1206–16.
- [88]. Dunstan JA, Mori TA, Barden A, Beilin LJ, Taylor AL, Holt PG, Prescott SL. Fish oil supplementation in pregnancy modifies neonatal allergen-specific immune responses and clinical outcomes in infants at high risk of atopy: a randomized, controlled trial. *J Allergy Clin Immunol*. 2003 Dec ;112(6):1178–84.
- [89]. Harb H, Amarasekera M, Ashley S, Tulic MK, Pfefferle PI, Potaczek DP, et al. Epigenetic regulation in early childhood: a miniaturized and validated method to assess histone acetylation. *Int Arch Allergy Immunol*. 2015;168(3):173–81.
- [90]. Harb H, Irvine J, Amarasekera M, Hii CS, Kesper DA, Ma Y, et al. The role of PKC $\zeta$  in cord blood T-cell maturation towards Th1 cytokine profile and its epigenetic regulation by fish oil. *Biosci Rep*. 2017 Mar 27;37(2):BSR20160485.
- [91]. Kobayashi Y, Bossley C, Gupta A, Akashi K, Tsartsali L, Mercado N, et al. Passive smoking impairs histone deacetylase-2 in children with severe asthma. *Chest*. 2014 Feb ;145(2):305–12.
- [92]. Kabesch M, Adcock IM. Epigenetics in asthma and COPD. *Biochimie*. 2012 Nov ;94(11):2231–41.
- [93]. Kidd CDA, Thompson PJ, Barrett L, Baltic S. Histone modifications and asthma. the interface of the epigenetic and genetic landscapes. *Am J Respir Cell Mol Biol*. 2016 Jan;54(1):3–12.
- [94]. Liu R, Leslie KL, Martin KA. Epigenetic regulation of smooth muscle cell plasticity. *Biochim Biophys Acta*. 2015 Apr ;1849(4):448–53.
- [95]. Royce SG, Ververis K, Karagiannis TC. Histone deacetylase inhibitors: can we consider potent anti-neoplastic agents for the treatment of asthma? *Ann Clin Lab Sci*. 2012 Summer ;42(3):338–45.
- [96]. Salam MT. Asthma epigenetics. *Adv Exp Med Biol*. 2014;795:183–99.



- [97]. Tost J, Gay S, Firestein G. Epigenetics of the immune system and alterations in inflammation and autoimmunity. *Epigenomics*. 2017 Apr;9(4):371–3.
- [98]. Tost J. Engineering of the epigenome: synthetic biology to define functional causality and develop innovative therapies. *Epigenomics*. 2016 Feb ;8(2):153–6.
- [99]. Potaczek DP, Garn H, Unger SD, Renz H. Antisense molecules: a new class of drugs. *J Allergy Clin Immunol*. 2016 May;137(5):1334–46.
- [100]. Garn H, Renz H. GATA-3-specific DNzyme—a novel approach for stratified asthma therapy. *Eur J Immunol*. 2017 Jan ;47(1):22–30.
- [101]. Krug N, Hohlfeld JM, Kirsten A-M, Kornmann O, Beeh KM, Kappeler D, et al. Allergen-induced asthmatic responses modified by a GATA3-specific DNzyme. *N Engl J Med*. 2015 May 21;372(21):1987–95.
- [102]. Chen CH, Wang CZ, Wang YH, Liao WT, Chen YJ, Kuo CH, et al. Effects of low-level laser therapy on M1-related cytokine expression in monocytes via histone modification. *Mediators Inflamm*. 2014;2014:625048.
- [103]. Nestor CE, Barrenäs F, Wang H, Lentini A, Zhang H, Bruhn S, et al. DNA methylation changes separate allergic patients from healthy controls and may reflect altered CD4+ T-cell population structure. *PLoS Genet*. 2014 Jan ;10(1):e1004059.
- [104]. Martino D, Joo JE, Sexton-Oates A, Dang T, Allen K, Saffery R, Prescott S. Epigenome-wide association study reveals longitudinally stable DNA methylation differences in CD4+ T cells from children with IgE-mediated food allergy. *Epigenetics*. 2014 Jul;9(7):998–1006.
- [105]. Martino D, Dang T, Sexton-Oates A, Prescott S, Tang MLK, Dharmage S, et al. Blood DNA methylation biomarkers predict clinical reactivity in food-sensitized infants. *J Allergy Clin Immunol*. 2015 May;135(5):1319.e1-12–1328.e1-12.
- [106]. Cell signaling technology. Histone modification table. . Accessed 21 July 2017.
- [107]. Kuo CH, Lin CH, Yang SN, Huang MY, Chen HL, Kuo PL, et al. Effect of prostaglandin I2 analogs on cytokine expression in human myeloid dendritic cells via epigenetic regulation. *Mol Med*. 2012 May 9;18(1):433–44.
- [108]. Zhong F, Zhou N, Wu K, Guo Y, Tan W, Zhang H, et al. A SnoRNA-derived piRNA interacts with human interleukin-4 pre-mRNA and induces its decay in nuclear exosomes. *Nucleic Acids Res*. 2015 Dec 2;43(21):10474–91.
- [109]. Zheng J, van de Veerdonk FL, Crossland KL, Smeekens SP, Chan CM, Al Shehri T, et al. Gain-of-function STAT1 mutations impair STAT3 activity in patients with chronic mucocutaneous candidiasis (CMC). *Eur J Immunol*. 2015 Oct;45(10):2834–46.

- [110]. Vicente CT, Edwards SL, Hillman KM, Kaufmann S, Mitchell H, Bain L, et al. Long-range modulation of PAG1 expression by 8q21 allergy risk variants. *Am J Hum Genet.* 2015 Aug 6;97(2):329–36.
- [111]. Naranbhai V, Fairfax BP, Makino S, Humburg P, Wong D, Ng E, et al. Genomic modulators of gene expression in human neutrophils. *Nat Commun.* 2015 Jul 7;6:7545.
- [112]. Lin CC, Lin WN, Hou WC, Hsiao LD, Yang CM. Endothelin-1 induces VCAM-1 expression-mediated inflammation via receptor tyrosine kinases and Elk/p300 in human tracheal smooth muscle cells. *Am J Physiol Lung Cell Mol Physiol.* 2015 Aug 1 ;309(3):L211–25.
- [113]. Sharma S, Zhou X, Thibault DM, Himes BE, Liu A, Szeffler SJ, et al. A genome-wide survey of CD4(+) lymphocyte regulatory genetic variants identifies novel asthma genes. *J Allergy Clin Immunol.* 2014 Nov;134(5):1153–62.
- [114]. Escobar TM, Kanellopoulou C, Kugler DG, Kilaru G, Nguyen CK, Nagarajan V, et al. miR-155 activates cytokine gene expression in Th17 cells by regulating the DNA-binding protein Jarid2 to relieve polycomb-mediated repression. *Immunity.* 2014 Jun 19;40(6):865–79.
- [115]. Gschwandtner M, Zhong S, Tschachler A, Mlitz V, Karner S, Elbe-Bürger A, Mildner M. Fetal human keratinocytes produce large amounts of antimicrobial peptides: involvement of histone-methylation processes. *J Invest Dermatol.* 2014 Aug ;134(8):2192–201.
- [116]. Han H, Xu D, Liu C, Claesson H-E, Björkholm M, Sjöberg J. Interleukin-4-mediated 15-lipoxygenase-1 trans-activation requires UTX recruitment and H3K27me3 demethylation at the promoter in A549 cells. *PLoS ONE.* 2014 Jan 20 ;9(1):e85085.
- [117]. Lakshmi SP, Reddy AT, Zhang Y, Sciruba FC, Mallampalli RK, Duncan SR, Reddy RC. Down-regulated peroxisome proliferator-activated receptor  $\gamma$  (PPAR $\gamma$ ) in lung epithelial cells promotes a PPAR $\gamma$  agonist-reversible proinflammatory phenotype in chronic obstructive pulmonary disease (COPD). *J Biol Chem.* 2014 Mar 7 ;289(10):6383–93.
- [118]. Wiegman CH, Li F, Clarke CJ, Jazrawi E, Kirkham P, Barnes PJ, et al. A comprehensive analysis of oxidative stress in the ozone-induced lung inflammation mouse model. *Clin Sci.* 2014 Mar;126(6):425–40.
- [119]. Che W, Parmentier J, Seidel P, Manetsch M, Ramsay EE, Alkhouri H, et al. Corticosteroids inhibit sphingosine 1-phosphate-induced interleukin-6 secretion from human airway smooth muscle via mitogen-activated protein kinase phosphatase 1-mediated repression of mitogen and stress-activated protein kinase 1. *Am J Respir Cell Mol Biol.* 2014 Feb;50(2):358–68.

- [120]. Kallsen K, Andresen E, Heine H. Histone deacetylase (HDAC) 1 controls the expression of beta defensin 1 in human lung epithelial cells. *PLoS ONE*. 2012;7(11):e50000.
- [121]. Robertson ED, Weir L, Romanowska M, Leigh IM, Panteleyev AA. ARNT controls the expression of epidermal differentiation genes through HDAC- and EGFR-dependent pathways. *J Cell Sci*. 2012 Jul 15;125:3320–32.
- [122]. Vazquez BN, Laguna T, Notario L, Lauzurica P. Evidence for an intronic cis-regulatory element within CD69 gene. *Genes Immun*. 2012 Jun;13(4):356–62.
- [123]. Zijlstra GJ, ten Hacken NHT, Hoffmann RF, van Oosterhout AJM, Heijink IH. Interleukin-17A induces glucocorticoid insensitivity in human bronchial epithelial cells. *Eur Respir J*. 2012 Jun 13;39(4):439–45.
- [124]. Cao J, Yan Q, Cancer epigenetics, tumor immunity, and immunotherapy, 2020 mar 31.
- [125]. *Cancer Quest*, cancer epigenetics.
- [126]. Alison MR, Sarraf CE, *Understanding cancer*, 1997, 39-40.
- [127]. Lakshminarasimhan R, Liang G. The Role of DNA Methylation in Cancer. *Adv Exp Med Biol*. 2016;945:151-172.
- [128]. Wajed SA, Laird PW, DeMeester TR. DNA methylation: an alternative pathway to cancer. *Ann Surg*. 2001 Jul;234(1):10-20.
- [129]. Kulis M, Esteller M. DNA methylation and cancer. *Adv Genet*. 2010;70:27-56.

