**Cardiac Bioengineering**

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**Abstract :**

Cardiovascular diseases (CVDs) remain a leading cause of morbidity and mortality worldwide. In recent decades, cardiac bioengineering has emerged as a promising field with the potential to revolutionize the diagnosis, treatment, and prevention of CVDs. This abstract provides an overview of the advancements made in cardiac bioengineering, highlights the challenges faced by researchers, and discusses the future prospects of this rapidly evolving discipline. The future of cardiac bioengineering holds immense potential. Researchers are actively exploring novel approaches to enhance vascularization, such as integrating microvascular networks within the engineered tissues. Advancements in induced pluripotent stem cell (iPSC) technology have opened new avenues for personalized medicine, enabling the derivation of patient-specific cardiac cells for transplantation. Integration of nanotechnology and gene editing techniques offers the possibility of targeted delivery of therapeutic agents and manipulation of cellular functions. Additionally, the advent of artificial intelligence and machine learning can aid in the identification of optimal biomaterials, cell types, and culture conditions for bioengineering cardiac tissues.

Key words: Cardiovascular Bioengineering; cell-free patch; pluripotent stem cells; Heart diseases;3D Printing

**1.Introduction:**

Heart disease is a leading cause of death worldwide, despite advancements in the current methods of diagnosis and treatment, including pharmaceutical therapy and revascularization treatments, heart disease continues to have a poor prognosis and outcomes [1,2]. To add to it, mammalian hearts are the least capable of regeneration among tissues and organs; in humans, most of the cardiomyocytes turnover rates are between 0.3-1 % annually. [3]. Cardiac bioengineering offers a promising approach by leveraging advances in stem cell biology, biomaterials science, and tissue engineering techniques to engineer living tissues that can replace or repair damaged heart tissue.

Coronary artery bypass surgery, coronary reperfusion therapy, and fibrinolytic therapy are examples of traditional treatments for MI that focus more on short-term symptom relief than on promoting the repair and regeneration of the injured myocardium [4]. The golden standard method for treating heart failure patients is a left ventricular assist device (LVAD) or a heart transplant [5]. The difficulty of the invasive transplant operation and its associated acute or chronic immune rejections results in prognosis to vary [6,7].

The effective pharmacological treatments for Myocardial infarction patients include β- -blockers and ACE inhibitors [8,9] these current approaches necessitate the investigation of novel treatment modalities that aim to regenerate the infarcted myocardium while also being feasible for use in clinical settings [10].

Recent developments in stem cell biology and biotechnologies, such as cardiac tissue engineering and human pluripotent stem cells (hPSCs), show considerable potential for establishing new therapeutic routes for heart regeneration and repair in the treatment of heart disease [11,12].

Over the past ten years, scientists have started to realize that paracrine processes are principally responsible for the beneficial clinical and functional results of stem cell therapy [13]. So-called non-coding RNAs including microRNA (miRNA), long non-coding RNAs (lncRNAs), and exosomes have drawn growing interest in emerging cell-free methods [14]. Alternatively, utilizing a unique cocktail of transcription factors that were verified in vitro and in vivo, researchers have recently revealed the idea of direct reprogramming of scar tissue after MI into a cardiomyocyte [15]. This system is still infancy requires preclinical validation safety and efficacy.

Recent studies reveals that 95% of delivered cells to the injured cardiomyocytes are lost due to poor oxygenation, host immune reaction and less nutrients. The cell-based therapy with bioengineering technologies inclusive of functional tools such as biomaterials and bio fabrications can stimulate the engraftment of transplanted cells [16-19].

Cardiac bioengineering offers a promising approach by leveraging advances in stem cell biology, biomaterial science, and tissue engineering techniques to engineer living tissues that can replace or repair damaged heart tissue. In this chapter we summarize the Progress in cardiac tissue engineering techniques, with a focus on cardiomyocyte maturation and stem cell engineering, as well as the creation of novel functional biomaterials (like hydrogels and decellularized scaffolds) and bio fabrication tools (like engineered heart tissues and bioprinting) and their therapeutic uses in the areas of drug discovery, disease modelling, and heart disease regenerative medicine which is also represented in Figure 1.1. Finally, we go through the present difficulties and potential futures of cardiac tissue engineering technologies from the standpoint of their therapeutic importance.



**FIGURE 1.1 Represents Cardiac Bioengineering**

**2.Over View of Cardiovascular Bioengineering in therapeutics:**

Bioengineering methods for cardiac applications involve the use of various techniques and approaches to study, repair, or enhance the structure and function of the heart. These methods aim to address cardiac diseases, improve cardiac regeneration, and develop new therapies for cardiovascular conditions.

The goal of biomedical engineering is to improve medical care, involving examination, observation, and treatment. The heart regenerating methods have been evolving into viable CVD therapeutics as a result [20]. Various biotechnologies have been used to develop treatments for myocardial that has been injured by ischemic/reperfusion or has undergone an infarction. Patient-specific cardiomyocytes (CMs) were produced using cell reprogramming technologies with the intention of utilizing autologous cells for cardiac treatment [21]. Various viral and non-viral vectors have been utilized for gene editing to affect gene expression throughout the cardiac remodeling process in order to increase the capacity of CMs to regenerate.[22,23].Cell therapy interventions, retention, tumorigenicity, immunogenicity can be overcome by isolating cell-derived protein factors, miRNAs and exosomes through nanosized or micro particles [24] Scientist also claimed that usage of antibodies, proteins and platelet membranes can aim for therapeutic targets. For better retention and integration, cardiac patches have been designed by transfer therapeutics in vehicles made of various biomaterials [25] Additionally, 3D printing [26] and 3D culture [27] technologies were utilized to create replaceable cardiac tissue represented in figure 2.1



**FIGURE 2.1** **Represents Bioengineering methods including scaffolds,cell sheets, decellularized heart tissues,3D bioprinting and cell free patches.**

**3.Precision Therapies for Cardiac Diseases:**

**3.1 Cell Reprogramming:**

Cell reprogramming refers to the process of converting special cells such as somatic cells into different cell type with a broader range of potential functions. This technique has been successfully used to generate induced pluripotent stem cells (iPSCs), cardiomyocytes (CMs), endothelial cells (ECs), and other cell types.

**3.1.1** **Stem cell engineering in Cardiomyocytes:**

Human induced pluripotent stem cells and human embryonic stem cells both have a strong capacity for cell renewal and differentiation in any portion of the body [28,29]. It is even capable of differentiating into vascular smooth muscle cells, vascular endothelial cells, and cardiomyocytes. These human heart-forming cells can be produced in vitro, making them more accessible for use in cardiac bioengineering, disease models, and regenerative therapeutics.

**3.1.1 Types of Cell Reprogramming:**

**a. Indirect cell reprogramming:**

It is the process of transforming indirect cell reprogramming from adult somatic cells to induced pluripotent stem cell-derived cardiomyocytes(iPSC-CMs) for cardiac regenerative medicine.

Induced pluripotent stem cells (iPSCs) are a type of stem cell that can be generated from adult somatic cells through a reprogramming process. iPSCs have the ability to differentiate into various types, including cardiomyocytes. They are commercially available and have become a popular choice in research and clinical applications due to their patient -specific genomic information which could be used for cardiac regenerative medicine.

* **Reprogramming process:** The process of converting adult fibroblasts into iPSCs includes various steps such as activation of alkaline phosphatase, silencing of somatic -specific gene expression, expression of SSEA1 (a marker of pluripotency), and progressive silencing of exogenous genes with upregulation of Oct4 and Nanog (critical pluripotency markers). [30]
* **iPSCs and their advantages:** According to recent research clearly states that “Induced pluripotent stem cells (iPSCs) are a type of stem cell that can be generated from adult somatic cells through a reprogramming process” . iPSCs have the ability to differentiate into various cell types, including cardiomyocytes. They are commercially available and have become a popular choice in research and clinical applications due to their patient-specific genomic information, which could be used for autologous (self-derived) cardiac regenerative medicine [31,].
* **Limitations of iPSC-CMs:** While iPSC-CMs have shown great promise for cardiac regenerative medicine, they still exhibit certain immaturity in terms of marker expression, ultrastructural features, metabolic signature, and electrophysiological properties compared to fully matured native cardiomyocytes [32]
* **Distinct metabolic flow technology:** To overcome purification obstacles and obtain mature iPSC-CMs, a metabolic flow technology has been developed. This involves large-scale purification through glucose depletion and lactate supplementation. Mature iPSC-CMs have a higher oxygen consumption rate and increased mitochondrial maturity, making them more suitable for transplantation.[33].

**b. Direct cell Reprogramming:**

Direct cell reprogramming as a potential approach for cardiac tissue repair, where somatic cells are transformed into desired cell types without going through a pluripotent or multipotent state. This process is considered more suitable for in vivo cardiac regeneration as it involves generating reprogrammed cells directly in the diseased heart. However, there are several challenges associated with in vivo direct cell reprogramming. It also holds promise for cardiac tissue repair and regeneration, it still faces challenges related to efficiency, safety, and long-term outcomes. Researchers continue to investigate and refine these reprogramming methods to overcome these obstacles and potentially develop more effective strategies for cardiac regenerative medicine [34].

* **Low transforming efficiency:** One of the main challenges of in vivo direct cell reprogramming is achieving efficient and reliable cell conversion. The process of introducing transcription factors or miRNAs to initiate the reprogramming is not always highly efficient, limiting its effectiveness for cardiac tissue repair.[35]
* **Transcription factors for direct reprogramming:** Transcription factors like Gata4, Oct4, Tbx5, Sox2, and Klf4 have been explored for direct reprogramming of somatic cells into cardiac cell lineages. While some studies have shown promising results, the overall efficiency remains a concern.[36]
* **Tumorigenic risks:** Direct cell reprogramming does not completely eliminate tumorigenic risks associated with reprogramming approaches. The use of retroviruses or other methods to deliver transcription factors or miRNAs could still pose potential risks of tumorigenesis, and ensuring the safety of this approach is crucial.[37].
* **Fate of transduced cells:** Understanding the fate of transduced cells after in vivo direct reprogramming is essential. While single-cell transcriptomics can provide insights into the mechanisms of fate conversion from fibroblasts to cardiomyocytes, there may still be uncertainties about the long-term behaviours and stability of the reprogrammed cells.
* **miRNAs as an alternative:** MicroRNAs (miRNAs) have also been explored as an alternative for cardiac reprogramming. miRNAs can regulate multiple signaling pathways simultaneously, making them a promising option for inducing cell fate changes. However, similar to transcription factor-based reprogramming, achieving sufficient efficiency with miRNA-based approaches remains a challenge.
* **Enhancement with JAK inhibitor:** Some studies have shown that combining certain miRNAs with the JAK inhibitor I can enhance cardiac reprogramming. This highlights the potential for synergistic effects by combining different approaches to improve the efficiency of direct cell reprogramming [38].

**3.2 Gene therapy:**

Numerous genes are necessary for CM growth and heart healing. By encouraging cardiomyocyte dedifferentiation and proliferation through YAP activation and EMT (epithelial-mesenchymal transition)-like processes, for instance, ERBB2 has been found to stimulate mammalian heart regeneration [39]. In order to stimulate cardiac repair, Cyclin A2, also known as CCNA2, a gene that is generally silenced after birth, has been shown to be a crucial cell cycle regulatory gene by mediating both the G1-S and G2-M transitions of the cell cycle [40].Therefore, for cardiac gene therapy, effective transport of the desired gene to the heart is crucial.

Gene therapy for cardiac regeneration is an approach aimed at improving heart function by introducing therapeutic genes into the heart cells (cardiomyocytes). The target protein encoded by the introduced DNA can either repair essential proteins for normal cardiac function or knock down proteins that may negatively affect heart function. However, there are several challenges associated with this approach [41].

1. **Gene transfer efficiency:** One of the main problems in cardiac gene therapy is achieving efficient gene transfer into the cardiomyocytes. For plasmid transfection, where naked DNA is introduced, the uptake by cardiomyocytes is too low, and measurable levels of gene expression are only detectable for a short period, limiting the therapeutic effect.
2. **Safety concerns with viral vectors:** Viral vectors, such as adenovirus, adeno-associated virus (AAV), and lentivirus, are more efficient in gene transfer than plasmids. However, adenoviral vectors can stimulate strong immune and inflammatory responses, which can be fatal. The dose of vector that can be injected is also limited. AAV vectors have lower immunogenicity, but their permanent expression of the transgene restricts their use.
3. **Lack of control over gene expression:** Currently, there is no effective method for regulating gene expression in vivo once the gene is delivered to the target cells. This permanent expression may not be desirable in some cases.
4. **Identifying the appropriate target gene(s):** Repairing a single gene may not be sufficient for complete cardiac repair, except for cases of enzymatic deficiencies with specific mutations. Cardiovascular gene therapy trials have started to focus on combinations of genes for better outcomes.
5. **Lack of an appropriate cell source:** To effectively express the required gene in injured hearts, a suitable source of regenerative cells is necessary. This aspect needs further research and development.

Despite several clinical trials in cardiovascular gene therapy over the last three decades, there has not been a significant breakthrough with a substantial impact on heart failure or acute myocardial infarction (MI). This has led to the realization that gene therapy might not be the optimal solution for addressing these cardiac deficiencies. Even though gene therapy holds promise as a potential approach for cardiac regeneration, there are still significant challenges to overcome before it can be widely used as an effective treatment in the cardiovascular field. Continued research and advancements in gene delivery techniques and gene expression control are needed to unlock the full potential of gene therapy for heart-related conditions.

Non-viral gene delivery methods offer an alternative to viral vectors in cardiac gene therapy. These methods utilize natural or synthetic compounds or physical forces to deliver the desired gene to the target tissue. Some of the non-viral gene delivery methods include:

1. **Needle or Jet Injection:** This method involves using a fine needle or a jet device to directly inject the gene of interest into the target tissue. However, the jet injection method with high-force piercing is not suitable for cardiac applications due to the potential damage it may cause.
2. **Hydrodynamic Gene Transfer:** Hydrodynamic gene transfer involves the rapid injection of a large volume of solution containing the desired gene into the bloodstream. The force generated by this rapid injection can help deliver the gene to some cells, but the efficiency is still relatively low.
3. **Electroporation:** Electroporation uses short electrical pulses to create temporary pores in the cell membrane, allowing the gene to enter the cells more effectively. This method has shown promise in improving gene delivery efficiency, but optimization is required for cardiac applications.
4. **Cationic Lipids:** Cationic lipids can form complexes with the gene of interest, creating liposomes that facilitate cellular uptake. They are easy to modify and can be designed for cell specificity, making them a valuable tool for gene delivery.

While non-viral gene delivery methods have advantages like low toxicity, ease of modification, and cell specificity, their efficiency in delivering the gene to target cells remains a challenge. For instance, hydrophilic naked DNA may be taken up by cells, but only a small percentage of the target cells will express the delivered genes, making them inefficient for widespread gene therapy applications.

Researchers continue to work on improving non-viral gene delivery methods to enhance their efficiency and specificity for cardiac gene therapy. Addressing these challenges will be crucial for advancing the field of cardiac regenerative medicine using non-viral gene delivery techniques.

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**4.0 Fabrication of Biomimetic Cardiac Tissues**

The fabrication of biomimetic cardiac tissues involves creating artificial tissues that closely resemble the structure and function of native heart tissues. These tissues can serve as models for studying cardiac biology, drug testing, and potentially as a source for cardiac tissue repair and regeneration. Several approaches are being explored for the fabrication of biomimetic cardiac tissues [42].

**Hydrogels:**

One of the best polymers for cardiac tissue engineering is hydrogel because of its biocompatibility and adaptable chemical and physical properties. One benefit of employing polymers is their potential to contain cell-binding sites like ECM proteins and RGD integrin-binding domains, as well as their high water content, scalability, and flexibility efficient metabolite exchange. Natural hydrogels for creating heart tissues. The application of hydrogels as functional engineered tissues may be constrained by poor cell adherence caused by the inherent cell repellency of most hydrogels, such as poly (ethylene glycol) or zwitterionic hydrogels [43].

Due to their innate softness and large porosity, macroporous hydrogels' lower modulus in comparison to real tissues may affect CM maturation [44].

Proteoglycans, glycoproteins, fibrous protein growth factors, and other tiny molecules make up the natural cardiac ECM's lack of biological complexity and are crucial for regulating cell-ECM interaction. Although a promising material, its application in cardiac tissue engineering needs to be investigated further.

**Bioprinting:**

A new technique for fabricating tissue-engineered constructs is called "bioprinting," which involves precisely layer-by-layer deposition of biomaterials, biochemicals, and living cells [45]. Technologies for bioprinting include inkjet printing and laser-assisted printing **[46**]. Researchers created an acellular neonatalsized heart analogue (37 mm diameter, 55 mm height) and the beating ventricle (truncated ellipsoid; 5.7 mm diameter, 8 mm height) using this method [47].. In another study, the FRESH technique was used to print a heart-shaped model (14 mm in diameter and 20 mm in height) using a bioink made of a hydrogel from a patient's decellularized omentum and the patient's own cells (CMs and endothelial cells)[48].

Microfabrication:

The proposed synthetic 3D cell niches can replicate the natural cell microenvironment thanks to microfabrication. There are many ways to make surface patterns, including lithography methods (or microprinting), which include photolithography and soft-lithography.By transferring the geometric pattern from a photomask to a light-sensitive chemical on the substrate under the light, one can create defined topographies for anisotropic tissue organisation via photolithography. Simple manufacturing techniques like hydrogels are unable to create cardiac tissues with perfusable and highly branching endothelialized channels as the vascular network. This can be accomplished by trying the simple soft lithography technique[49,50].

**5.0 Decellularization and Recellularization Methods:**

Decellularization and recellularization are innovative techniques used in tissue engineering and regenerative medicine. These methods involve the removal of cellular components from a tissue or organ (decellularization) and subsequent repopulation with new cells (recellularization). The goal is to create bioengineered tissues or organs that can be used for transplantation, disease modelling, drug testing, and more. The methods are explained in detail below:

1. Decellularization: Decellularization is the process of removing cellular material, including cells, cell debris, and cell-associated proteins, from a tissue or organ while preserving the underlying extracellular matrix (ECM). The ECM is a complex network of proteins and molecules that provide structural support and biochemical cues for cells. By eliminating the cellular components, the risk of immune rejection upon transplantation can be significantly reduced, as the new tissue can be populated with cells from the recipient without triggering an immune response.

**Methods of Decellularization:** Several methods can be used for decellularization, including:

* Chemical methods: Using detergents, such as sodium dodecyl sulfate (SDS), Triton X-100, or ethylenediaminetetraacetic acid (EDTA), to disrupt and solubilize cellular membranes and remove cellular content.
* Physical methods: Applying mechanical forces, such as agitation, to break down cells and release their contents.
* Enzymatic methods: Using enzymes, such as nucleases and proteases, to degrade nucleic acids and proteins, respectively, thereby facilitating cell removal.
* Perfusion-based methods: These involve the perfusion of decellularization solutions through blood vessels within the tissue or organ to ensure thorough cell removal.
1. Recellularization: Recellularization is the subsequent step after decellularization, where the acellular scaffold created through the decellularization process is repopulated with new cells. The cells used for recellularization can be derived from various sources, such as the patient's own cells (autologous), donor cells (allogeneic), or stem cells. The chosen cell type depends on the application and the desired tissue or organ.

**Methods of Recellularization:** There are different techniques for recellularization, including:

* Perfusion: Cells are delivered through the blood vessels of the decellularized tissue or organ using a perfusion system. This method is often used for larger tissues and organs.
* Seeding: Cells are directly seeded onto the surface of the scaffold, allowing them to attach and proliferate within the ECM.
* Bioprinting: In this advanced technique, cells are incorporated into bioinks and then deposited layer-by-layer to recreate the complex tissue structure.
1. Combination of Decellularization and Recellularization: Combining both decellularization and recellularization processes results in tissue-engineered constructs with a biologically functional ECM and living cells. This approach holds great promise in the field of regenerative medicine, where it could provide a renewable and patient-specific source of organs and tissues for transplantation.

It is important to note that decellularization and recellularization processes are still active areas of research, and the specific protocols and techniques may vary depending on the tissue or organ being targeted and the intended application **[51,52,53].**

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**6.0 Whole-Heart Engineering**

Whole heart engineering, also known as whole heart regeneration or bioengineered heart, is a cutting-edge area of research in tissue engineering and regenerative medicine. The ultimate goal of whole heart engineering is to create a functional and transplantable heart using a combination of decellularization and recellularization techniques, as well as other advanced bioengineering approaches. The aim is to address the critical shortage of donor hearts for transplantation and to provide personalized and immune-compatible solutions for patients with end-stage heart failure.

The process of whole heart engineering involves several key steps:

1. Heart Decellularization: The first step is to obtain a donor heart or an animal heart that is structurally similar to a human heart. The heart is then subjected to the decellularization process, during which the cellular components, including cardiomyocytes (heart muscle cells), endothelial cells, and other cell types, are removed, leaving behind the acellular heart scaffold. This scaffold consists of the extracellular matrix (ECM), which provides the framework and biochemical cues necessary for cell attachment and tissue formation.
2. Recellularization: After successful decellularization, the acellular heart scaffold needs to be repopulated with new cells. Various cell types are required to recreate the different components of the heart, such as cardiomyocytes, endothelial cells, fibroblasts, and potentially others. The cells used for recellularization can be derived from the patient's own cells (autologous) or other sources (allogeneic or stem cells). The challenge lies in effectively populating the scaffold with the right cell types and ensuring their proper organization to generate functional heart tissue.
3. Maturation and Functionality: Once the recellularization process is complete, the bioengineered heart undergoes maturation, where the cells align, differentiate, and establish functional connections similar to those found in a native heart. Maturation techniques include electrical and mechanical stimulation to mimic the natural physiological conditions in which the heart develops and functions.
4. Testing and Validation: Bioengineered hearts need to undergo rigorous testing and validation to ensure that they function properly and can withstand the demands of the circulatory system. This involves assessing their contractile ability, electrical conductivity, and response to various stimuli, as well as testing their long-term stability and safety.
5. Transplantation and Clinical Application: If successfully developed and validated, bioengineered hearts could be transplanted into patients with end-stage heart failure, providing a potentially curative treatment option. Additionally, these bioengineered hearts could serve as invaluable tools for drug testing, disease modelling, and understanding heart development and pathophysiology.

While whole heart engineering holds great promise, it is a highly complex and challenging field that requires multidisciplinary collaboration among experts in tissue engineering, cardiac biology, biomaterials, and transplantation medicine. As of my last update in September 2021, whole heart engineering was still largely in the realm of preclinical research and had not yet been translated into routine clinical practice. Nevertheless, ongoing advancements in the field offer hope for the development of viable bioengineered hearts in the future.**Top of Form**

**7.0 Challenges Should Be Addressed for Whole-Heart Engineering:**

Whole-heart engineering is an ambitious and complex endeavor that faces several significant hurdles that need to be addressed to achieve success in creating functional bioengineered hearts. Some of the key challenges include:[59,60]

1. Cell Sourcing and Differentiation: Obtaining a sufficient number of high-quality cells for recellularization is a critical challenge. Differentiating stem cells into the required cell types, especially functional cardiomyocytes, and ensuring their proper integration and function within the heart scaffold is a major obstacle.
2. Vascularization: Proper vascularization is essential for supplying nutrients and oxygen to the entire bioengineered heart. Ensuring that blood vessels can effectively integrate into the engineered tissue and support the metabolic demands of the heart remains a significant challenge.
3. Electrical Integration: The heart's coordinated beating relies on a complex electrical conduction system. Replicating this electrical integration within the bioengineered heart to ensure synchronized and efficient pumping is a formidable task.
4. Immunological Compatibility: To avoid rejection by the recipient's immune system, whole-heart engineering must address immunological compatibility. Developing strategies to suppress immune responses and ensure long-term acceptance of the bioengineered heart is critical.
5. Maturation and Functionality: Achieving the full functional maturity of the bioengineered heart tissue remains a challenge. Ensuring that the cells can contract rhythmically, respond appropriately to physiological stimuli, and adapt to changing conditions is vital.
6. Scalability: Current methods for whole-heart engineering are often time-consuming and resource-intensive, limiting the scalability of the process. Developing efficient and reproducible techniques to generate bioengineered hearts is essential for potential clinical application.
7. Biomechanical Properties: The mechanical properties of the bioengineered heart, such as its ability to withstand pressure and pumping forces, need to be carefully considered to ensure long-term viability and functionality.
8. Ethical Considerations: The development of whole-heart engineering raises ethical questions about using animals for research and the potential creation of human-animal hybrids for organ generation. Ethical guidelines and regulations must be carefully considered and adhered to during research.
9. Clinical Translation and Regulatory Approval: Before bioengineered hearts can be used in human patients, they must undergo extensive preclinical testing and clinical trials to demonstrate safety and efficacy. Regulatory approval from relevant authorities will be necessary.
10. Long-Term Viability and Durability: Ensuring the long-term viability and durability of the bioengineered heart is crucial for its clinical application. Understanding how the bioengineered heart responds to aging, disease, and other stressors is essential.

Addressing these hurdles requires collaborative efforts from researchers in various disciplines, including tissue engineering, cell biology, biomaterials, immunology, and transplantation medicine. Additionally, continued investment in research, technological advancements, and a deep understanding of the complexities of the heart and its interactions are vital to overcome these challenges and bring whole-heart engineering closer to clinical reality.**Top of Form**

**8.0 Cardiac Patch:**

A cardiac patch, also known as a cardiac tissue patch or heart patch, is a bioengineered construct designed to repair damaged or injured heart tissue. It serves as a regenerative therapy to restore heart function and improve cardiac performance, particularly after a heart attack (myocardial infarction) or in cases of heart failure.The cardiac patch is typically made from biocompatible materials that mimic the extracellular matrix (ECM) of the heart and may include a combination of cells, growth factors, and biomaterials. The primary goal of the cardiac patch is to promote tissue regeneration, prevent adverse remodeling, and support the injured heart's healing process [61.62].

**Components of a Cardiac Patch:**

1. Biomaterial Scaffold: The cardiac patch's structural foundation is a biomaterial scaffold, which provides mechanical support and serves as a platform for cell attachment and tissue growth. The ideal biomaterial should have properties that closely resemble the heart's ECM to facilitate integration with the native tissue.
2. Cardiomyocytes: Cardiomyocytes are the heart muscle cells responsible for the heart's contractile function. Including viable and functional cardiomyocytes in the cardiac patch helps restore contractility to the damaged area, improving the heart's overall pumping ability.
3. Supporting Cells: In addition to cardiomyocytes, the cardiac patch may contain other types of supporting cells, such as endothelial cells and fibroblasts. Endothelial cells promote blood vessel formation (angiogenesis) within the patch, enhancing its blood supply and oxygen delivery. Fibroblasts play a role in tissue repair and remodeling.
4. Growth Factors and Bioactive Molecules: Growth factors and bioactive molecules are included in the cardiac patch to stimulate cell proliferation, differentiation, and tissue regeneration. They promote the migration of cells to the damaged area and modulate the healing process.

**Application of Cardiac Patch:**

When a patient suffers from heart damage, such as in the case of a heart attack, a cardiac patch can be surgically implanted onto the affected area of the heart. The patch adheres to the damaged tissue and provides structural support while releasing bioactive molecules to promote tissue repair and regeneration. Over time, the cardiac patch integrates with the host tissue, and the implanted cells may begin to form new blood vessels and contractile tissue, leading to the restoration of heart function[63]

Benefits of Cardiac Patch:

* Minimally Invasive: Cardiac patches can be delivered through minimally invasive procedures, reducing the risks associated with open-heart surgery.
* Localized Therapy: The patch specifically targets the damaged area, enhancing the regenerative potential at the site of injury.
* Reduced Scar Formation: By promoting tissue regeneration, the cardiac patch may help reduce scar formation, which is common after a heart attack.

While cardiac patches show promising results in preclinical studies and early-phase clinical trials, further research and refinement are necessary to optimize their efficacy and safety. They are part of the broader field of regenerative medicine, offering hope for improved treatments for heart conditions in the future[64].**Top of FormBottom of Form**

**9. The Role of Cardiac Tissue Engineering in Clinic:**

Cardiac tissue engineering plays a crucial role in the clinic by offering innovative regenerative therapies and treatment strategies for various heart conditions. Its goal is to repair or replace damaged heart tissue and restore cardiac function, addressing the limitations of traditional treatments for heart diseases. Here are some key roles of cardiac tissue engineering in the clinical setting:[65.66].

1. **Heart Repair and Regeneration:** One of the primary roles of cardiac tissue engineering is to promote the repair and regeneration of damaged or injured heart tissue. By using bioengineered constructs, such as cardiac patches or tissue grafts, the aim is to restore functional tissue at the site of injury, ultimately improving heart function.
2. **Myocardial Infarction (Heart Attack) Therapy:** Cardiac tissue engineering offers potential therapies for patients who have experienced a myocardial infarction (heart attack). After a heart attack, there is irreversible damage to the heart tissue due to lack of blood supply. Cardiac patches or engineered tissues can be applied to the infarcted area to replace damaged tissue and prevent adverse remodeling.
3. **Heart Failure Treatment:** For patients with heart failure, where the heart's ability to pump blood efficiently is compromised, cardiac tissue engineering provides strategies to augment or replace the dysfunctional cardiac muscle. Engineered cardiac tissues, containing functional cardiomyocytes, can be transplanted into the heart to improve contractile function.
4. **Minimally Invasive Procedures:** Cardiac tissue engineering offers the potential for minimally invasive procedures, reducing the need for open-heart surgeries. These less invasive approaches can lead to faster recovery times and lower risks for patients.
5. **Patient-Specific Treatment:** Tissue engineering allows for the creation of patient-specific treatments. By using the patient's own cells (autologous cells) to create bioengineered constructs, the risk of immune rejection is minimized, increasing the likelihood of successful treatment outcomes.
6. **Drug Testing and Disease Modeling:** In addition to direct patient treatment, cardiac tissue engineering has an essential role in drug testing and disease modeling. Bioengineered heart tissues can be used to study the effects of drugs, understand disease mechanisms, and screen potential therapeutic agents, leading to more efficient drug development.
7. **Implantable Bioartificial Organs:** Advanced cardiac tissue engineering aims to develop implantable bioartificial hearts or ventricles. Although still in early stages, these bioartificial organs could serve as temporary solutions for patients awaiting heart transplantation or as long-term alternatives to donor organs.
8. **Understanding Heart Development and Function:** Cardiac tissue engineering allows researchers to study heart development, structure, and function in a controlled laboratory setting. This understanding is essential for advancing our knowledge of heart biology and pathophysiology.[67]

Despite the significant progress made in cardiac tissue engineering, challenges remain. Issues such as scalability, long-term functionality, immunological responses, and the need for large-scale clinical trials need to be addressed for widespread clinical adoption[68].

In conclusion, cardiac tissue engineering holds great promise for revolutionizing cardiovascular medicine by providing regenerative therapies, personalized treatments, and improved drug testing platforms, ultimately improving the quality of life for patients with heart conditions. As the field continues to advance, it has the potential to significantly impact the landscape of cardiology and patient care in the clinic[69,70].

**References:**

1. Roth, G.A.; Johnson, C.; Abajobir, A.; Abd-Allah, F.; Abera, S.F.; Abyu, G.; Ahmed, M.; Aksut, B.; Alam, T.; Alam, K.; et al. Global, regional, and national burden of cardiovascular diseases for 10 causes, 1990 to 2015. J. Am. Coll. Cardiol. 2017, 70, 1–25.
2. Dai, H.; Zhang, Q.; Much, A.A.; Maor, E.; Segev, A.; Beinart, R.; Adawi, S.; Lu, Y.; Bragazzi, N.L.; Wu, J. Global, regional, and national prevalence, incidence, mortality, and risk factors for atrial fibrillation, 1990–2017: Results from the Global Burden of Disease Study 2017. Eur. Heart J. Qual. Care Clin. Outcomes 2021, 7, 574–582.
3. Alonzo, M., AnilKumar, S., Roman, B., Tasnim, N., & Joddar, B. (2019). 3D Bioprinting of cardiac tissue and cardiac stem cell therapy. *Translational Research*, *211*, 64-83.
4. Awada, H. K., Hwang, M. P., and Wang, Y. (2015). Towards comprehensive cardiac repair and regeneration after myocardial infarction: aspects to consider and proteins to deliver. Biomaterials 82, 94–112. doi: 10.1016/j.biomaterials.2015. 12.025
5. Rose, E. A., Gelijns, A. C., Moskowitz, A. J., Heitjan, D. F., Stevenson, L. W., Dembitsky, W., et al. (2001). Long-term use of a left ventricular assist device for end-stage heart failure. N. Engl. J. Med. 345, 1435–1443.
6. White, H. D., and Chew, D. P. (2008). Acute myocardial infarction. Lancet 372, 570–584.
7. Wilhelm, M. J. (2015). Long-term outcome following heart transplantation: current perspective. J. Thorac. Dis. 7, 549–551.
8. Packer, M., Coats, A. J., Fowler, M. B., Katus, H. A., Krum, H., Mohacsi, P., et al. (2001). Carvedilol prospective randomized cumulative survival study group. Effect of carvedilol on survival in severe chronic heart failure. N. Engl. J. Med. 344, 1651–1658.
9. McMurray, J. J., Packer, M., Desai, A. S., Gong, J., Lefkowitz, M. P., Rizkala, A. R., et al. (2014). Angiotensin-neprilysin inhibition versus enalapril in heart failure. N. Engl. J. Med. 371, 993–1004.
10. Raziyeva, K., Smagulova, A., Kim, Y., Smagul, S., Nurkesh, A., and Saparov, A. (2020). Preconditioned and genetically modified stem cells for myocardial infarction treatment. Int. J. Mol. Sci. 21:7301. doi: 10.3390/ijms21197301
11. Witman, N.; Zhou, C.; Beverborg, N.G.; Sahara, M.; Chien, K.R. Cardiac progenitors and paracrine mediators in cardiogenesis and heart regeneration. Semin. Cell Dev. Biol. 2020, 100, 29–51. [CrossRef] [PubMed]
12. Florian, W.; Ingra, M.; Thomas, E. Engineering cardiac muscle tissue: A maturating field of research. Circ. Res. 2017, 120, 1487–1500.
13. Kitsara, M., Agbulut, O., Kontziampasis, D., Chen, Y., & Menasché, P. (2017). Fibers for hearts: A critical review on electrospinning for cardiac tissue engineering. *Acta biomaterialia*, *48*, 20-40.
14. Wang, L., Wu, Y., Hu, T., Guo, B., & Ma, P. X. (2017). Electrospun conductive nanofibrous scaffolds for engineering cardiac tissue and 3D bioactuators. *Acta biomaterialia*, *59*, 68-81.
15. Li, Y., Huang, G., Zhang, X., Wang, L., Du, Y., Lu, T. J., & Xu, F. (2014). Engineering cell alignment in vitro. *Biotechnology advances*, *32*(2), 347-365.
16. Sheikh, A.Y.; Lin, S.A.; Cao, F.; Cao, Y.; van der Bogt, K.E.; Chu, P.; Chang, C.P.; Contag, C.H.; Robbins, R.C.; Wu, J.C. Molecular imaging of bone marrow mononuclear cell homing and engraftment in ischemic myocardium. Stem Cells 2007, 25, 2677–2684. [CrossRef]
17. van den Akker, F.; Feyen, D.A.; van den Hoogen, P.; van Laake, L.W.; van Eeuwijk, E.C.; Hoefer, I.; Pasterkamp, G.; Chamuleau, S.A.; Grundeman, P.F.; Doevendans, P.A.; et al. Intramyocardial stem cell injection: Go(ne) with the flow. Eur. Heart J. 2017, 38, 184–186. [CrossRef]
18. Madonna, R.; Van Laake, L.W.; Davidson, S.M.; Engel, F.B.; Hausenloy, D.J.; Lecour, S.; Leor, J.; Perrino, C.; Schulz, R.; Ytrehus, K.; et al. Position paper of the european society of cardiology working group cellular biology of the heart: Cell-based therapies for myocardial repair and regeneration in ischemic heart disease and heart failure. Eur. Heart J. 2016, 37, 1789–1798. [CrossRef]
19. Gaetani, R.; Rizzitelli, G.; Chimenti, I.; Barile, L.; Forte, E.; Ionta, V.; Angelini, F.; Sluijter, J.P.; Barbetta, A.; Messina, E.; et al. Cardiospheres and tissue engineering for myocardial regeneration: Potential for clinical application. J. Cell Mol. Med. 2010, 14, 1071–1077. [CrossRef]
20. Lee, R. T., and Walsh, K. (2016). The future of cardiovascular regenerative medicine. Circulation 133, 2618–2625. doi: 10.1161/circulationaha.115.01 9214
21. Wang, H., Yang, Y., Liu, J., et al. (2021). Direct cell reprogramming: approaches, mechanisms and progress. Nat. Rev. Mol. Cell Biol. [Epub ahead of print]. doi: 10.1038/s41580-021-00335-z
22. Rincon, M. Y., VandenDriessche, T., and Chuah, M. K. (2015). Gene therapy for cardiovascular disease: advances in vector development, targeting, and delivery for clinical translation. Cardiovasc. Res. 108, 4–20. doi: 10.1093/cvr/cvv205
23. Kohama, Y., Higo, S., Masumura, Y., Shiba, M., Kondo, T., Ishizu, T., et al. (2020). Adeno-associated virus-mediated gene delivery promotes S-phase entry-independent precise targeted integration in cardiomyocytes. Sci. Rep. 10, 1–13.
24. Su, T., Huang, K., Ma, H., Liang, H., Dinh, P.-U., Chen, J., et al. (2018b). Plateletinspired nanocells for targeted heart repair after ischemia/reperfusion injury. Adv. Funct. Mater. 0, 1803567. doi: 10.1002/adfm.201803567
25. Mei, X., and Cheng, K. (2020). Recent development in therapeutic cardiac patches. Front. Cardiovasc. Med. 7:610364. doi: 10.3389/fcvm.2020.610364.
26. Maiullari, F., Costantini, M., Milan, M., Pace, V., Chirivì, M., Maiullari, S., et al. (2018). A multi-cellular 3D bioprinting approach for vascularized heart tissue engineering based on HUVECs and iPSC-derived cardiomyocytes. Sci. Rep. 8, 1–15
27. Jackman, C. P., Ganapathi, A. M., Asfour, H., Qian, Y., Allen, B. W., Li, Y., et al. (2018). Engineered cardiac tissue patch maintains structural and electrical properties after epicardial implantation. Biomaterials 159, 48–58. doi: 10.1016/ j.biomaterials.2018.01.002
28. Thomson, J.A.; Itskovitz-Eldor, J.; Shapiro, S.S.; Waknitz, M.A.; Swiergiel, J.J.; Marshall, V.S.; Jones, J.M. Embryonic stem cell lines derived from human blastocysts. Science 1998, 282, 1145–1147. [CrossRef]
29. Takahashi, K.; Tanabe, K.; Ohnuki, M.; Narita, M.; Ichisaka, T.; Tomoda, K.; Yamanaka, S. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. Cell 2007, 131, 861–872.
30. Teshigawara, R., Cho, J., Kameda, M., and Tada, T. (2017). Mechanism of human somatic reprogramming to iPS cell. Lab. Investig. 97, 1152–1157. doi: 10.1038/ labinvest.2017.56
31. Fernández-Avilés, F.; Sanz-Ruiz, R.; Climent, A.M.; Badimon, L.; Bolli, R.; Charron, D.; Fuster, V.; Janssens, S.; Kastrup, J.; Kim, H.-S. Global position paper on cardiovascular regenerative medicine. Eur. Heart J. 2017, 38, 2532–2546.
32. Pianezzi, E., Altomare, C., Bolis, S., Balbi, C., Torre, T., Rinaldi, A., et al. (2020). Role of somatic cell sources in the maturation degree of human induced pluripotent stem cell-derived cardiomyocytes. Biochim. Biophys. Acta Mol. Cell Res. 1867:118538. doi: 10.1016/j.bbamcr.2019.118538.
33. Tohyama, S., Hattori, F., Sano, M., Hishiki, T., Nagahata, Y., Matsuura, T., et al. (2013). Distinct metabolic flow enables large-scale purification of mouse and human pluripotent stem cell-derived cardiomyocytes. Cell Stem Cell 12, 127– 137. doi: 10.1016/j.stem.2012.09.013.
34. Wang, H., Yang, Y., Liu, J., et al. (2021). Direct cell reprogramming: approaches, mechanisms and progress. Nat. Rev. Mol. Cell Biol. [Epub ahead of print]. doi: 10.1038/s41580-021-00335-z
35. Hashimoto, H., Olson, E. N., and Bassel-Duby, R. (2018). Therapeutic approaches for cardiac regeneration and repair. Nat. Rev. Cardiol. 15, 585–600. doi: 10. 1038/s41569-018-0036-6
36. Wang, L., Liu, Z., Yin, C., Asfour, H., C
37. Barreto, S., Hamel, L., Schiatti, T., Yang, Y., and George, V. (2019). Cardiac progenitor cells from stem cells: learning from genetics and biomaterials. Cells 8:1536. doi: 10.3390/cells8121536hen, O., Li, Y., et al. (2015). Stoichiometry of Gata4, Mef2c, and Tbx5 influences the efficiency and quality of induced cardiac myocyte reprogramming. Circ. Res. 116, 237–244. doi: 10.1161/circresaha.116. 305547.
38. Liu, Z., Wang, L., Welch, J. D., Ma, H., Zhou, Y., Vaseghi, H. R., et al. (2017). Single-cell transcriptomics reconstructs fate conversion from fibroblast to cardiomyocyte. Nature 551, 100–104. doi: 10.1038/nature24454.
39. Aharonov, A., Shakked, A., Umansky, K. B., Savidor, A., Genzelinakh, A., Kain, D., et al. (2020). ERBB2 drives YAP activation and EMT-like processes during cardiac regeneration. Nat. Cell Biol. 22, 1346–1356. doi: 10.1038/s41556-020- 00588-4.
40. Shapiro, S. D., Ranjan, A. K., Kawase, Y., Cheng, R. K., Kara, R. J., Bhattacharya, R., et al. (2014). Cyclin A2 induces cardiac regeneration after myocardial infarction through cytokinesis of adult cardiomyocytes. Sci. Transl. Med. 6:224ra27. doi: 10.1126/scitranslmed.3007668.
41. Bulcha, J. T., Wang, Y., Ma, H., Tai, P. W. L., Gao, G. (2021). Viral vector platforms within the gene therapy landscape. Signal Transduct. Target Ther. 6:53.
42. Tenreiro, M.F.; Louro, A.F.; Alves, P.M.; Serra, M.J. Next generation of heart regenerative therapies: Progress and promise of cardiac tissue engineering. Npj Regen. Med. 2021, 6, 30.
43. Xu, F.; Dawson, C.; Lamb, M.; Mueller, E.; Stefanek, E.; Akbari, M.; Hoare, T. Hydrogels for Tissue Engineering: Addressing Key Design Needs Toward Clinical Translation. Front. Bioeng. Biotechnol. 2022, 10, 849831
44. Gao, L.; Gan, H.; Meng, Z.; Gu, R.; Wu, Z.; Zhang, L.; Zhu, X.; Sun, W.; Li, J.; Zheng, Y. Effects of genipin cross-linking of chitosan hydrogels on cellular adhesion and viability. Colloids Surf. B Biointerfaces 2014, 117, 398–405.
45. Murphy, S.V.; Atala, A. 3D bioprinting of tissues and organs. Nat. Biotechnol. 2014, 32, 773–785
46. Sorkio, A.; Koch, L.; Koivusalo, L.; Deiwick, A.; Miettinen, S.; Chichkov, B.; Skottman, H. Human stem cell based corneal tissue mimicking structures using laser-assisted 3D bioprinting and functional bioinks. Biomaterials 2018, 171, 57–71.
47. Lee, A.; Hudson, A.; Shiwarski, D.; Tashman, J.; Hinton, T.; Yerneni, S.; Bliley, J.; Campbell, P.; Feinberg, A. 3D bioprinting of collagen to rebuild components of the human heart. Science 2019, 365, 482–487.
48. Noor, N.; Shapira, A.; Edri, R.; Gal, I.; Wertheim, L.; Dvir, T. 3D printing of personalized thick and perfusable cardiac patches and hearts. Adv. Sci. 2019, 6, 1900344.
49. Xia, Y.; Whitesides, G.M. Soft lithography. Angew. Chem. Int. Ed. 1998, 37, 550–575
50. Feinberg, A.W.; Feigel, A.; Shevkoplyas, S.S.; Sheehy, S.; Whitesides, G.M.; Parker, K.K. Muscular thin films for building actuators and powering devices. Science 2007, 317, 1366–1370.
51. Crapo, P.M.; Gilbert, T.W.; Badylak, S.F. An overview of tissue and whole organ decellularization processes. Biomaterials 2011, 32, 3233–3243.
52. Fernández-Pérez, J.; Ahearne, M.J.S.R. The impact of decellularization methods on extracellular matrix derived hydrogels. Sci. Rep. 2019, 9, 14933.
53. Neishabouri, A.; Khaboushan, A.S.; Daghigh, F.; Kajbafzadeh, A.M.; Zolbin, M.M. Decellularization in Tissue Engineering and Regenerative Medicine: Evaluation, Modification, and Application Methods. Front. Bioeng. Biotechnol. 2022, 10, 805299.
54. Lu, T.-Y.; Lin, B.; Kim, J.; Sullivan, M.; Tobita, K.; Salama, G.; Yang, L. Repopulation of decellularized mouse heart with human induced pluripotent stem cell-derived cardiovascular progenitor cells. Nat. Commun. 2013, 4, 2307.
55. Tao, Z.-W.; Mohamed, M.; Hogan, M.; Salazar, B.; Patel, N.M.; Birla, R.K. Establishing the framework for fabrication of a bioartificial heart. ASAIO J. 2015, 61, 429–436.
56. Garry, M.G.; Kren, S.M.; Garry, D.J. Neonatal cardiac scaffolds: Novel matrices for regenerative studies. JoVE (J. Vis. Exp.) 2016, 5, e54459. [CrossRef] 88. Nguyen, D.T.; O’Hara, M.; Graneli, C.; Hicks, R.; Miliotis, T.; Nyström, A.-C.; Hansson, S.; Davidsson, P.; Gan, L.-M.; Magnone, M.C.J.S.r. Humanizing miniature hearts through 4-flow cannulation perfusion decellularization and recellularization. Sci. Rep. 2018, 8, 7458.
57. Barbulescu, G.I.; Bojin, F.M.; Ordodi, V.L.; Goje, I.D.; Buica, T.P.; Gavriliuc, O.I.; Baderca, F.; Hoinoiu, T.; Paunescu, V. Innovative biotechnology for generation of cardiac tissue. Appl. Sci. 2021, 11, 5603.
58. Park, S.M.; Yang, S.; Rye, S.-M.; Choi, S.W. Effect of pulsatile flow perfusion on decellularization. BioMedical Eng. OnLine 2018, 17, 15.
59. Guyette, J.P.; Charest, J.M.; Mills, R.W.; Jank, B.J.; Moser, P.T.; Gilpin, S.E.; Gershlak, J.R.; Okamoto, T.; Gonzalez, G.; Milan, D.J. Bioengineering human myocardium on native extracellular matrix. Circ. Res. 2016, 118, 56–72.
60. Hülsmann, J.; Aubin, H.; Sugimura, Y.; Lichtenberg, A.; Akhyari, P. Electrophysiological stimulation of whole heart constructs in an 8-pole electrical field. Artif. Organs 2018, 42, E391–E405.
61. Chan, G.; Mooney, D.J. New materials for tissue engineering: Towards greater control over the biological response. Trends Biotechnol. 2008, 26, 382–392.
62. Wang, B.; Borazjani, A.; Tahai, M.; de Jongh Curry, A.L.; Simionescu, D.T.; Guan, J.; To, F.; Elder, S.H.; Liao, J. Fabrication of cardiac patch with decellularized porcine myocardial scaffold and bone marrow mononuclear cells. J. Biomed. Mater. Res. Part A 2010, 94, 1100–1110.
63. Bassat, E.; Mutlak, Y.E.; Genzelinakh, A.; Shadrin, I.Y.; Baruch Umansky, K.; Yifa, O.; Kain, D.; Rajchman, D.; Leach, J.; Riabov Bassat, D. The extracellular matrix protein agrin promotes heart regeneration in mice. Nature 2017, 547, 179–184.
64. Chamberland, C.; Martinez-Fernandez, A.; Beraldi, R.; Nelson, T.J. Embryonic decellularized cardiac scaffold supports embryonic stem cell differentiation to produce beating cardiac tissue. Int. Sch. Res. Not. 2014, 2014, 625164.
65. Hong, X.; Yuan, Y.; Sun, X.; Zhou, M.; Guo, G.; Zhang, Q.; Hescheler, J.; Xi, J. Skeletal extracellular matrix supports cardiac differentiation of embryonic stem cells: A potential scaffold for engineered cardiac tissue. Cell. Physiol. Biochem. 2018, 45, 319–331.
66. Hochman-Mendez, C.; Pereira de Campos, D.B.; Pinto, R.S.; Mendes, B.J.d.S.; Rocha, G.M.; Monnerat, G.; Weissmuller, G.; Sampaio, L.C.; Carvalho, A.B.; Taylor, D.A. Tissue-engineered human embryonic stem cell-containing cardiac patches: Evaluating recellularization of decellularized matrix. J. Tissue Eng. 2020, 11, 2041731420921482.
67. Carvalho, J.L.; de Carvalho, P.H.; Gomes, D.A.; Goes, A.M. Characterization of decellularized heart matrices as biomaterials for regular and whole organ tissue engineering and initial in-vitro recellularization with ips cells. J. Tissue Sci. Eng. 2012, 11, 002.
68. Eitan, Y.; Sarig, U.; Dahan, N.; Machluf, M. Acellular cardiac extracellular matrix as a scaffold for tissue engineering: In vitro cell support, remodeling, and biocompatibility. Tissue Eng. Part C Methods 2010, 16, 671–683.
69. Wainwright, J.M.; Hashizume, R.; Fujimoto, K.L.; Remlinger, N.T.; Pesyna, C.; Wagner, W.R.; Tobita, K.; Gilbert, T.W.; Badylak, S.F. Right ventricular outflow tract repair with a cardiac biologic scaffold. Cells Tissues Organs 2012, 195, 159–170.
70. Wang, B.; Wang, G.; To, F.; Butler, J.R.; Claude, A.; McLaughlin, R.M.; Williams, L.N.; de Jongh Curry, A.L.; Liao, J. Myocardial scaffold-based cardiac tissue engineering: Application of coordinated mechanical and electrical stimulations. Langmuir 2013, 29, 11109–11117.