PHARMACOLOGICAL PROPERTIES OF NORTH INDIAN HERBS OF RUMEX

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ABSTRACT:

The genus Polygonaceae includes more than 200 species of annual, biennial and perennial herbaceous plants. Some species of this genus are found all over the world, some of them are cultivated. They are mainly distributed in the northern hemisphere and the countries of Europe, Asia, America and Africa. These species are traditionally used for culinary and medicinal purposes. The aerial parts, leaves and roots of plants in this genus are used as vegetables to treat various health problems such as mild diabetes, constipation, infections, diarrhea, jaundice. hepatobiliary disorders edema, and inflammation. Pharmacological activities have been studied in these species, including anti-inflammatory, antioxidant, antitumor, antibacterial, antiviral and antifungal properties. In their phytochemical studies, accumulations of anthraquinones, 1,8-naphthalenediol, flavonoids and stilbenes were found. R.Acetosa, R.acetosella, R.thyrsiflorus and R. vesicarius are known for their various ethnomedical uses. Although the genus contains more than 200 species, only more than 50 species have been studied and pharmacologically. phytochemically Moreover, the hepatoprotective, antiviral and antidiabetic activities of R. cerevisiae are a priority for future pharmacological studies. This review aims to describe the pharmacological and phytochemical properties of Rumex.

INTRODUCTION:

In Polygonaceae (buckwheat) family rumex is the second largest genus which contains more than 200 species [1]. Rumex, which refers to the form of the leaves, is a name that is derived from the Greek words "dart" and "spear" and is mostly distributed in the northern areas at an altitude of above 1000m [2]. This genus contains more perennial herbs along with strong roots, indeterminate inflorescence (racemose

inflorescence), and fruits that are covered with a large inner perianth. The eligible parts of leaves (used as food in the form of sour soup, sauces, and salads) flowers, and seeds are taken as vegetables and medicinal herbs also [3]. The flowers are small, green or red in color, branched clusters are present at the terminal and it grows in the month of April to May. The seeds become mature from the month of May to June [4]. Consuming large amounts of oxalic acid and hydroxy anthracene derivatives that are present in this genus rumex plant and make the food digest. Oxalic acid forms a calcium oxalate stone in the kidney and decreases iron absorption [5]. If the leaves are not cooked properly it can cause vomiting, nausea, stomach cramp, and headache [6]. Because of the resemblance in their morphology, the species belonging to this genus are very hard to differentiate between themselves [7]. It is a good source of primary nutritional compounds like vitamins, fat, sugar, protein, and minerals [8]. These plants are rich in biologically active secondary metabolites like polyphenols, anthraquinones, flavonoids, tannins, quinones, glycosides, tocopherols, carotenoids, terpenoids, glucosinolates, diterpene alkaloids, proanthocyanidins stilbenes and lignans [9]. It exhibits pharmacological action like antivirus, antitumor, antidiabetic, antiantibacterial. inflammatory, antiulcer. antifungal. anti-aging, phytotoxic, cytotoxic, dysenteric, anticoagulant, anti-diarrheal, antiemetic, anti-suppressive agents, anti-mutagenic, anti-hypertensive, hepatoprotective, gastrointestinal and cardiovascular anticancer. protective effects [10]. Some plants are used as natural colourants, paints, cosmetics, toys and textile industries [11].

BIOLOGICAL SOURCE:

Kingdom: Plantae Class : Magnoliopsida Phylum: Tracheophyta Order: Caryophyllales Family: Polygonaceae Subfamily: Polygonoideae Genus : Rumex [12]



Fig:1. Rumex Plant



Fig:1. Rumex Flower

GEOGRAPHICAL SOURCE:

Rumex species plays a crucial role in global folks medicine in Southern Africa, India, America, China and Turkey. It is also distributed in France, Italy, Kazakhstan, Newzealand, Norway, Germany, Finland, Europe, Australia, Denmark, United Kingdom, Russia, Ukraine, Switzerland and sweden [13].

MEDICINAL USES:

Rumex species were first used medicinally in China in "Shennong's Herbal Classic," which is used in the treatment of fever, gynecological illnesses, scabies, and head lice [14]. Rumex roots, commonly known as dock roots, are used to treat bacterial infections, jaundice, constipation, inflammation, tumors, and cardiovascular problems. Some of the rumex species are used as herbal medicine in the treatment of urinary inflammation, dysentery, skin burn, laxative, diuretic, chronic cutaneous diseases, astringent, osteomyelitis skin treatment, hepatitis sedative properties, gallstones, and liver diseases. Leaves and roots of Rumex maritimus are used as CNS depressants, Cathartic, Antipruritic, Purgative, and Antidiarrheal [15].

EXTRACTIONS:

- Leaves, roots, flowers, and seeds are collected
- washed and dried at room temperature or dark place
- Cut into small pieces then carefully loaded in a paper bag.
- 100g of rumex species are extracted in 70% hydro-methanol at room temperature 27°C.
- Filtered and evaporated at 55°C.

• Again, dried to obtain dried gum [16].

CHEMICAL CONSTITUENTS:

The acid plays a crucial role in Rumex species that mostly contain fatty acids like Linoleic acid, steric acids, benzoic acid, Ferulic acid and palmitic acids, which are separated from R. indurates. Likewise, Oxalic acid from R. abyssinicus and tartaric acid, citric acid from species of R. nervous. Antifungal activity has been detected in the presence of Chrysophanic acid are separated from R. abissinicus and R. nepalensis. Similarly, rumex species root contains ferulic acid and leave contain ascorbic acids act as antitumor and antioxidants agent [17]. More numbers of species are rich source of quinones, which are present in root including anthraquinones, anthraquinones, seco-anthraquinones. They are three significant anthraquinones, 1,5-dihydroxy-3-methyl anthraquinone, 1,3,5-trihydroxy-6-hydroxymethyl anthraquinone, and 1,5-dihydroxy-3-methoxy-7-methyl anthraquinone were isolated from the roots of R. crispus through chemical analysis of pharmaceutically important anthraquinones. These substances are used to treat constipation and purify the blood in cases of skin complaints [18]. Most of the flavonoids are obtained from kaempferol and quercetin associated with rhamnosyl, arabinosyl, glucosyl and galactosyl at different positions [19]. A chromone glucoside, 2,5-dimethyl-7-hydroxy chromone-7-O-\beta-D-glucoside were separated from the root of R. gmelini, and five chromones, 5,7-dihydroxy-1(3H)-chromone, 5methoxy-7-hydroxy-1(3H)-chromone2,5-dimethyl-7-hydroxy chromone-7-O-β-D-glucoside, 7-hydroxy-2,3-dimethyl-chromone and 2,5-dimethyl-7-hydroxy chromone were separated from R. gmelinite, R. nepalensis, R. patientia and R. cristatus [20]. The aerial segments of R. acetosa were extracted using acetone and water, and the EtOAc fraction

of that extract proved that R. acetosa was high in tannins. By using LC-

MS methods, the chemical constituents of Rumex species were also examined. Mostly on flowers and stems of R. tetanus, untargeted metabolomic profiling employing UHPLC-Q-TOF-MS analysis confirmed the presence of 60 compounds. The most common phenolic compound in flowers has been identified as quercetin-3-O-Dglucuronide and in stems epicatechin-3-O-gallate. The highest content of hydroxycinnamic acids are present in R. acetosella leaves (18 mg g-1), proanthocyanids from leaves of R. crispus, R. obtusifolius, and R. sanguineus (6.4 to 7.2 mg g-1) and catechins are found in R. sanguineus leaves (11 mg g-1). Isolation of rumex species produce six Resveratrol, 5.4'-dihydroxy-stilbene-3-O-αstilbenes that are (Z)-resveratrol, 5,4'-Dihydroxy-3-methoxystilbene, arabinoside. polydatin and 3,5-dihydroxy-4'-methoxystilbene. These stilbenes are used as antioxidants agent and cardiovascular protective. 6-hydroxy-11deoxy-13-dehydrohetisane, 7,11,14-trihydroxy-2,13-dioxohetisane and 6,13,15-trihydroxyhetisan are synthesized from tetra- or pentacyclic diterpenes that are present in the Rumex pictus [21]. The steroids stigmastane, α -cholestanol and β -cholestanol are separated from R.induratus, likewise, β -sitosterol-3-O- β -D-glycoside and β -cholestane are isolated from R. patientia [22].

PHARMACOLOGICAL ACTIVITY:

Rumex species are used as food and medicine. These species have more pharmacological and phytochemistry properties. The parts of plants are collected, washed, and dried, and extract from the chemical constituents. Through the chemical constitution pharmacological properties are identified that are used for treating disease and developing modern science [23].

ANTI-INFLAMMATORY ACTIVITY:

The high dose of R. patientia root (aqueous extract) 150 mg/kg reduced the edema with indomethacin the positive control at 10 mg/kg, 36.6%. Intestinal epithelial cells treated with methanolic extracts of R. roseus's roots and stem indicated anti-inflammatory properties, reducing TNF-induced gene expression of IL-6 and IL-8. R. crispus root ethyl acetate extract exhibited anti-inflammatory action by preventing Nitric oxide formation and reducing the release of proinflammatory cytokines [24].

ANTIOXIDANT ACTIVITY:

The antioxidant activity of the rumex sample was evaluated in its capacity to scavenge free radicals using the stable radical DPPH. Each Rumex sample was dissolved in 1 mL of methanol, mixed thoroughly, and then incubated for 15 minutes in the dark at room temperature (25°C) and varying concentration with the same volume of 0.3 mm DPPH in a solution of methanol. Using a UV-visible spectrophotometer to measure the absorption. antioxidant activity was found in R. acetosa by the root of 80% methanol extract showing IC50 = 118.8 μ M high scavenging activity to DPPH free radicals and for leaves IC50 = 201.6 μ M, flowers and fruits IC50 = 230.1 μ M, stems IC50 = 411.2 μ M [25].

ANTIMICROBIAL ACTIVITY:

The agar diffusion assay was done using the process given by Cappuccino and Sherman. The Whatman filter paper discs were sterilized and saturated with 100 l of the extract. Inoculation of young cultures on the surface agar plates. Petri dishes are incubated at 37°C for 24 hours. Experiments were performed in triplicate, and the diameter of the inhibition zone were calculated and compared with

DMSO. Antibiotics, penicillin, clotrimazole, and chloramphenicol were utilized. Isolate the root of R.japonicus obtained methoxy-stypandrone, naphthalenes and torachrysone indicate the inhibitory effect on both gram-positive and gram-negative bacteria. R.sanguineus and R.crispus plant have wound healing properties because they have anti-Acinetobacter baumannii activities [26].

ANTIVIRUS ACTIVITY:

R. aquaticus' 1,4-naphthoquinone and naphthalenes exhibited antiviral activity against Vero cells that were infected with HSV-2 (herpes simplex virus type 2) replication. On a conventional virus produce reduction test and qPCR assay, musizin (145) exhibited dose-dependent inhibitory property, resulting in a 2.00 log10 reduction in HSV-2 at a concentration of 6.25 M. It implied that R. aquaticus might be used to treat people with HSV-2 infections. R. acetosa exhibits antiviral activity through plaque reduction and MTT assay against vero cells [27].

ANTITUMOR ACTIVITY:

Inhibition of cancer cell proliferation by using MTT assays on MCF7 (human breast adenocarcinoma) cell lines, HeLa (human cervical carcinoma) and A431 (skin epidermoid carcinoma) exhibit the R. acetosa and R. thyrsiflorus properties. R. vesicarius shows greater cytotoxic in vitro with nontoxic on zebrafish development (IG50 = 33.45—62.56 µM) at concentration 30 µg/mL. When applied to zebrafish embryos, the chloroform extract of the stems prevented the growth of 100% of subintestinal vein blood vessels and around 70% of intersegmental blood vessels. This implies that the chloroform extract of R. vesicarius stems has evident anticancer potential [28].

ANTIDIABETIC ACTIVITY:

R. crispus root produces physcion and Chrysophanol and the alcohol extract of R. acetosella exhibits Inhibitory activity in α -glucosidase with IG₅₀ value, they show anti-diabetic properties. The seed of ethanolic extract of R. obtusifolius is administrated through the oral route, increasing glucose tolerance, decreasing the fasting glucose level, and increasing the glycogen level in the liver. It decreases the low-density lipoprotein cholesterol level, liver enzymes level, and total cholesterol level but it increases the high-density lipoprotein cholesterol level. This R. obtusifolius plant shows high potential to cure diabetes [29].

CONCLUSION:

The genus rumex is widely distributed in the world, it contains more than 200 species belongs to the family Polygonaceae. Rumex species are used in food, medicine and pharmaceutical industries because it is rich in nutrition, secondary metabolites and pharmacological activities. Leaves, roots, seeds, flowers, and some extracts are used to treat many various diseases like kidney disorder, urinary inflammation, jaundice, skin disease, diuretics, laxatives, regulations of digestive system, diabetes, constipation, chronic cutaneous, analgesic, cancer, gallstones. These species have greater potential for regrowth after wound. Some species contains oxalic acid it makes difficult to digest the food and create stone in kidney or digestive problems. The current studies are focused on the bioactive compounds, biosafety, more evaluation in chemical constituents and their efficiency for developing the modern science.

REFERENCES:

- Saleh NAM, Elhadidi N, Arafa RFM. Flavonoids and anthraquinones of some egyptian Rumex species (Polygonaceae) Biochem Syst Ecol. 1993;21:301–303. doi: 10.1016/0305-1978(93)90049-W.
- Liddell HG, Scott R, Jones HS, McKenzie R. A Greek-English lexicon. In: Fontaine D, editor. 9th edn. Oxford: Clarendon Press; 1940. p. 442.
- Wegiera, M., D, S. H. and D, W. (2007). Anthracene derivatives in some Rumex L. species. Acta. Socie. Botanicorum. Poloniae, 76: 103-108.
- 4. Munavum M R, Mudamba L O and Ogur J A. (1984). Isolation and Characterization of the major Anthraquinone Pigments from Rumexabysinica. PlantaMedica, 50: 111-111.
- 5. Siener R, Honow R, Seidler A, Voss S, Hesse A. Oxalate contents of species of the Polygonaceae, Amaranthaceae and Chenopodiaceae families. Food Chem. 2006;98:220–224.
- Tukappa, A.N.K.; Londonkar, R.L.; Nayaka, H.B.; Kumar S.C.B. Cytotoxicity and hepatoprotective attributes of methanolic extract of Rumex vesicarius L. Biological Research 2015, 48, 1-9.
- 7. Trease, G.E.; Evans W.C. Pharmacognosy (13th edn). Bailliere Tindall, London, 1989, 176-180.
- Litvinenko, Y.A.; MuzychKina, R. Phytochemical investigation of biologically active substances in certain Kazakhstan Rumex species. 1. Journal Chemistry of Natural Compounds 2003, 39, 446-449.
- Chelly M, Chelly S, Salah HB, Athmouni K, Bitto A, Sellami H, Kallel C, Allouche N, Gdoura R, Bouaziz-Ketata H. Characterization, antioxidant and protective effects of edible

Rumex roseus on erythrocyte oxidative damage induced by methomyl. J Food Meas Charact. 2020;14:229–243.

- Quradha MM, Khan R, Mujeeb-ur-Rehman, Abohajeb A. Chemical composition and in vitro anticancer, antimicrobial and antioxidant activities of essential oil and methanol extract from Rumex nervosus. Nat Prod Res. 2019;33:2554–2559.
- 11. Rogowska, A.; Szakiel, A. The role of sterols in plant response to abiotic stress. Phytochemistry Reviews 2020, 19, 1525-1538.
- 12. Jiang L, S Z. and L X. (2007). Oxanthrone C-glycosides and epoxynaphthoquinol from the roots of Rumex japonicus, Phytochemistry, 68: 2444-2449.
- Korpelainen H, Pietiläinen M. Sorrel (Rumex acetosa L.): not only a weed but a promising vegetable and medicinal plant. Bot Rev. 2020;86:234–246.
- Martin AC, Zim HS, Nelson AL. American wildlife and plants: a guide to wildlife food habits. New York: Dover Publications; 1951.
- 15. Khare CP. Indian medicinal plants. An illustrated Dictionary. New York: Springer-Verlag; 2007.
- 16. Harborne, J.B. Phenolic compounds, In Phytochemical methods, Springer, Dordrecht, 1973, 33-88,
- 17. Kerem Z, Bilkis I, Flaishman M. A. and Sivan, L. (2006). Antioxidant Activity and Inhibition of r-glucosidase by trans-Resveratrol, Piceid, and a Novel trans-Stilbene from the Roots of Israeli Rumexbucephalophorus L. Journal Of Agriculture and Food Chemistry,, 54: 1243-1247.
- Mei RQ, Liang HX, Wang J, Zeng LH, Lu Q, Cheng YX. New seco-anthraquinone glucosides from Rumex nepalensis. Planta Med. 2009;75:1162–1164.

- 19. Gao LM, Wei XM, Zheng SZ, Shen XW, Su YZ. Studies on chemical constituents of Rumex patientia. Chin Tradit Herbal Drugs. 2002;33:207–209.
- 20. Wang ZY, Cheng JR, Li RM, Wang ZQ. New chromone glucoside from roots of Rumex gmelini. Nat Prod Res Dev. 2009;21:189–191.
- 21. Bicker J, Petereit F, Hensel A. Proanthocyanidins and a phloroglucinol derivative from Rumex acetosa L. Fitoterapia. 2009;80:483–495.
- 22. Yuan Y, Chen W, Zhang S Q and Yang G J. (2001). Study on the structure and activity of new phenolic acid compounds from Erigeron breviscapus. Zhongguozhongyaozazhi, 26: 256-258.
- 23. Cappuccino J.G.; Sherman N. Microbiology: A laboratory Manual. Peason Benjamin Cummings. San Francisco, US, 2008.
- 24. Kim HY, Jeon H, Bae CH, Lee Y, Kim H, Kim S. Rumex japonicus Houtt. alleviates dextran sulfate sodium-induced colitis by protecting tight junctions in mice. Integr Med Res. 2020
- 25. Litvinenko YA, Muzychkina RA. New antioxidant phytopreparation from Rumex thyrsiflorus roots III. Chem Nat Compd. 2008;44:239–240.
- Nishina A, Kubota K, Osawa T. Antimicrobial components, trachrysone and 2-methoxystypandrone, in Rumex Japonicus Houtt. J Agr Food Chem. 1993;41:1772–1775.
- 27. Gescher K, Hensel A, Hafezi W, Derksen A, Kühn J. Oligomeric proanthocyanidins from Rumex acetosa L. inhibit the attachment of herpes simplex virus type-1. Antivir Res. 2011;89:9–18.
- 28. Tukappa NKA, Londonkar RL, Nayaka HB, Kumar CBS. Cytotoxicity and hepatoprotective attributes of methanolic extract of Rumex vesicarius L. Biol Res. 2015;48:19.
- 29. Aghajanyan A, Nikoyan A, Trchounian A. Biochemical activity and hypoglycemic effects of Rumex obtusifolius L. seeds used in

Armenian traditional medicine. Biomed Res Int. 2018 doi: 10.1155/2018/4526352.